# Corrieopone nouragues gen. nov., sp. nov., a new Ponerinae from French Guiana (Hymenoptera, Formicidae) 

Flavia A. Esteves', Brian L. Fisher ${ }^{\prime}$<br>I Entomology, California Academy of Sciences, 55 Music Concourse Drive, San Francisco, CA 94118, USA<br>Corresponding author: Flavia A. Esteves (flaviaesteves@gmail.com)

Academiceditor:MarekBorowiec|Received21September2021|Accepted13November2021|Published3December2021
http://zoobank.org/OEE73BBC-E98A-46AC-B6B9-83BF15868F8B
Citation: Esteves FA, Fisher BL (2021) Corrieopone nouragues gen. nov., sp. nov., a new Ponerinae from French Guiana (Hymenoptera, Formicidae). ZooKeys 1074: 83-173. https://doi.org/10.3897/zookeys.1074.75551


#### Abstract

This study describes the worker and queen castes of the Neotropical ponerine Corrieopone nouragues gen. nov., sp. nov., an ant from the tropical rainforest in French Guiana. Worker morphology of the taxon is compared with those of other Ponerinae and the similarities between them are discussed, refining the definition of character states for some diagnostic characters at the generic level, providing an identification key to the Neotropical genera, and making some adjustments to the taxonomic framework within the subfamily. Descriptions, diagnosis, character discussion, identification key, and glossary are illustrated with more than 300 images and line drawings. Open science is supported by providing access to measurement data for specimens of the new genus, a matrix of character states for all ponerine taxa evaluated in this study, and specimen data for all examined material. The new or revived combinations presented here are Pachycondyla procidua Emery, comb. rev., Neoponera curiosa (Mackay and Mackay), comb. nov., Leptogenys butteli (Forel), comb. nov., and Bothroponera escherichi (Forel), comb. nov. In addition, Leptogenys butteli is synonymized with Leptogenys myops (Emery), syn. nov.


## Keywords

Ant Course, ants, myrmecology, neotropical, new genus, ponerines, South America

## Introduction

Since 2001, 17 Ant Course editions have trained nearly 500 students from 59 countries, included more than 60 international instructors, and offered opportunities to explore the biological diversity in different parts of the globe, from Australia to Southeast Asia, to East Africa, and to North, Central, and South America (www.antcourse.org). Specimens collected in past editions enhanced our understanding of several aspects of ant biology, such as functional morphology (e.g., Peeters et al. 2017; Larabee et al. 2017, 2018; Gibson et al. 2018), ecology (Kronauer 2004), reproductive biology (Peeters 2017), and natural history (LaPolla et al. 2004; Lattke and Delsinne 2016). The course also created opportunities for remarkable discoveries. For example, two new genera records for USA fauna, Typhlomyrmex Mayr (CASENT0173317) and Fulakora Mann (MCZ-ENT00528501); the first record of the worker caste of the Afrotropical genus Aenictogiton Emery (CASENT0906052); and two genera recorded for the first time in Borneo (Rhopalothrix Mayr and Tyrannomyrmex Fernández; Fisher et al. 2015).

Here we describe a novel ponerine genus and species discovered during the 2018 Ant Course in the Natural Reserve of Nouragues, French Guiana. Ponerinae Lepeletier de Saint-Fargeau (see diagnosis in Fisher and Bolton 2016: 53-54) currently consists of 47 extant genera and 1,263 valid species (AntCat.org). Of these, 17 genera and 673 species are present in the Neotropics (AntWeb.org). Schmidt and Shattuck (2014) recently revised the higher classification of the subfamily and split the former genus Pachycondyla into 19 genera. The authors also described Iroponera odax, a name coined by WL Brown Jr (Cornell University, Ithaca, USA), who unfortunately passed away before describing that unknown genus and species. Iroponera Schmidt \& Shattuck is the most recent ponerine genus description based on a new species rather than reclassification.

As part of the process of diagnosing the new genus, we make a few adjustments to the taxonomic framework within Ponerinae, provide a new identification key for workers of the Neotropical genera, and reassess morphological characters across the subfamily.

## Materials and methods

## Study site

Ant Course 2018 was held in August at Nouragues Research Station (4.08796N, $52.68002^{\circ} \mathrm{W}$ ) in the Natural Reserve of Nouragues in French Guiana (Figs 1, 2). A French overseas territory on the northeastern edge of South America, French Guiana sits on a Precambrian massif known as the Guiana Shield (Grimaldi and Riéra 2001). The Nouragues station's hilly landscape features a granitic inselberg that rises to 430 m (Fig. 2C, E; Grimaldi and Riéra 2001). The station lies in the Guianan Lowland Moist Forests ecoregion, a subset of the Tropical and Subtropical Moist Forests biome (Dinerstein et al. 2017); the climate is equatorial humid, and seasonality is determined by the movements of the Inter-tropical Convergence Zone (Grimaldi and Riéra 2001).


Figure I. Shaded relief maps of the type locality of Corrieopone nouragues. Clockwise from top-left: South America, French Guiana, and Natural Reserve of Nouragues. Symbols: red circle, collection locality; circle with white fill and black outline, Cayenne, the capital of French Guiana.


Figure 2. Images of the Nouragues Research Station A ant Course 2018 students working in the lab at the Inselberg camp B view of the Inselberg camp kitchen facilities $\mathbf{C}$ forested landscape of the reserve viewed from the Nouragues Inselberg $\mathbf{D}$ rainforest along the Approuague river $\mathbf{E}$ the Nouragues Inselberg (top left of the image) and surrounding landscape $\mathbf{F}$ collection site of Corrieopone nouragues. Photography by BL Fisher.

The region experiences an average annual temperature of $-26^{\circ} \mathrm{C}$ and yearly average precipitation $\sim 2990 \mathrm{~mm}$, with the rainy season occurring from November to August but interrupted in March by a short dry period (Grimaldi and Riéra 2001).

## Material examined and specimen records

During this study, we contrasted the morphology of the new genus with 129 species or subspecies representing all 47 currently valid extant Ponerinae genera (AntCat 2021; see Table 1 for taxa list, Suppl. material 1: Table S1 for specimen data). Suppl. mate-
rial 3: Table S3 (character matrix) documents characters evaluated that could be unambiguously discretized, and we share it here to disclose our methodology and foster validation, replication, and reinterpretation of our results (see also Suppl. material 2: Table S2 for character statements).

