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# **Research Article**

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# Extrafloral nectar as a driver of ant community spatial structure along disturbance and rainfall gradients in Brazilian dry forest

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# Abstract

Although extrafloral nectar (EFN) is a key food resource for arboreal ants, its role in structuring ground-nesting ant communities has received little attention, despite these ants also being frequent EFN-attendants. We investigated the role of EFN as a driver of the spatial structure of ground-nesting ant communities occurring in dry forest in north-eastern Brazil. We examined the effects on this relationship of two global drivers of biodiversity decline, chronic anthropogenic disturbance and climate change (through decreasing rainfall). We mapped EFN-producing plants and ant nests in 20 plots distributed along independent gradients of disturbance and rainfall. We categorized ant species into three types according to their dependence on EFN: heavy users, occasional users and non-users. We found a strong relationship between ant dependence on EFN and nest proximity to EFN-producing plants: heavy-users (mean distance 1.1 m) nested closer to EFN-producing plants than did non-users (2.3 m). Neither disturbance nor rainfall affected the proximity of heavy-user nests to EFN-producing plants. Our study shows for the first time that EFN is a key driver of the spatial structure of entire communities of ground-nesting ants.

# Introduction

Optimal foraging theory predicts that organisms seek to maximize net energetic yield in minimum foraging time (Grundel 1992, Oster & Wilson 1978, Stephens & Krebs 1986), and that this drives animal behaviour in terms of food selection and decisions about where, when and for how long foraging occurs (Pyke *et al.* 1977, Schoener 1971). According to optimal foraging theory (MacArthur & Pianka 1966), central-place foragers such as social insects optimize net energy gain by balancing trade-offs relating to colony location in a way that maximizes fitness (Covich 1976, Orians & Pearson 1979, Schoener 1979). Several studies have demonstrated that the foraging of social insects can be optimized by the location of nests close to key food resources (Grundel *et al.* 2010, Kacelnik *et al.* 1986, Murray 1938, Potts *et al.* 2003). However, there have been few empirical studies addressing how the distribution of food resources influences spatial patterns of social insect colonies.

Ants are ecologically dominant social insects in most terrestrial ecosystems, living in highly organized colonies where foragers retrieve food items to their nests, where they are stored, eaten or fed to offspring (Hölldobler & Wilson 1990, Traniello 1989). Plants provide many resources for ants as part of mutualistic interactions involving defence against herbivores (Bennett & Breed 1985, Bequaert 1922). Such resources can include nesting sites in the form of hollow thorns, stipules, leaf pouches, and chambers within epiphytic tubers (Janzen 1966, Rico-Gray & Oliveira 2007). However, most mutualistic interactions between plants and ants involve ant species that make their own nests, and therefore make their own decisions about where to locate their nests in relation to plant-based food resources such as leaves, nectar, seeds, honey-dew, or insects that live on vegetation (Holway & Case 2000, Kay 2002, Wagner & Fleur Nicklen 2010).

Extrafloral nectar (EFN) is a carbohydrate-rich food resource produced by at least 3941 plant species (Weber & Keeler 2013, Zhang *et al.* 2015) for attracting ants, which helps protect plants from herbivores (Heil 2011, Rico-Gray & Oliveira 2007). Many specialist nectar-feeding ants have specialized digestive systems designed to exploit liquid carbohydrates, allowing intensified exploitation of such resources, and thus generating high fidelity with their host plants (Byk & Del-Claro 2011, Davidson 1997). Many studies have shown that the availability of EFN strongly

influences arboreal ants, promoting ant foraging activity and diversity on trees (Blüthgen & Fiedler 2004, Davidson *et al.* 2003, Davidson 1997, Koptur 1992), and increasing colony survivorship, growth and reproduction (Byk & Del-Claro 2011). EFN is also exploited by ground-nesting ants, but its role in structuring ground-nesting ant communities has received little attention. Previous studies suggest that EFN production may encourage ants to build their nests near plants (Holway & Case 2000, Van Wilgenburg & Elgar 2007, Wagner & Fleur Nicklen 2010), but the community-wide influence of EFN on the spatial structure of ground-nesting ants remains to be examined.

In this study, we investigate the relationships between the distributions of EFN-producing plants and the nests of epigaeic ants occurring in caatinga dry forest of north-eastern Brazil. First, we hypothesize that the availability of EFN-producing plants plays an important role in structuring the spatial distribution of nests of ant species that are heavily dependent on nectar. We predict that the nests of such species are closer to trees producing EFN than are the nests of other ant species. Second, given that both disturbance and decreasing rainfall can negatively impact both populations of EFN-producing plants and nectar production (Heil 2011, Leal et al. 2015, Pacini et al. 2003), we hypothesize greater effects on the density and distribution of nests of heavily nectar-dependent ant species compared with other ants. Compared with other species, for ant species that are heavily dependent on nectar we predict a greater reduction in nest density with increasing CAD and decreasing rainfall. We also predict that the nests of ant species that are heavily dependent on nectar will be located even closer to EFNproducing plants with increasing CAD and decreasing rainfall, because nectar becomes an increasingly limited resource.

## **Materials and methods**

#### Study area

The study was carried out in Catimbau National Park ( $8^{\circ}24'00''-$ 8°36'35''S; 37°09'30''-37°14'40''W), in the state of Pernambuco, north-eastern Brazil, located in caatinga dry forest (Figure 1). Caatinga is the world's most species-rich dry forest (Pennington *et al.* 2009, Silva *et al.* 2017), and a large proportion of its woody plant species produce EFN (Leal *et al.* 2015, Melo *et al.* 2010). Caatinga is also one of the world's most highly populated and threatened semi-arid biomes, sustaining 28 million people who are highly dependent on forest natural resources for their livelihoods (Ribeiro *et al.* 2015, Silva *et al.* 2017) in a typical regime of chronic anthropogenic disturbance (Singh 1998). Moreover, caatinga is highly threatened by climate change, with climate models forecasting a 22% reduction in rainfall and a 3–6°C increase in temperatures by 2100 (Magrin *et al.* 2014).