Specimens were examined with a Leica M125 microscope with $187.5 \times$ total magnification power (Leica Microsystems, Switzerland). Minute characters were accessed with scanning electron microscopy (SEM) or through SEM images previously available on AntWeb (www.antweb.org). Each specimen evaluated bears a registered unique identifier (e.g., CASENT0830464) associated with collection and specimen information, images, and identification on AntWeb. Data is accessible on AntWeb through the persistent URL of a given unique identifier (e.g., www.antweb.org/specimen/CASENT0830464).

Maxillary and labial palpal counts are based on direct observation of specimens with protracted maxillolabial complex $(N=41)$, dissections ( $N=47$ ), and existing SEM images of the maxillolabial complex on AntWeb.org $(N=26)$. For dissected specimens, the maxillolabial complex was removed from the buccal cavity with a pin or forceps. Larger specimens were observed with a Leica M125 microscope; smaller specimens were accessed with SEM. See Suppl. material 3: Table S3 (column Notes_char_9) for specimen data on palps evaluated, including unique specimen identifiers and observation method.

In subsequent sections, we refer to several other species whose morphology was only assessed through extended-focus images of specimens databased on AntWeb. Those species were not included in Table 1 or Tables S1 and S3. Instead, and when necessary, references are accompanied by the unique identifier of relevant specimens, which are hyperlinked to respective webpages on AntWeb.

## Images, drawings, and maps

Extended focus montage images were acquired with a Leica DFC 425 camera and LEICA APPLICATION SUITE software (version 3.8; Leica Microsystems, Switzerland). For most SEM images, samples were coated with gold-palladium in a Cressington 108 Sputter Coater (Cressington Scientific Instruments, United Kingdom), and micrographs were taken at high vacuum secondary electron emission (accelerating voltage: 15 kV , spot intensity diameter: 40) in a Hitachi SU3500 microscope (Hitachi High-Technologies, Japan). Uncoated specimens (e.g., primary types, unique specimens) were also imaged at high vacuum secondary electron emission, but with the accelerating voltage set to 1.5 kV and spot intensity to 50 . Image enhancement (e.g., contrast, levels, sharpness, darken background) occurred in ADOBE PHOTOSHOP (version 22.4.2; Adobe Inc., United States of America). All images produced in this study are available on AntWeb.

Line drawings were originally traced in ADOBE ILLUSTRATOR (version 25.2.3; Adobe Inc., United States of America) or modified from artwork produced by scientific illustrator Jessica Huppi, whose authorship is credited in figure captions when pertinent. Jessica Huppi line art is part of a work-for-hire agreement that makes the California Academy of Sciences a copyright holder for her artwork.

Table I. List of taxa used for direct comparisons with the morphology of the new genus. In addition, it contains information on how specimens were determined.

| Taxon name and Author(s)year | Determination methodology |
| :---: | :---: |
| Anochetus angolensis Brown | Type examined |
| Anochetus emarginatus (Fabricius) | Det. CA Schmidt; compared with type images |
| Asphinctopone differens Bolton \& Fisher | Type examined |
| Asphinctopone silvestrii Santschi | Det. BL Fisher |
| Austroponera castanea (Mayr) | Det. WL Brown |
| Belonopelta deletrix Mann | Det. SP Cover; FA Esteves; BL Fisher |
| Boloponera ikemkha Hawkes | Type examined |
| Boloponera vicans Fisher | Type examined |
| Bothroponera cariosa Emery | Det. BL Fisher |
| Bothroponera crassa (Emery) | Det. G Fischer; BL Fisher |
| Bothroponera pachyderma (Emery) | Det. BL Fisher; compared with type images |
| Bothroponera silvestrii (Santschi) | Det. G Fischer; compared with type images |
| Bothroponera talpa André | Det. BL Fisher |
| Brachyponera chinensis (Emery) | Det. WP Mackay; compared with type images |
| Brachyponera croceicornis (Emery) | Type examined |
| Brachyponera lutea (Mayr) | Det. BL Fisher; compared with type images |
| Brachyponera luteipes (Mayr) | Det. WL Brown; compared with type images |
| Brachyponera obscurans (Walker) | Det. BL Fisher |
| Brachyponera sennaarensis (Mayr) | Det. FA Esteves; compared with type images |
| Buniapone amblyops (Emery) | Det. FA Esteves; compared with type images |
| Centromyrmex brachycola (Roger) | Det. ES Ross; compared with type images |
| Centromyrmex decessor Bolton \& Fisher | Type examined |
| Centromyrmex ereptor Bolton \& Fisher | Type examined |
| Centromyrmex raptor Bolton \& Fisher | Type examined |
| Cryptopone gilva (Roger) | Det. BL Fisher; J Lattke; compared with type images |
| Cryptopone guianensis (Weber) | Det. SP Cover; PS Ward |
| Cryptopone hartwigi Arnold | Det. P Hawkes |
| Diacamma ceylonense Emery | Det. C Peeters |
| Dinoponera longipes Emery | Det. PA Lenhart |
| Dinoponera lucida Emery | Det. JRM Santos |
| Dolioponera fustigera Brown | Det. BL Fisher; compared with type images |
| Ectomomyrmex javanus Mayr | Det. WL Brown |
| Emeryopone buttelreepeni Forel | Det. D Agosti; FA Esteves |
| Euponera brunoi (Forel) | Det. BL Fisher; compared with type images |
| Euponera sikorae (Forel) | Det. BL Fisher |
| Euponera sjostedti (Mayr) | Det. BL Fisher; RR Snelling |
| Feroponera ferox Bolton \& Fisher | Type examined |
| Fisheropone ambigua (Weber) | Det. FA Esteves; BL Fisher |
| Hagensia havilandi marleyi (Arnold) | Det. C Peeters; compared with type images |
| Harpegnathos saltator Jerdon | Det. C Peeters |
| Hypoponera punctatissima (Roger) | Det. BL Fisher |
| Iroponera odax Schmidt \& Shattuck | Det. WL Brown |
| Leptogenys ixta Lattke | Type examined |
| Leptogenys peruana Lattke | Type examined |
| Leptogenys podenzanai (Emery) | Det. RA Keller (?); compared with type images |
| Leptogenys pucuna Lattke | Type examined |
| Leptogenys sonora Lattke | Type examined |
| Leptogenys wheeleri Forel | Det. FA Esteves; compared with type images |
| Loboponera obeliscata Bolton \& Brown | Det. taxon author(s) |
| Loboponera vigilans Bolton \& Brown | Det. taxon author(s) |
| Mayaponera arhuaca (Forel) | Det. WL Brown; compared with type images |
| Mayaponera becculata (Mackay \& Mackay) | Type examined |
| Mayaponera cernua (Mackay \& Mackay) | Type examined |
| Mayaponera conicula (Mackay \& Mackay) | Type examined |


| Taxon name and Author(s)year | Determination methodology |
| :---: | :---: |
| Mayaponera constricta (Mayr) | Det. JT Longino |
| Mayaponera pergandei (Forel) | Det. WP Mackay |
| Megaponera analis (Latreille) | Det. FA Esteves |
| Mesoponera ambigua (André) | Det. G Fischer; BL Fisher |
| Mesoponera australis (Forel) | Det. FA Esteves |
| Mesoponera caffraria (Smith) | Det. BL Fisher |
| Mesoponera elisae rotundata (Emery) | Det. G Fischer |
| Mesoponera melanaria macra (Emery) | Det. FA Esteves |
| Mesoponera papuana (Viehmeyer) | Type examined |
| Mesoponera rubra (Smith) | Det. FA Esteves; compared with type images |
| Mesoponera subiridescens (Wheeler) | Det. G Fischer; BL Fisher |
| Myopias darioi Probst \& Boudinot | Type examined |
| Myopias maligna (Smith) | Det. FA Esteves; compared with type images |
| Neoponera aenescens (Mayr) | Det. WL Brown; compared with type images |
| Neoponera apicalis (Latreille) | Det. WP Mackay; compared with type images |
| Neoponera bugabensis (Forel) | Det. WP Mackay; compared with type images |
| Neoponera carinulata (Roger) | Det. WP Mackay; compared with type images |
| Neoponera cavinodis Mann | Det. WL Brown |
| Neoponera commutata (Roger) | Det. WL Brown |
| Neoponera crenata (Roger) | Det. WP Mackay; compared with type images |
| Neoponera dismarginata (Mackay \& Mackay) | Det. taxon author(s) |
| Neoponera eleonorae (Forel) | Det. WL Brown |
| Neoponera cf. fiebrigi | Det. BL Fisher |
| Neoponera fisheri (Mackay \& Mackay) | Type examined |
| Neoponera foetida (Linnaeus) | Det. WP Mackay |
| Neoponera globularia (Mackay \& Mackay) | Type examined |
| Neoponera insignis (Mackay \& Mackay) | Det. BL Fisher |
| Neoponera inversa (Smith) | Det. WP Mackay; compared with type images |
| Neoponera laevigata (Smith) | Det. WP Mackay; compared with type images |
| Neoponera luteola (Roger) | Det. WP Mackay; compared with type images |
| Neoponera moesta (Mayr) | Det. WP Mackay; compared with type images |
| Neoponera obscuricornis (Emery) | Det. WP Mackay; compared with type images |
| Neoponera schoedli (Mackay \& Mackay) | Type examined |
| Neoponera striatinodis (Emery) | Det. WP Mackay |
| Neoponera unidentata (Mayr) | Det. BL Fisher; compared with type images |
| Neoponera verenae (Forel) | Det. WP Mackay; compared with type images |
| Neoponera villosa (Fabricius) | Det. LR Davis; compared with type images |
| Odontomachus bauri Emery | Det. FA Esteves; RA Keller |
| Odontoponera transversa (Smith) | Det. FA Esteves; RA Keller |
| Ophthalmopone berthoudi Forel | Det. FA Esteves; compared with type images |
| Pachycondyla crassinoda (Latreille) | Det FA Esteves, Kempf (1964) ID key |
| Pachycondyla harpax (Fabricius) | Det FA Esteves, Kempf (1964) ID key; compared with type images |
| Pachycondyla impressa (Roger) | Det FA Esteves, Kempf (1964) ID key; compared with type images |
| Pachycondyla lattkei Mackay \& Mackay | Type examined |
| Pachycondyla lenis Kempf | Det FA Esteves, Kempf (1964) ID key |
| Pachycondyla procidua Emery | Det FA Esteves, Kempf (1964) ID key; compared with type images |
| Pachycondyla striata Smith | Det. WP Mackay; compared with type images |
| Paltothyreus tarsatus (Fabricius) | Det. G Alpert; G Fischer |
| Parvaponera darwinii madecassa (Emery) | Det. BL Fisher |
| Phrynoponera pulchella Bolton \& Fisher | Type examined |
| Phrynoponera transversa Bolton \& Fisher | Type examined |
| Platythyrea cribrinodis (Gerstäcker) | Det. FA Esteves, Brown (1975) ID key; compared with type images |
| Platythyrea punctata (Smith) | Det. LR Davis; RA Keller; compared with type images |
| Platythyrea turneri Forel | Det. RA Keller (?); CA Schmidt; PS Ward |
| Plectroctena strigosa Emery | Det. C Peeters; HJ Robertson |
| Ponera alpha Taylor | Det. taxon author(s) |
| Ponera pennsylvanica Buckley | Det. RA Keller; J Lattke |


| Taxon name and Author(s)year | Determination methodology |
| :--- | :---: |
| Promyopias silvestrii (Santschi) | Det. B Bolton |
| Psalidomyrmex procerus Emery | Det. WL Brown; A Dejean; BL Fisher |
| Pseudoneoponera porcata (Emery) | Det. RA Keller |
| Pseudoneoponera tridentata (Smith) | Det. BL Fisher |
| Pseudoponera gilberti (Kempf) | Det. FA Esteves, Mackay and Mackay (2010) ID key; compared with Kempf |
|  | $(1960)$ description |
| Pseudoponera stigma (Fabricius) | Type examined |
| Rasopone costaricensis Longino \& Branstetter | Type examined |
| Rasopone cryptergates Longino \& Branstetter | Type examined |
| Rasopone cubitalis Longino \& Branstetter | Type examined |
| Rasopone guatemalensis Longino \& Branstetter | Type examined |
| Rasopone panamensis (Forel) | Type examined |
| Rasopone pluviselva Longino \& Branstetter | Type examined |
| Rasopone politognatha Longino \& Branstetter | Type examined |
| Simopelta oculata Gotwald \& Brown | Type examined |
| Simopelta transversa Mackay \& Mackay | Type examined |
| Streblognathus peetersi Robertson | Det. C Peeters |
| Thaumatomyrmex fraxini D'Esquivel \& Jahyny | Det. taxon author(s) |
| Thaumatomyrmex zeteki Smith | Det. FA Esteves, Kempf (1975) ID key |