The climate in Catimbau National Park is semi-arid, with annual temperature averaging 23°C, and mean annual rainfall varying from 480 to 1100 mm y<sup>-1</sup>, concentrated between March and July (Sociedade Nordestina de Ecologia 2002). Approximately 70% of its 607 km<sup>2</sup> is covered by quartzite sandy soils, but planosols and lithosols are also present (15% each one; Siqueira *et al.* 2017). The vegetation is composed of shrubs and small trees up to 7 m in height, dominated by the families Leguminosae, Euphorbiaceae, Boraginaceae and Burseraceae (Rito *et al.* 2017). Catimbau National Park is one of the most important conservation areas within the caatinga dry forest (Silva *et al.* 2004). However, the Park was only recently established (in 2002) and its inhabitants still remain; therefore, it continues to

be subject to a regime of chronic anthropogenic disturbance, including livestock grazing, timber harvesting, firewood collection and hunting (Rito *et al.* 2017, Siqueira *et al.* 2017, Sociedade Nordestina de Ecologia 2002). Catimbau includes a major gradient of decreasing rainfall from 1100 mm in the south-east to 480 mm in the north-west; such bioclimatic gradients provide powerful frameworks for addressing climate change impacts through space-for-time substitution (Blois *et al.* 2013; Caddy-Retalic *et al.* 2017).

Based on satellite imagery, soil maps, and data on rainfall and disturbance, we established 20 0.1-ha plots ( $50 \times 20$  m) covering a wide range of rainfall and disturbance intensity. All plots were on sandy soil, on flat terrain, and supported old-growth vegetation that had not experienced slash-and-burn agriculture for at least 50 y (Rito *et al.* 2017). Plots were separated by a minimum of 2 km and located within a total area of 21 430 ha (Rito *et al.* 2017).

#### Characterizing CAD and rainfall gradients

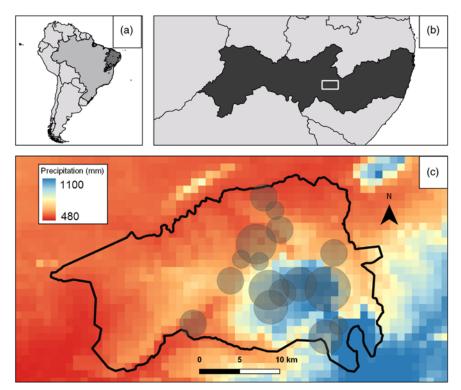
In order to assess the effects of CAD on the density and distribution of epigaeic ant nests, we used a global multi-metric disturbance index previously established for the study plots (Arnan et al. 2018). The index combines three types of source information: (1) indirect landscape measures using satellite imagery in ArcGIS 10.1 software (proximity to the nearest house, and proximity to the nearest road); (2) socio-economic information obtained by interviewing local households (number of people in the nearest village with influence weighted by distance); and (3) direct measures of disturbance in the field (length of goat trails, the density of goat and cattle dung, and extents of live-wood extraction and fire-wood collection). The values of each disturbance metric were first standardized between 0 and 1 to make component metrics of equal importance. We then computed the global multi-metric index ranged from 2 to 58. Full details on this disturbance index are provided in Arnan et al. (2018).

We obtained data on mean annual rainfall for each plot from the WorldClim global climate data repository (Hijmans *et al.* 2005) with 1-km resolution using the maptolls package for R v3.1.2. There are six meteorological stations around Catimbau National Park from which rainfall data have been interpolated to estimate values for our plots, which ranged from 510 to 940 mm. Such extreme variation in a small geographic area makes the study area ideal for analysing the ecological effects of rainfall on plant (Rito *et al.* 2017) and ant (Arnan *et al.* 2018, Leal *et al.* 2015) community attributes and interactions (Câmara *et al.* 2018, Oliveira *et al.* 2019).

#### Plant and ant surveys

We used the abundance of EFN-bearing plants as an indirect measure of overall availability of extrafloral nectar. To do so, we identified and counted all adult (> 1.5 m in height and > 3 cm diameter at soil level, following Rodal *et al.* 1992) EFN-producing woody plants in each plot between August 2014 and February 2015. Plant species were identified by comparing field-collected samples with samples from the Federal University of Pernambuco herbarium (UFP- Geraldo Mariz Herbaria), where voucher specimens of each species have been deposited. All EFN-producing plants were mapped according to a Cartesian coordinate system within each plot.

We surveyed ant nests in each plot between August 2014 and February 2015 by following foragers attracted to tuna baits returning to their nests. Thirty baits were used in each plot, placed



**Figure 1.** Location of study plots in Catimbau National Park (dark grey) in South America (a), location of Catimbau National Park (white rectangle) in Pernambuco state, Brazil (b), limits of Catimbau National Park (in black) with the 20 study plots represented by grey circles (increasing circle size represents higher levels of disturbance) (c).

on the soil for 2 h in a  $6 \times 5$  grid with 10-m spacing, located over the  $50 \times 20$ -m plots used to sample EFN-producing plants. We chose to use protein- rather than carbohydrate-based baits because these are attractive to a greater range of ant species, including both ants that are and are not heavily dependent on EFN (Andersen 1992). All ant nests were mapped according to the Cartesian coordinate system within each plot. All ants were sorted to morphospecies and identified to genus using Baccaro *et al.* (2015). Voucher specimens of all morphospecies were sent to the Systematics and Ant Biology Laboratory at the Universidade Federal do Parana for species identification. A complete set of voucher specimens is held in the ant collection at the Universidade Federal de Pernambuco. All applicable institutional and/or national guidelines for the care and use of animals were followed.