Mapping of the study area occurred on RStudio Desktop (version 1.4.1717; RStudio Team 2021), an integrated development environment for R (version 4.1.0; R Core Team 2021). Spatial data was sourced directly from the following: NASADEM HGT v001 products (NASA JPL 2020), Global Lakes and Wetlands Database (GLWD2 dataset; Lehner and Döll 2004), Shuttle Radar Topography Mission Water Body models (NASA JPL 2013), Global River Classification dataset (GloRiC version 1.0, Ouellet Dallaire et al. 2018), and Global Tree Cover dataset (Hansen et al. 2013). Alternatively, data were acquired and imported into R with functions of the following R packages: OSMDATA (Padgham et al. 2017), RASTER (Hijmans 2021), RWORLDMAP (South 2011), and RWORLDXTRA (South 2012); the OSMDATA package imports features from OpenStreetMap (OpenStreetMap.org data were available under the Open Database License). Data geoprocessing was executed with functions of DPLYR (Wickham et al. 2021), GDALUTILS (Greenberg and Mattiuzzi 2020), RASTER, and SF (Pebesma 2018) packages. Map projection and mapping were performed with functions available in the TMAP package (Tennekes 2018).

## Terminology

Positional and directional terminology references a hypothetical worker seen in profile, with the head oriented to the left and standing with legs slightly spread over the horizontal plane of a multidimensional space (Fig. 3). Mouthparts are retracted, and antennal scapes are directed posteriad, parallel to the head dorsum. We defined three main directional axes from this orientation, which guided the description of direction and relative position of morphological characters in this study. The anteroposterior axis extends horizontally across the space, from the anterior left side to the posterior right; the dorsoventral axis extends vertically, with "dorsal" directed upwards and "ventral" directed downwards. Finally, the


Figure 3. Anatomical position and coordinate system adopted as framework for positional and directional terminology. Corrieopone nouragues is shown in profile, with the head oriented to the left. The coordinate system is defined by the median plane and three directional axes (anteroposterior, dorsoventral, and mediolateral). Illustration by FA Esteves.
mediolateral axis is perpendicular to a median plane that bisects the ant through its bilateral line, with "medial" directed towards the plane and "lateral" diverging from it (Fig. 3).

Additionally, we utilized a fourth axis for aiding the description of positions and directions of characters located on appendages, sclerites, processes, or any structure that may project away from the body. The basoapical axis arises from the median plane, with "basal" being close to the plane while "apical" is distant from it (Fig. 4A). Morphological terminology is based on Keller (2011) unless otherwise stated. Wherever possible, terms in the glossary are hyperlinked to correspondent terms in the Hymenoptera Anatomy Ontology portal (Yoder et al. 2010). Terms absent from the glossary below are defined, illustrated, or associated with references to pertinent literature at first use.

Anepisternum (an, Fig. 5B): Dorsal subdivision of the mesopleuron, separated from the katepisternum by the mesepisternal sulcus (= anapleural sulcus in Fisher and Bolton 2016).

Antenna (pl antennae; ant, Fig. 6A): Paired head appendage that, from base to apex, is composed of scape, pedicel, and flagellum. We refer to pedicel and flagellomeres as antennomeres; Roman numerals in ascending order indicate the position of individual antennomeres along the antenna's basoapical axis (e.g., scape $=$ antennomere I, pedicel $=$ antennomere II, basalmost flagellomere $=$ antennomere III).

Arolium (pl arolia; ar, Fig. 4B): The adhesive pretarsal organ; a soft cuticular structure between the pretarsal claws.

Bulbus neck: The constricted portion of the antennal scape, bordered basally by the bulbus (= condylar bulbus in Fisher and Bolton 2016) and apically by the main shaft of the scape.


Figure 4. Glossary of terminology, part I A hypothetical hindleg in a three-dimensional coordinate system defined by the anteroposterior, dorsoventral, and basoapical directional axes $\mathbf{B}$ apicalmost tarsomere and pretarsus in ventroapical view, with the retracted arolium highlighted in orange $\mathbf{C}$ probasitarsus and protibial apex in posterior view, with the comb of strigil highlighted in orange. Abbreviations: ar, arolium; bt, basitarsus; ca, calcar of strigil; cx, coxa; cw, claw; fm, femur; ma, manubrium; sgc, comb of strigil; tb, tibia; t1-5, tarsi; tch, trochanter. Illustrations by FA Esteves.

Calcar of strigil (ca, Fig. 4C): The protibial spur; together with the comb of strigil, forms the antennal cleaning organ.

Comb of strigil (sgc, Fig. 4C): Comb-like structure on the ventral face of the basal portion of the probasitarsus; together with the calcar of strigil, forms the antennal cleaning organ.

Epistomal sulcus (es, Fig. 6A, B; as in Richards 1977): Sulcus delimitating the clypeus posteriorly and laterally from the remainder of the head.

Galea (ga, Fig. 6E): The spatulate lobe located at the apical part of the stipes; part of the maxilla.

Galeal comb: Row of setae located on the outer face of the medial margin of the galea, opposite the maxillary comb located near the inner face of the margin.

Galeal crown: The apicalmost part of the galea.