We categorized ant species into three types according to their dependence on EFN, based on published literature (Davidson 1997; Davidson *et al.* 2003) and our own expert knowledge: (1) heavy users: ants that are highly dependent on exudates and have special adaptations for a diet consisting primarily of liquid food; (2) occasional users: ants that feed on nectar opportunistically, and do not possess adaptations for it; and (3) non-users: ants that rarely or never feed on nectar.

# Data analysis

We constructed a General Linear Mixed Model (GLMM) in order to test whether the nests of ant species that are heavily dependent on EFN are closer to EFN-producing plants than are those of other species, where the distance of each ant nest to the nearest EFNproducing plant was the response variable, and ant feeding type was the fixed factor. Ant species nested within plot was added to the model as random factor. Second, we used Kolmogorov– Smirnov tests to compare the frequency distributions of the nearest distances between ant nests and EFN-producing plants among the three ant feeding types. To test our second prediction that increasing CAD and decreasing rainfall will have greater negative effects on nest density of species that are heavily dependent on nectar than in the other groups, we conducted General Linear Models (GLMs) where the density of nests of all ant species, heavy users, occasional users and non-users were the response variables, and CAD and rainfall were the fixed factors. Finally, to test our third prediction that nests of ant species that are heavily dependent on nectar will be located closer to EFN-producing plants with increasing CAD and decreasing rainfall, we ran a GLMM where the distance of each nest of heavy users to the nearest EFN-producing plant species was the response variable, CAD and rainfall were the fixed factors, and ant species nested within plots was the random factor. For the tests associated to our second and third predictions, we conducted a model comparison approach based on the Akaike's information criterion with a correction for finite sample sizes (AICc) to select the best-supported models; this approach reduces the problems associated with multiple testing, co-linearity of explanatory variables, and small sample sizes (Burnham & Anderson 2002). We compared four models, containing: both CAD and rainfall; only CAD; only rainfall; and only the intercept. The best-supported models were selected based on their AICc weights, which reveal the relative likelihood of a given model based on the data and the fit scaled to one; thus, models with a delta (AICc difference) of <2 were selected (Burnham & Anderson, 2002). The relevant variables were those that were retained in the best-supported models (except when the best-supported model consisted only of the intercept). Model selection was carried out using the dredge function in the MuMIn package in R.

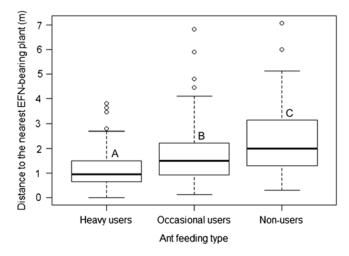
# Results

A total of 2243 individuals belonging to 12 genera and 21 species of EFN-producing plants were recorded, with a mean (±SE) density of  $1151 \pm 623$  plants ha<sup>-1</sup> and mean plot richness of  $5.1 \pm 0.5$ 

**Table 1.** List of ground-nesting ant species per subfamily showing feeding type (H = heavy user; O = occasional user; N = non-user) in relation to extrafloral nectar, total number of nests, and number of plots in which it occurred in Catimbau National Park, Pernambuco State, Brazil.