Figure 5. Glossary of terminology, part II A body in profile, with the abdomen highlighted in orange hatching B mesosoma and petiole in profile, with the mesopleuron highlighted in orange hatching; laterotergite in solid orange $\mathbf{C}$ gaster in profile, with the hypopygium highlighted in orange $\mathbf{D}$ ventral face of the petiole; laterotergite highlighted in orange $\mathbf{E}$ dorsal face of the gaster. Abbreviations: A1-7, abdominal segments; an, anepisternum; cx1-3, coxae; he, helcium; hy, hypopygium; kn, katepisternum; lt, laterotergite; mn, mesonotum; mtp, metapleuron; mts, metanotal sulcus; nps, notopleural suture; pn, pronotum; ppd, propodeum; $\mathbf{p p z}$, petiolar proprioceptor zone; $\mathbf{p r}$, prora; $\mathbf{p s t}$, petiolar sternite; $\mathbf{p s t g}$, posttergite; $\mathbf{p t}$, petiole; $\mathbf{p t g}$, pretergite; py, pygidium; sp, subpetiolar process; spl, spiracular lobe; str, stridulitrum. Illustrations by FA Esteves.

Gaster (Fig. 5A, C, E; as in Fisher and Bolton 2016): Tagma formed by the third, fourth, fifth, sixth, and seventh abdominal segments in female ponerines. When mentioning individual segments, we refer to the homologous abdominal segment labeled


Figure 6. Glossary of terminology, part III A head in full-face view $\mathbf{B}$ head in lateral view $\mathbf{C}$ mandibles and the anterior area of the clypeus in dorsal view $\mathbf{D}$ labrum and retracted maxillolabial complex in ventral view $\mathbf{E}$ maxillolabial complex in central view; labial palps highlighted in green, maxillary palps in orange. Abbreviations: ant, antenna; bm, basal margin of the mandible; cl, clypeus; es, epistomal sulcus; ga, galea; gl, glossa; $\mathbf{l b r}$, labrum; $\mathbf{l m}$, lateral margin of the mandible; $\mathbf{m d}$, mandible; $\mathbf{m m}$, masticatory margin of the mandible; plb, labial palps; pm, premental shield; pmx, maxillary palps; sc, scape; st, stipes; tol, torular lobe; trm, torular median arch. Illustrations by FA Esteves ( $\mathbf{A}, \mathbf{B}, \mathbf{C}, \mathbf{E}$ ), and modified from artwork by Jessica Huppi (D).
by a corresponding Roman numeral (e.g., abdominal segment V ). For aesthetic reasons in figures and measurements abbreviations, segments are noted by "A" followed by the corresponding Arabic numeral (e.g., A5).

Helcium (he, Fig. 5C): Structure formed by the specialized presclerites of abdominal segment III, which articulates with the petiole.

Hypopygium (hy, Fig. 5C): The sternite of abdominal segment VII in adult female ants.

Katepisternum (kn, Fig. 5B): Ventral subdivision of the mesopleuron, separated from the anepisternum by the mesepisternal sulcus (= anapleural sulcus in Fisher and Bolton 2016). Note that this usage does not claim homology between the katepisternum in ants and homonymous mesothoracic areas in other insects.

Labrum (lbr, Fig. 6D): Mouthpart appendage that connects to the anterior margin, or the ventral face of the clypeus, and usually folds over the retracted maxillolabial complex.

Labial palps (plb, Fig. 6E): Labial ring-shaped sclerites articulated with the apicolateral portion of the prementum. We refer to each sclerite as a palpomere and use Roman numerals in ascending order to indicate the position of individual palpomeres along the basoapical axis of the palps (e.g., basalmost labial palpomere $=$ labial palpomere I).

Lower and upper metapleuron (as in Snodgrass 1910): Secondary division of the metapleuron into dorsal wing-bearing and ventral leg-bearing portions.

Maxillary palps (pmx, Fig. 6E): Ring-shaped sclerites articulated with the apicomedial or the apicalmost portion of the stipes in the maxilla. We refer to each sclerite as a palpomere and use Roman numerals in ascending order to indicate the position of individual palpomeres along the basoapical axis of the palps (e.g., basalmost maxillary palpomere $=$ maxillary palpomere I).

Mesoscutellar-axillar complex (as in Gibson et al. 1998): In winged ants, the area of the mesonotum that comprises the mesoscutellum and axillae; located posteriad to the transscutal line.

Mesoscutum (as in Gibson et al. 1998): In winged ants, the region of the mesonotum anterior to the transscutal line and mesoscutellar-axillar complex, whose laterals usually bear parapsidal lines.

Mesometapleural suture (as in Fisher and Bolton 2016): Groove-like suture delimiting the mesopleuron from the metapleuron.

Mesonotum (Fig. 5A): The tergum of the mesothorax.
Mesopleuron (Fig. 5B; as in Fisher and Bolton 2016): Pleuron of the mesothorax; extends over the lateral and most of the ventral external surfaces of the second thoracic segment.

Mesosoma (Fig. 5A): Tagma formed by the three thoracic segments (pro-, meso-, and metathorax) plus the propodeum (abdominal segment I).

Mesosternal process (as in Fisher and Bolton 2016): The pair of cuticular projections surrounding the midline of the mesothorax's ventral face, anterior to the mesocoxal cavities.

Metasternal process (as in Fisher and Bolton 2016): The pair of cuticular projections surrounding the midline of the metathorax's ventral face, anterior to the metacoxal cavities.

Metanotum (mts, Fig. 5B): The tergum of the metathorax; reduced to a groove (Fig. 5B) or completely indistinct in Ponerinae workers.

Metapleuron (mtp, Fig. 5B; as in Fisher and Bolton 2016): Pleuron of the metathorax; extends over the lateral and ventral external surfaces of the third thoracic segment; bears the metapleural gland opening laterally.

Notopleural suture (nps, Fig. 5B): Suture between mesopleuron and mesonotum.
Petiolar laterotergite (lt, Fig. 5B, D): Paired, long, narrow, strip-like area of the ventral margin of the petiolar tergite that is separated from the main part of the sclerite by an impression; flanks the petiolar sternite.

Petiolar proprioceptor zone (ppz, Fig. 5D): A depression on the anteriormost part of the petiolar sternite, sharply delineated anteriorly, and bearing numerous sensilla.

Petiole (pt, Fig. 5A, B, D): Abdominal segment II.
Postsclerite: Posterior portion of each abdominal sclerite not concealed by an articulation. The term posttergite refers to the postsclerite of a tergum (pstg, Fig. 5E), and poststernite to the postsclerite of a sternum (Fig. 5D).

Presclerite: Anterior articulatory region of each abdominal sclerite overlapped by the anterior segment. The term pretergite refers to the presclerite of a tergum (ptg, Fig. 5E), and presternite to the presclerite of a sternum.