Subfamily     Ant     recting     No.     wor.       Dolichoderinae     Dorymyrmex goldii Forel, 1904     H     28     9       Dorymyrmex thoracicus     H     20     5       Gallardo, 1916     Dorymyrmex sp. A     H     15     5       Dorymyrmex sp. C     H     9     1       Dorymyrmex sp. D     H     7     1       Dorymyrmex sp. E     H     4     1       Dorymyrmex sp. B     H     4     1       Dorymyrmex sp. A     N     3     2       Ectatomminae     Ectatomma muticum Mayr, 1870     0     58     12       Formicinae     Brachymyrmex sp. A     O     4     4       Camponotus crassus Mayr, 1862     N     1     1       Myrmicinae     Cyphomyrmex transversus Emery, 1894     N     1     1       Pheidole radoszkowskii Mayr, 1884     0     18     10       Pheidole sp. B     N     9     4       Pheidole sp. K     N     1     1       Phe		Ant	Fooding	Ne	Ne
Image: Description of the image: Description of	Subfamily	Ant species	Feeding type	No. nests	No. plots
Gallardo, 1916       Dorymyrmex sp. A     H     15     5       Dorymyrmex sp. C     H     9     1       Dorymyrmex sp. D     H     7     1       Dorymyrmex sp. E     H     4     1       Dorymyrmex sp. H     H     6     1       Tapinoma sp. A     N     3     2       Ectatomminae     Ectatommuticum Mayr, 1870     O     58     12       Formicinae     Brachymyrmex sp. A     O     4     4       Camponotus crassus Mayr, 1862     H     7     3       Myrmicinae     Cyphomyrmex sp. A     N     1     1       Pheidole radoszkowskii Mayr, 1884     O     18     10       Pheidole sp. B     N     9     4       Pheidole sp. D     N     11     5       Pheidole sp. B     N     9     4       Pheidole sp. B     N     9     4       Pheidole sp. F     N     1     1       Pheidole sp. F     N     3     3	Dolichoderinae	Dorymyrmex goldii Forel, 1904	Н	28	9
Dorymyrmex sp. C     H     9     1       Dorymyrmex sp. D     H     7     1       Dorymyrmex sp. E     H     4     1       Dorymyrmex sp. H     H     6     1       Tapinoma sp. A     N     3     2       Ectatomminae     Ectatomma muticum Mayr, 1870     O     58     12       Formicinae     Brachymyrmex sp. A     O     4     4       Camponatus crassus Mayr, 1862     H     7     3       Myrmicinae     Cyphomyrmex transversus Emery, 1894     N     2     1       Kalathomyrmex sp. A     N     1     1     1       Pheidole radoszkowskii Mayr, 1884     O     18     10       Pheidole sp. B     N     9     4       Pheidole sp. E     N     7     3       Pheidole sp. F     N     1     1       Pheidole sp. F     N     3     3       Pheidole sp. F     N     3     3       Pheidole sp. F     N     3     3			Н	20	5
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Dorymyrmex sp. E     H     4     1       Dorymyrmex sp. H     H     6     1       Tapinoma sp. A     N     3     2       Ectatomminae     Ectatomma muticum Mayr, 1870     0     58     12       Formicinae     Brachymyrmex sp. A     0     4     4       Camponotus crassus Mayr, 1862     H     7     3       Myrmicinae     Cyphomyrmex transversus Emery, 1894     N     2     1       Pheidole radoszkowskii Mayr, 1884     0     18     10       Pheidole sp. B     N     9     4       Pheidole sp. B     N     1     1       Pheidole sp. B     N     1     1       Pheidole sp. B     N     1     1       Pheidole sp. F     N     3     3       Pheidole sp. R     N     1     1       Solenopsis sp. C <td></td> <td><i>Dorymyrmex</i> sp. C</td> <td>н</td> <td>9</td> <td>1</td>		<i>Dorymyrmex</i> sp. C	н	9	1
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Ectatomminae     Ectatomma muticum Mayr, 1870     O     58     12       Formicinae     Brachymyrmex sp. A     O     4     4       Camponotus crassus Mayr, 1862     H     7     3       Myrmicinae     Cyphomyrmex transversus Emery, 1894     N     2     1       Kalathomyrmex sp. A     N     1     1       Pheidole radoszkowskii Mayr, 1884     O     18     10       Pheidole sp. B     N     9     4       Pheidole sp. B     N     9     4       Pheidole sp. B     N     1     1       Pheidole sp. B     N     9     4       Pheidole sp. B     N     1     1       Pheidole sp. B     N     1     1       Pheidole sp. F     N     1     1       Pheidole sp. K     N     4     3       Pheidole sp. P     N     3     3       Solenopsis sp. C     N     3     3       Solenopsis sp. J     N     2     1       Solenopsis sp.		Dorymyrmex sp. H	н	6	1
IBTO     O     4     4       Camponotus crassus Mayr, 1862     O     4     4       Myrmicinae     Cyphomyrmex sp. A     N     2     1       Kalathomyrmex transversus Emery, 1894     N     2     1       Kalathomyrmex sp. A     N     1     1       Pheidole radoszkowskii Mayr, 1884     O     18     10       Pheidole sp. B     N     9     4       Pheidole sp. B     N     1     1       Pheidole sp. F     N     1     1       Pheidole sp. K     N     4     3       Pheidole sp. P     N     3     3       Solenopsis sp. B     N     1     1       Solenopsis sp. C     N     3     3       Solenopsis sp. N     N     1     1  S		Tapinoma sp. A	Ν	3	2
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Tetramorium sp. A N 1 1		Solenopsis virulens Smith, 1858	0	9	6
		Tetramorium sp. A	Ν	1	1

species. Most of the EFN-producing plant species (75%) and individuals (70%) belonged to Leguminosae. The most common species were *Pityrocarpa moniliformis* (Leguminosae; 27.8% of total individuals); *Poincianella microphylla* (Leguminosae; 19.6%) and *Croton argyrophylloides* (Euphorbiaceae; 18.8%). The density of EFN-



**Figure 2.** Boxplots showing the nearest distance between ant nests and EFN-bearing plants for the three ant feeding types according to EFN dependence (heavy-users, occasional users, and non-users) in Catimbau National Park, north-eastern Brazil. Different letters indicate significant differences (P < 0.05) according to post hoc contrasts.

bearing plants was not associated with variation in either CAD (GLM:  $F_{1,17} = 0.7$ ; P = 0.391) or rainfall ( $F_{1,17} = 0.5$ ; P = 0.482).

We recorded 257 ant nests belonging to 33 species (Table 1). The number of nests per plot varied from 5 to 24, corresponding to a density range of 50–240 nests ha<sup>-1</sup>. The most common ant species were *Ectatomma muticum* (Ectatomminae; 22.5% of all nests; occurring in 12 plots), *Dorymyrmex goldii* (Dolichoderinae; 10.8%; 9) and *D. thoracicus* (7.8%; 5). Heavy users comprised seven species of *Dorymyrmex*, which collectively represented 93% of all heavy-user nests, along with *Camponotus crassus*. Heavy users, occasional users and non-users accounted for 37.3%, 41.2% and 21.5% respectively of all ant nests. Ant nest density of any feeding category was not related to the density of EFN-producing plants.

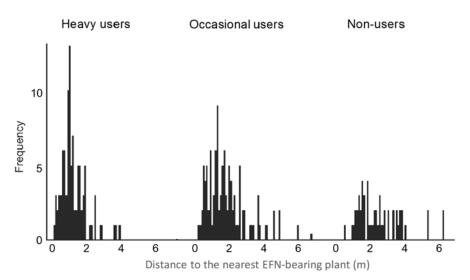
Mean nest distance from the nearest EFN-producing plant varied significantly among ant feeding types ( $F_{2,83} = 23.4$ ; P <0.001), with heavy-users (mean distance  $1.1 \pm 0.51$  m) nesting closer to EFN-producing plants than did occasional users (1.7 m;  $\pm 1.22$ ), which in turn nested closer to EFN-producing plants than did non-users (2.3 m;  $\pm 2.03$ ; Figure 2). Most nests of heavy users were located within 1 m of the nearest EFN-bearing plant, and none was located >4 m distant (D = 0.32; P <0.001). In contrast, most nests of other feeding types were located >1.5 and some >6 m distant (D = 0.50; P <0.001) (Figure 3).

The best-supported models always consisted only of the intercept, except for the density of heavy users, which included rainfall (Table 2). Thus, there was no relationship between CAD and the density of total ant nests or of those of any feeding type. The density of nests of heavy users declined with decreasing rainfall (Figure 4), but there was no relationship for either occasional users or non-users. Similarly, neither CAD nor rainfall were related to the proximity of heavy-user nests to EFN-producing plants (Table 2).

# Discussion

The availability of EFN is an important factor influencing arboreal ant communities, but its effects on ground-nesting ants remains largely unknown. Our study examined the relationship between the locations of EFN-producing plants and ant nests, and whether **Table 2.** Statistical outputs from the model comparison approach conducted to test the effects of chronic anthropogenic disturbance (CAD) and rainfall on the density of total nests, ant ground nests of heavy-users, occasional users and non-users and on the proximity of heavy users to EFN-bearing plants in Catimbau National Park, north-eastern Brazil. The best-supported models are highlighted by \*. Int., Intercept.

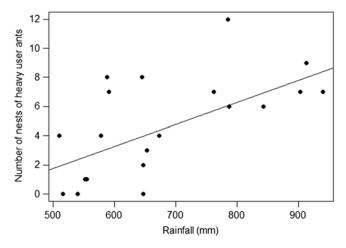
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Response variable	Model rank	Model variables	df	AICc	delta	weight
Density of total nests	1*	Int.*	2*	120.4*	0.00*	0.582*
	2	Int. + rainfall	3	122.5	2.12	0.202
	3	Int. + CAD	3	122.8	2.45	0.171
	4	Int. + CAD + rainfall	4	125.5	5.11	0.045
Density of heavy user nests	1*	$Int. + rainfall^*$	3*	102.6*	0.00*	0.797*
	2	Int. + CAD + rainfall	4	105.7	3.08	0.171
	3	Int.	2	109.7	7.09	0.023
	4	Int. + CAD	3	111.7	9.11	0.008
Density of occasional user nests	1*	Int.*	2*	118.8*	0.00*	0.637*
	2	Int. + rainfall	3	121.5	2.65	0.170
	3	Int. + CAD	3	121.6	2.79	0.158
	4	Int. + CAD + rainfall	4	124.6	5.79	0.035
Density of non-user nest	1*	Int. $+ rainfall^*$	3*	98.0*	0.00*	0.425*
	2	Int.	2	98.2	0.19	0.388
	3	Int. + CAD	3	101.0	2.92	0.099
	4	Int. + CAD + rainfall	4	101.2	3.14	0.088
Proximity of heavy users to plants	1*	Int.*	4*	74.5*	0.00*	0.992*
	2	Int. + CAD	5	84.4	9.98	0.007
	3	Int. + Rainfall	5	87.8	13.38	0.001
	4	Int. + CAD + rainfall	6	98.7	24.22	0.000



**Figure 3.** Distribution of the nearest distances between ant nests and EFN-producing plants for the three ant feeding types according to EFN dependence (heavy-users, occasional users and non-users) in Catimbau National Park, north-eastern Brazil.

this varies along gradients of CAD and rainfall, both of which are known to influence EFN production. For the first time, we have shown that EFN-bearing plants are a key driver of the spatial structure of entire ground-nesting ant communities. This occurred across a wide range of disturbance and climatic conditions.

Species of *Dorymyrmex* and *Camponotus* were heavily dependent on EFN (heavy users) in our study system. They belong to subfamilies (Dolichoderinae and Formicinae respectively) that are considered to be specialized exudate feeders (Eisner 1957), possessing anatomical traits that allow them to retain and process large volumes of plant exudates (Davidson & Patrell-Kim 1996, Davidson *et al.* 2003, Eisner 1957). Species of *Camponotus* have a sclerotized proventriculus that can store large volumes of fluids for long periods of time (Eisner 1957). As in other dolichoderine genera such as *Iridomyrmex*, *Froggattella* and *Turneria*, species of *Dorymyrmex* possess a complex proventriculus that is capable of carrying more liquid than is required by an individual forager (Cook & Davidson 2006, Eisner 1957).



**Figure 4.** Relationship between mean annual rainfall and the density of nests of EFN heavy-user ants in the 20 study sites at Catimbau National Park, north-east Brazil. The line depicts the linear fit between the two variables.

Nesting in close proximity to a key food resource reduces travel times, which both increases foraging efficiency and reduces exposure to natural enemies (Davidson 1997, McIver 1991, Pfeiffer & Linsenmair 1998). As we predicted, heavy-user ants established their nests closer to EFN-bearing plants than did other ants. Mean nest distance to the nearest EFN-producing plant was 1.2 m for heavy users, compared with about 2 m for other ant species. Most nests of heavy users were located within 1 m of an EFN-producing plant, whereas most nests of other species occurred more than 1.5 m away. We also found a more-nuanced effect of ant feeding type: although nests of occasional users were more distant than those of heavy users, they were closer than those of non-users. Our study therefore provides empirical evidence that EFN drives the nest distributions of ant species according to the extent to which they use nectar resources.

A relationship between nest distribution and nectar resources has been previously suggested for individual ground-nesting ant species. For example, Bennett & Breed (1985) found an association between *Pentaclethra macroloba* (Leguminosae) trees and nests of the giant tropical ant *Paraponera clavata* in Costa Rica, likely related to the provision of EFN. Similarly, there is an association mediated by EFN between *Acacia constricta* and *Dorymyrmex* and *Forelius* species in the Sonoran Desert (Wagner & Fleur Nicklen 2010). However, our study is the first to demonstrate a community-wide impact of EFN on the spatial structure of epigaeic ants.