Premental shield (pm, Fig. 6D, E): Labial sclerite that articulates apicolaterally with the labial palps.

Pronotum (pn, Fig. 5A): The tergum of the prothorax, which is hypertrophied in workers, and also in queens and males of some ant taxa.

Prora (pr, Fig. 5C; modified from Boudinot et al. 2020): Anteroventral process of abdominal sternite III that contacts the petiole sternite and gives stability to the ventral flexion of the gaster during stinging. It may be located anywhere from the area between the ventral margins of the helcial tergite arch to the anterior face of the poststernite.

Pygidium (py, Fig. 5A, C): In adult female ants, the tergite of abdominal segment VII.
Scape (sc, Fig. 6A): Basalmost antennomere followed apically by the pedicel; formed by the bulbus, the bulbus neck, and the main shaft of the scape.

Scutoscutellar sulcus (as in Gibson 1985): Sulcus impressed along the scutoscutellar suture. The scutoscutellar suture runs across the mesoscutellar-axillar complex, and separates axillae from mesoscutellum.

Stipes (st, Fig. 6D, E): Maxillary sclerite that articulates apically with the maxillary palps and with the galea.

Subpetiolar process (sp, Fig. 5B; as in Fisher and Bolton 2016): Ventral projection of the petiolar poststernite.

Suture and sulcus: A suture is a groove formed by the fusion of two sclerites; sulcus is an impression that corresponds to an apodeme.

Torular lobes (tol, Fig. 6A): Trait formed by the laterally projected median arches of the torulus; similar to frontal lobes.
Additionally, we used the following adjectives to describe vestiture:
Aristate: Shaped basally like a spine-like seta, and bearing a long, thin, flexuous apex.

Buoyant (Fig. 7A): Shaped as if floating in air and weightless.
Elliptic (Fig. 7B): Elongate oval, with the widest part near the middle.
Filiform (Fig. 7C): Filamentous, shaped like a thread. Here used to describe seta with a regular, hair-like shape.

Glabrous: Devoid of hair or cuticular projections.
Helicoid (Fig. 7D): Seta compressed longitudinally and twisted, resembling a helix.

Hook-shaped (Fig. 7E): Seta with the apical portion curved like a hook.
Lanceolate (Fig. 7F): Shaped like a lancehead, with the widest part near the base and the narrowest part near the apex.

Microtrichium (pl microtrichia; Fig. 7F, H, L): Setae-like, minute cuticular projection.

Serrate: With one margin bearing a series of small, sharp, teeth-like projections.
Spatulate (Fig. 7G, H): Shaped like a spatula, with a broad, flat apex that tapers to the base.

Spatulate-costate (Fig. 7I): Spatulate seta with longitudinal parallel ridges.
Spatulate-bicuspid (Fig. 7J): Spatulate seta in which the apex is much wider than the base, so that its apicolateral corners arch over the apicomedial area, giving the seta a bicuspid appearance.

Stout (Fig. 7K): Heavily built.
Tubiform (Fig. 7L): Tubular, not tapering to a point.
Sculpture terminology follows Harris (1979), as below.
Colliculate (Fig. 8A): Covered with continuous, short, rounded prominences.
Confused (Fig. 8E): Sculpture without definite outlines or definite pattern.
Costulate (Fig. 8B): With thin, longitudinal, parallel ridges.
Fossula (pl fossulae; Fig. 8C): Elongate, oblong depressions.
Punctate (Fig. 8D): With fine, impressed punctures.
Rugose (Fig. 8E): Wrinkled.
Smooth: Devoid of any sculpturing.
Strigate (Fig. 8F): With narrow, transverse ridges.
Strigulate (Fig. 8F): Finely or minutely strigate.

## Measurements and indices

We utilized linear morphometry to quantify size and offer a means of comparison with other Ponerinae taxa. Measurements, indices, and abbreviations follow Fisher (2006). All measurements were taken at $80 \times$ power with a Leica MZ125 microscope using an orthogonal pair of micrometers, recorded to the nearest 0.001 mm , rounded to two


Figure 7. Glossary of terminology, part IV: vestiture. Individual parts show SEM images taken from Corrieopone nouragues; paratypes, worker caste. Distinct types of seta and microtrichium are highlighted in orange $\mathbf{A}$ buoyant seta $\mathbf{B}$ elliptic seta $\mathbf{C}$ filiform seta $\mathbf{D}$ helicoid seta $\mathbf{E}$ hook-shaped seta $\mathbf{F}$ lanceolate microtrichium $\mathbf{G}$ spatulate seta $\mathbf{H}$ spatulate microtrichium I spatulate-costate seta $\mathbf{J}$ spatulate-bicuspid seta $\mathbf{K}$ stout, spine-like seta $\mathbf{L}$ tubiform microtrichium. Specimens imaged: CASENT0872031 (A, C, D, K) and CASENT0923158 (B, E-J, L). Images by FA Esteves; available at AntWeb.org. Scale bars: 0.01 mm .


Figure 8. Glossary of terminology, part V: sculpture. Individual parts show SEM images taken from Corrieopone nouragues; holotype, worker (CASENT0830464) A rectangle encloses the colliculate sculpture on the metatibial apicoposterior surface $\mathbf{B}$ rectangle encloses the costulate lateral surface of the pronotum $\mathbf{C}$ arrows indicate the fossulae on the dorsal face of the mandible $\mathbf{D}$ punctate sculpture on the dorsoposterior area of the head $\mathbf{E}$ rectangle encloses the confused rugose sculpture on the petiolar profile $\mathbf{F}$ declivitous face of the propodeum and petiole in dorsal view; arrow indicates the strigulate sculpture; star highlights the strigate sculpture. Images by M Esposito ( $\mathbf{A}, \mathbf{E}$ ) and FA Esteves ( $\mathbf{B}-\mathbf{D}, \mathbf{F}$ ); available at AntWeb.org. Scale bars: $0.1 \mathrm{~mm}(\mathbf{A}, \mathbf{D}) ; 0.2 \mathrm{~mm}(\mathbf{B}, \mathbf{C}, \mathbf{E}, \mathbf{F})$.
decimal places, and presented as minimum and maximum values with holotype measurement within parentheses. Indices are rounded to the nearest integer and expressed as minimum and maximum values with holotype index within parentheses (see original data in Suppl. material 4: Table S4).