Many heavy-user ant species are behaviourally dominant, relying on large volumes of liquid carbohydrate for powering their rapid locomotory activity and high levels of aggression (Blüthgen & Fiedler 2004, Davidson et al. 2003). In our study, this applies to species of Dorymyrmex (Dominant Dolichoderinae sensu Andersen 1995), which represented >90% of all heavy-user nests. These dominant species can exercise competitive control in the vicinity of their nests (Hölldobler & Wilson 1990), and this is likely to be a factor contributing to other species nesting more distantly from EFN-producing plants. However, given that the density of Dorymyrmex nests (mean of 41.5 ha<sup>-1</sup>) was only a small fraction of that of EFN-producing plants (1151 plants ha<sup>-1</sup>) such competitive exclusion could not be the dominant factor driving the spatial structure of occasional-user and non-user ant species. This is supported by non-significant correlations between the distance to the nearest EFN-bearing plant of Dorymyrmex species nests and the distance to the nearest EFN-bearing plant of occasional users (Spearman rho = -0.18; P = 0.450) and non-users (Spearman rho = -0.28; P = 0.235).

Our study also examined the effects of two global drivers of biodiversity decline, CAD and climate change, on nests of groundforaging ants and their spatial relationships with EFN-producing plants. We predicted that increasing anthropogenic disturbance and decreasing rainfall would have a greater impact on nest densities of ants that are heavily dependent on nectar than on other species, because disturbance and decreasing rainfall can negatively impact both populations of EFN-producing plants and nectar production (Heil 2011, Leal et al. 2015, Nichol & Hall 1988, Whitford et al. 1995). Nest density did not vary with CAD, but our prediction held for rainfall, where a decline in rainfall was associated with a decrease in the nest density of heavy-users, but not of other ants. The decline in nest density of heavy-users was not due to a decline in the density of EFN-bearing plants (which did not vary with either rainfall or CAD); it can therefore be attributed to a decline in nectar production rates, which is known to occur under conditions of low water availability (Jakobsen & Kristjánsson 1994, Keasar et al. 2008, Murcia 1995) due to stomatal closure induced by water stress (Heil 2011, Lange et al. 2013, Rico-Gray et al. 1998). This is supported by the finding that the size of EFN glands of P. moniliformis decreases with decreasing rainfall (Reis 2016). The decline in nest density of heavy-user ants with increasing aridity suggests that EFN-mediated ant protection services for plants also declines with aridity. This has implications for plant protection services under future climates, which are projected to be substantially drier in the region.

Our final prediction was that the nests of ant species that are heavily dependent on nectar will be located even closer to EFNproducing plants with increasing disturbance and decreasing rainfall, because nectar becomes an increasingly limited resource (Heil 2011, Leal et al. 2015, Pacini et al. 2003). The density of heavy-user nests did not vary with CAD, which suggests that nectar production likewise did not vary; in such circumstances our finding that proximity of heavy-user nests to EFN-producing plants did not vary with CAD is to be expected. However, nectar production did appear to decline with decreasing rainfall, but this did not result in heavy-user ants nesting even closer to EFN-bearing plants. One possible explanation is that colony size declines with increasing aridity due to factors unrelated to EFN production, such that even if nectar production declines in an absolute sense, it is not relatively more limiting. Alternatively, rate of nectar production might just not be a relevant factor in the optimization of nest proximity to EFN-bearing plants in our study system.

In conclusion, we have shown that EFN is not just important for arboreal ant communities, but that it is a key driver of the spatial structure of ground-nesting ant communities, especially through the supply of a key food resource for behaviourally dominant ants. This has a direct effect on the location of the nests of these ants in relation to EFN-producing plants, and likely also a competition-mediated indirect effect on the spatial structure of other ant species. These effects occur across a wide range of disturbance and climatic conditions.