HL Head length (Fig. 9A): Maximum longitudinal length of the head, measured from the anteriormost portion of the projecting clypeus to the midpoint of an imaginary line traced across the posterior margin of the head.
HW Head width (Fig. 9A): Maximum width of head, excluding eyes.
SL Scape length (Fig. 9A): Maximum chord length of the main shaft of the antennal scape, excluding basal bulbus and bulbus neck.
WL Weber's length (Fig. 9B): With the mesosoma in profile, the diagonal length from the posteroventral corner of the propodeum to the farthest point on
the anterior face of the pronotum (i.e., anterior inflection point of the pronotum), excluding the neck.
TL Total length: Sum of HL + WL + length of segments A2 to A7. A2 to A7 is measured as follows: maximum length of the petiole in profile (PL; Fig. 9B) + A3 to A7, or gaster length (GL; Fig. 9B).
CI Cephalic index: HW/HL $\times 100$.
SI Scape index: SL/HW $\times 100$.

## A



Figure 9. Measurements taken from Corrieopone nouragues workers and queen $\mathbf{A}$ head in full-face view B body in profile. Abbreviations: GL, gaster length; HL, head length; HW, head width; PL, petiolar length; SL, scape length; WL, Weber's length of mesosoma. Illustrations by FA Esteves.

## Acronyms

CASC California Academy of Sciences, San Francisco, California, U.S.A.
JTLC John T. Longino private collection, Salt Lake City, Utah, U.S.A.
MHNG Musee d'Histoire Naturelle Genève, Geneva, Switzerland
MNHN Muséum Nationale d'Histoire Naturelle, Paris, France
MZSP Museu de Zoologia da Universidade de São Paulo, São Paulo, S.P., Brazil

## Taxonomic treatments

## Taxonomic changes in Ponerinae

Before describing Corrieopone nouragues gen. nov., sp. nov., we must make some adjustments to the taxonomic framework proposed for Ponerinae by Schmidt and Shattuck (2014), which otherwise would render comments on morphological similarities and differences among genera in subsequent sections cumbersome. The following changes are based on assessing female morphology among specimens examined in this study, with images available on AntWeb, and information gathered from relevant taxonomic literature.

## Transfers between Neoponera and Pachycondyla

Neoponera Emery and Pachycondyla Smith are recognized among other Neotropical ponerines by the following combination of characters: The anterior clypeal margin lacks a pair of large teeth-like projections. Torular lobes are closely approximated. Mandibles are triangular to subtriangular, inserted on the anterolateral corner of the head, and armed with numerous teeth. The metapleural gland orifice is closely skirted medially and posteriorly by a well-developed carina. The propodeal spiracle is usually slitshaped; otherwise, the head presents a bilateral carina between the clypeal margin and the anterior margin of the compound eye, and the pretergite of abdominal segment IV presents a stridulitrum. The mesotibia lacks stout, spine-like setae along its dorsal face, and the metatibia presents two spurs. Pretarsal claws are not pectinate. The petiole sternite lacks a posterior spatulate projection that folds posteriad over the remaining sternite; otherwise, the anterior clypeal margin is convex and angulate (see Suppl. material 3: Table S3; Mackay and Mackay 2010; Schmidt and Shattuck 2014).

According to our assessment, Neoponera and Pachycondyla can be set apart from each other by only two characters: the former genus presents distinct arolia between the pretarsal claws and a stridulitrum on the pretergite of abdominal segment IV; the latter does not (see Suppl. material 3: Table S3; Kempf 1961a; Mackay and Mackay 2010; Fernandes et al. 2014; Schmidt and Shattuck 2014). Contrary to Schmidt and Shattuck (2014), the presence or absence of stout, spine-like setae on the posterior portion of the hypopygium is of little importance to distinguish the genera for two


Figure 10. Stout setae on the posterolateral face of the hypopygium in Neoponera and Pachycondyla A Neoponera bucki, worker (UFV-LABECOL-007493); hypopygium disassociated from the pygidium; arrows indicate the spine-like setae B $N$. carinulata, worker (CASENT0845443); arrow indicates the spinelike setae C Pachycondyla crassinoda, worker (CASENT0370917); arrow indicates the spine-like setae D P. harpax, worker (CASENT0374616); spine-like seta highlighted in orange E P. crassinoda, worker (CASENT0372235); left side; arrow indicates the aristate setae $\mathbf{F}$ P. crassinoda, worker (CASENT0372235); right side; arrow indicates the aristate setae; spine-like seta highlighted in orange. Images by JCM Chaul (A) and FA Esteves (B-F); available at AntWeb.org. Scale bars: $0.04 \mathrm{~mm}(\mathbf{A}-\mathbf{C}, \mathbf{E}, \mathbf{F}) ; 0.06 \mathrm{~mm}(\mathbf{D})$.
reasons. First, the hypopygium bears spine-like setae in some Neoponera species [e.g., $N$. bucki (Borgmeier), N. cavinodis, N. crenata, N. striadinodis, and $N$. unidentata; Fig. 10A, B], which are similar to those found in Pachycondyla (Fig. 10C, D). Second, in every Pachycondyla species examined $(N=6)$, some specimens presented a hypopygium armed with aristate setae instead (Fig. 10E). Moreover, among those aristate setae, a few had blunt-apices and resembled spines (Fig. 10F). We assume that this intraspecific and intraindividual shape variation is caused by wear. For example, contact with prey during stinging could erode the apices of pristine aristate setae on the hypopygium, which would then resemble spines.