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# Literature cited

- Andersen AN (1992) Regulation of "momentary" diversity by dominant species in exceptionally rich ant communities of the Australian seasonal tropics. *American Naturalist* 140, 401–420.
- Andersen AN (1995) A classification of Australian ant communities, based on functional groups which parallel plant life-forms in relation to stress and disturbance. *Journal of Biogeography* 22, 15–29.
- Arnan X, Leal IR, Tabarelli M, Andrade JF, Barros MF, Câmara, T, Jamelli D, Knoechelmann C, Menezes TGC, Menezes AGS, Oliveira FMP, de Paula AS, Pereira SG, Rito KF, Sfair JC, Siqueira FFS, Souza DG, Specht MJ, Vieira LA, Arcoverde GB and Andersen A (2018) A framework for deriving measures of chronic anthropogenic disturbance: surrogate, direct, single and multi-metric indices in Brazilian Caatinga. *Ecological Indicators* 94, 274–282.
- Baccaro FB, Feitosa RM, Fernández F, Fernandes IO, Izzo TJ, Souza JLP and Solar R (2015) Guia para os gêneros de formigas do Brasil. Editora INPA, Manaus.
- Bennett B and Breed MD (1985) The nesting biology, mating behavior, and foraging ecology of *Perdita opuntiae* (Hymenoptera: Andrenidae). *Journal of the Kansas Entomological Society* **58**, 185–194.
- Bequaert JC (1922) Ants in their diverse relations to the plant world. Bulletin of the American Museum of Natural History 45, 333–621.
- Blois JL, Williams JW, Fitzpatrick MC, Jackson ST and Ferrier S (2013) Space can substitute for time in predicting climate-change effects on biodiversity. *Proceedings of the National Academy of Sciences USA* 110, 374–9379.
- Blüthgen N and Fiedler K (2004) Competition for composition: lessons from nectar feeding ant communities. *Ecology* 85, 1479–1485.
- Burnham KP and Anderson DR (2002) Model Selection and Multimodel Inference: A Practical Information-theoretic Approach. New York, NY: Springer-Verlag.
- Byk J and Del-Claro K (2011) Ant-plant interaction in the neotropical savanna: direct beneficial effects of extrafloral nectar on ant colony fitness. *Population Ecology* 53, 327–332.
- Caddy-Retalic S, Andersen AN, Aspinwall MJ, Breed MF, Byrne M, Christmas MJ, Dong N, Evans BJ, Fordham DA, Guerin GR, Hoffmann AA, Hughes AC, van Leeuwen SJ, McInerney FA, Prober SM, Rossetto M, Rymer PD, Steane DA, Wardle GM and Lowe AJ (2017) Bioclimatic transect networks: powerful observatories of ecological change. Ecology and Evolution 7, 4607–4619.
- Câmara T, Leal, IR, Blüthgen N, Oliveira FMP, De Queiroz RT and Arnan X (2018) Effects of chronic anthropogenic disturbance and rainfall on the specialization of ant-plant mutualistic networks in the Caatinga, a Brazilian dry forest. *Journal of Animal Ecology* 87, 1022–1033.
- Cook SC and Davidson DW (2006) Nutritional and functional biology of exudate-feeding ants. Entomologia Experimentalis et Applicata 118, 1–10.
- Covich AP (1976) Analyzing shapes of foraging areas: some ecological and economic theories. Annual Review of Ecology, Evolution, and Systematics 7, 235–257.
- **Davidson DW** (1997) The role of resource imbalances in the evolutionary ecology of tropical arboreal ants. *Biological Journal of the Linnean Society* **61**, 153–181.
- Davidson DW and Patrell-Kim L (1996) Tropical arboreal ants: why so abundant? In Gibson AC (ed.), *Neotropical Biodiversity and Conservation*. Berkeley, CA: University of California, pp. 127–140.
- Davidson DW, Cook SC and Snelling RR (2003) Explaining the abundance of ants in lowland tropical rainforest canopies. *Science* **300**, 969–972.
- **Eisner T** (1957) A comparative morphological study of the proventriculus of ants (Hymenoptera: Formicidae). *Bulletin of the Museum of Comparative Zoology* **116**, 429–490.

- **Grundel R** (1992) How the mountain chickadee procures more food in less time for its nestlings. *Behavioral Ecology and Sociobiology* **31**, 291–300.
- Grundel R, Jean RP, Frohnapple KJ, Glowacki GA, Scott PE and Noel NB (2010) Floral and nesting resources, habitat structure, and fire influence bee distribution across an open-forest gradient. *Ecological Applications* **20**, 1678–1692.
- Heil M (2011) Nectar: generation, regulation and ecological functions. *Trends in Plant Science* 16, 191–200.
- Hijmans RJ, Cameron SE, Parra JL, Jones PG and Jarvis A (2005) Very high resolution interpolated climate surfaces for global land areas. *International Journal of Climatology* 25, 1965–1978.
- Hölldobler B and Wilson EO (1990) *The Ants*. Cambridge, MA: Belknap Press, Harvard University Press.
- Holway DA and Case TJ (2000) Mechanisms of dispersed central-place foraging in polydomous colonies of the Argentine ant. *Animal Behaviour* **59**, 433–441.
- Jakobsen H and Kristjánsson K (1994) Influence of temperature and floret age on nectar secretion in *Trifolium repens*. L. Annals of Botany 74, 327–334.
- Janzen DH (1966) Coevolution of mutualism between ants and acacias in Central America. *Evolution* 20, 249–275.
- Kacelnik A, Houston AI and Schmid-Hempel P (1986) Central-place foraging in honey bees: the effect of travel time and nectar flow on crop filling. *Behavioral Ecology and Sociobiology* **19**, 19–24.
- Kay A (2002) Applying optimal foraging theory to assess nutrient availability ratios for ants. *Ecology* 83, 1935–1944.
- Keasar T, Sadeh A and Shmida A (2008) Variability in nectar production and standing crop, and their relation to pollinator visits in a Mediterranean shrub. Arthropod–Plant Interactions 2, 117–123.
- Koptur S (1992) Extrafloral nectary-mediated interactions between insects and plants. In E. Bernays (ed.), *Insect-Plant Interactions*. Boca Raton, FL: CRC Press, pp. 81–129.
- Lange D, Dáttilo W and Del-Claro K (2013) Influence of extrafloral nectary phenology on ant-plant mutualistic networks in a neotropical savanna. *Ecological Entomology* 38, 463–469.
- Leal LC, Andersen AN and Leal IR (2015) Disturbance winners or losers? Plants bearing extrafloral nectaries in Brazilian Caatinga. *Biotropica* 47, 468–474.
- MacArthur RH and Pianka ER (1966) On optimal use of a patchy environment. American Naturalist 100, 603–609.
- Magrin GO, Marengo JA, Boulanger JP, Buckeridge MS, Castellanos E, Poveda G and Vicuña S (2014) Central and South America. In Barros VR et al. (eds), Climate Change 2014: Impacts, Adaptation, and Vulnerability.
  Part B: Regional Aspects. Contribution of Working Group II to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge: Cambridge University Press, pp. 1499–1566.
- McIver JD (1991) Dispersed central place foraging in Australian meat ants. Insectes Sociaux 38, 129–137.
- Melo Y, Machado SR and Alves M (2010) Anatomy of extrafloral nectaries in Fabaceae from dry-seasonal forest in Brazil. *Botanical Journal of the Linnean Society* 163, 87–98.
- Murcia C (1995) Edge effects in fragmented forests: implications for conservation. Trends in Ecology and Evolution 10, 58–62.
- Murray JM (1938) An investigation of the interrelationships of the vegetation, soils and termites. *South African Journal of Science* **35**, 288–297.
- Nichol P and Hall L (1988) Characteristics of nectar secretion by extrafloral nectaries of *Ricinus communis*. Journal of Experimental Botany 39, 573–586.
- Oliveira FMP, Andersen NA, Arnan X, Ribeiro-Neto JD, Arcoverde GB and Leal IR (2019) Effects of increasing aridity and chronic anthropogenic disturbance on seed dispersal by ants in Brazilian Caatinga. *Journal of Animal Ecology*, in press.
- Orians GH and Pearson NE (1979) On the theory of central place foraging. In Horn DJ, Mitchell RD and Stairs GR (eds), *Analysis of Ecological Systems*. Columbus, OH: Ohio State University Press, pp. 155–177.
- Oster GF and Wilson EO (1978) Caste and Ecology in the Social Insects. Princeton, NJ: Princeton University Press

- Pacini E, Nepi M and Vesprini JL (2003) Nectar biodiversity: a short review. Plant Systematic Evolution 238, 7–21.
- Pennington RT, Lavin M and Oliveira-Filho A (2009) Woody plant diversity, evolution, and ecology in the tropics: perspectives from seasonally dry tropical forests. *Annual Review of Ecology, Evolution, and Systematics* 40, 437–457.
- Pfeiffer M and Linsenmair KE (1998) Polydomy and the organization of foraging in a colony of the Malaysian giant ant *Camponotus gigas* (Hym./ Form.). Oecologia 117, 579–590.
- Potts SG, Vulliamy B, Dafni A, Ne'eman G, O'Toole C, Roberts S and Willmer P (2003) Response of plant-pollinator communities to fire: changes in diversity, abundance and floral reward structure. *Oikos* **101**, 103–112.
- **Pyke GH, Pulliam HR and Charnov EL** (1977) Optimal foraging: a selective review of theory and tests. *Quarterly Review of Biology* **52**, 137–154.
- **Reis DQA** (2016) Influência de perturbações antrópicas crônicas e mudanças climáticas na diversidade estrutural de plantas com nectários extraflorais em uma floresta seca. Master's Thesis; Universidade Federal de Pernambuco, Recife.
- Ribeiro EMS, Arroyo-Rodríguez V, Santos BA, Tabarelli M and Leal IR (2015) Chronic anthropogenic disturbance drives the biological impoverishment of the Brazilian Caatinga vegetation. *Journal of Applied Ecology* 52, 611–620.
- Rico-Gray V and Oliveira PS (2007) The Ecology and Evolution of Ant-Plant Interaction. Chicago, IL: University of Chicago Press.
- Rico-Gray V, Garcia-Franco JG, Palacios-Rios M, Iz-Castelazo C, Parra-Tabla V and Navarro JA (1998) Geographical and seasonal variation in the richness of ant-plant interactions in Mexico. *Biotropica* 30, 190–200.
- **Rito KF, Arroyo-Rodríguez V, Queiroz RT, Leal IR and Tabarelli M** (2017) Precipitation mediates the effect of human disturbance on the Brazilian Caatinga vegetation. *Journal of Ecology* **105**, 828–838.
- Rodal MJN, Sampaio EVSB, Figueiro MA and Figueiredo MA (1992) Manual sobre métodos de estudo florístico e fitossociológico. Brasília, DF: SBB.
- Schoener TW (1971) Theory of feeding strategies. Annual Review of Ecology, Evolution, and Systematics 2, 369–404.

- Schoener TW (1979) Generality of the size-distance relation in models of optimal feeding. American Naturalist 114, 902–914.
- Silva JD, Tabarelli M, Fonseca MD and Lins LV (2004) Biodiversidade da Caatinga: áreas e ações prioritárias para a conservação. Brasília, DF: Universidade Federal de Pernambuco.
- Silva JMC, Leal IR and Tabarelli M (2017) Caatinga: The Largest Tropical Dry Forest Region in South America. Cham: Springer.
- Singh SP (1998) Chronic disturbance: a principal cause of environmental degradation in developing countries. *Environmental Conservation* 25, 1–2.
- Siqueira FFS, Ribeiro-Neto JD, Tabarelli M, Andersen AN and Wirth R, Leal IR (2017) Leaf-cutting ant populations profit from human disturbances in tropical dry forest in Brazil. *Journal of Tropical Ecology* **33**, 337–344.
- Sociedade Nordestina de Ecologia (2002) Projeto técnico para a criação do Parque Nacional do Catimbau/PE. Recife.
- Stephens DW and Krebs JR (1986) Foraging Theory. Princeton, NJ: Princeton University Press.
- Traniello JFA (1989) Foraging strategies of ants. Annual Review of Entomology 34, 191–210.
- Van Wilgenburg E and Elgar MA (2007) Colony characteristics influence the risk of nest predation of a polydomous ant by a monotreme. *Biological Journal of the Linnean Society* **92**, 1–8.
- Wagner D and Fleur Nicklen E (2010) Ant nest location, soil nutrients and nutrient uptake by ant-associated plants: does extrafloral nectar attract ant nests and thereby enhance plant nutrition? *Journal of Ecology* 98, 614–624.
- Weber MG and Keeler KH (2013) The phylogenetic distribution of extrafloral nectaries in plants. Annals of Botany 111, 1251–1261.
- Whitford WG, Martinez-Turanzas G and Martinez-Meza E (1995) Persistence of decertified ecosystems: explanations and implications. *Environmental Monitoring and Assessment* **37**, 319–332.
- Zhang S, Zhang Y and Keming MA (2015) The equal effectiveness of different defensive strategies. *Scientific Reports* 5, Art. no. 13049.