Other characters with inconsistent diagnostic value include the presence of a metanotal sulcus and the shape of the petiole. The metanotal sulcus is distinctly impressed in most Neoponera species, but is absent to shallowly marked or shows variation in distinctiveness in N. bucki (CASENT0915250), N. crenata species-group members (sensu Mackay and Mackay 2010; CASENT0923100), and N. laevigata (ATPFOR2006, CASENT0902510). In most Pachycondyla, the sulcus is obliterated, yet is present in some specimens of $P$. harpax (specimen CASENT0104808),


Figure II. Pachycondyla procidua comb. rev.; worker A head in full-face view (CASENT0646046) B dorsal view (CASENT0646046) C mesosoma in profile (CASENT0830573) D body in profile (CASENT0646046). Images by JT Longino ( $\mathbf{A}, \mathbf{B}, \mathbf{D}$ ) and FA Esteves $(\mathbf{C})$; available at AntWeb.org. Scale bars: 1 mm .
P. impressa (ECOFOG-IT14-0276-06), P. lattkei (CASENT0217562), and P. striata (CASENT0178185, CASENT0249158). The shape of the petiole in profile ranges from scale-like to cuboid in Neoponera [as in N. carbonaria (Smith), CASENT0915655, and N. crenata, CASENT0915261, respectively]. It is cuboid in most Pachycondyla species, but also D-shaped in P. harpax (CASENT0249149) and scale-like in P. lenkoi Kempf (UFV-LABECOL-000002).

The Neotropical Pachycondyla procidua Emery, 1890 comb. rev. (Fig. 11) was previously assigned to Neoponera by Schmidt and Shattuck (2014). Yet, the species lacks a stridulitrum and distinct arolia (Fig. 12A, B; Kempf 1964). Additionally, it presents the above characters that discriminate Neoponera and Pachycondyla from ponerines in the neotropics (Fig. 11; Kempf 1964; Mackay and Mackay 2010). The metanotal sulcus is distinct (Fig. 11B), while the petiole is scale-like (Fig. 11C, D). The two specimens we examined present aristate hypopygial setae. The setae seem fragile, and in one specimen, most were abraded by the cleaning procedure that preceded SEM imaging (Fig. 12C, D).

Neoponera curiosa (Mackay \& Mackay, 2010) comb. nov. also conforms with the characters shared by Neoponera and Pachycondyla (Fig. 13; Mackay and Mackay 2010). In addition, the species possesses a stridulitrum on the pretergite of abdominal segment IV (Mackay and Mackay 2010). According to the authors of the taxon,


Figure I 2. Pachycondyla procidua comb. rev.; worker (CASENT0830573) A median area of the abdominal pretergite IV in dorsal view $\mathbf{B}$ pretarsus and apex of apicalmost tarsomere in posterodorsal view $\mathbf{C}$ posterolateral face of the hypopygium $\mathbf{D}$ close-up of the posterolateral face of the hypopygium. Images by FA Esteves; available at AntWeb.org. Scale bars: $0.02 \mathrm{~mm}(\mathbf{A}, \mathbf{B}) ; 0.04 \mathrm{~mm}(\mathbf{C}, \mathbf{D})$.
the arolia are underdeveloped. However, note that the adjective was likely used to compare the taxon with other species formerly assigned to Pachycondyla in which the arolium is distinct. For example, Mackay and Mackay (2010) also described the trait as underdeveloped in $N$. striadinodis; yet we examined one specimen determined by W. P. Mackay that has distinct arolia.

## Transfer from Euponera to Leptogenys and new synonymy

The genus Leptogenys Roger occurs in the tropics and subtropics around the world. Most of its species present pectinate or multidentate pretarsal claws, absent elsewhere in Ponerinae (Bolton 1975; Schmidt and Shattuck 2014; Fisher and Bolton 2016). The clypeal medial area usually projects anteriad into a well-developed, angular prominence, generally skirted anteriorly by a translucid lamella; the median area usually bears a longitudinal carina. Torular lobes are small and only conceal the medial portion of the antennal sockets in dorsal view. Mandibles insert on the anterolateral corners of the head. Mandible shapes range from subtriangular to oblique, to falcate, to linear, to bizarre forms in-between. The propodeal spiracle usually presents a round to oval ori-


Figure 13. Neoponera curiosa comb. nov.; holotype, dealated queen (LACMENT226103) A head in fullface view $\mathbf{B}$ dorsal view $\mathbf{C}$ petiole in profile $\mathbf{D}$ body in profile. Images by JE Lattke; available at AntWeb. org. Scale bars: 0.5 mm .
fice. The metatibia bears two spurs; the metabasitarsus does not bear stout, spine-like setae on its dorsal face. The helcium is infra-axial (i.e., helcium positioned ventrad to the midheight of the anterior face of abdominal segment III).

The species originally described as Pseudoponera butteli Forel, 1913, based on specimens collected in Java, Indonesia, was recently assigned to the genus Euponera Forel by Schmidt and Shattuck (2014). However, the above characters setting Leptogenys apart from other ponerines are visible in images of its syntypes (Fig. 14; specimens CASENT0907293, FOCOL1012, FOCOL1013) or were mentioned in its description (Forel 1913), except for the shape of the pretarsal claws. On our behalf, entomologist and curator Dr B. Landry examined one of the syntypes deposited at the NHMG and confirmed the presence of three preapical teeth on the hindleg claws. Therefore, E. butteli becomes Leptogenys butteli (Forel, 1913) comb. nov.

Leptogenys butteli belongs to the L. processionalis species group (sensu Schmidt 2013), which contains $\sim 15$ taxa occurring in the Indomalayan and Australasian regions, viz.: L. birmana Forel, specimen CASENT0907581; L. breviceps Viehmeyer, CASENT0902610; L. crassicornis Emery, CASENT0903953; L. chelifera (Santschi), CASENT0915218; L. dentilobis Forel, CASENT0907365; L. fallax (Mayr), CASENT0915877; L. fortior Forel, CASENT0907393; L. iridescens (Smith),


Figure 14. Leptogenys butteli comb. nov.; syntypes, worker caste $\mathbf{A}$ head in full-face view (CASENT0907293) B dorsal view (CASENT0907293) C head in full-face view (FOCOL1013) D view in profile (CASENT0907293). Images by Z Lieberman (A, B, D) and C Klingenberg (C); available at AntWeb.org. Scale bars: 0.2 mm .

CASENT0901354; L. lucidula Emery, CASENT0235337; L. mutabilis (Smith), CASENT0901355; L. myops (Emery), CASENT0903952; L. processionalis (Jerdon), CASENT0270567; L. processionalis distinguenda (Emery), CASENT0903955; L. strena Zhou; and $L$. tricosa Taylor. Members of this species group are characterized by a quadrate or subquadrate head; oblique mandibles armed with more than three teeth, without distinct basal and masticatory margins (somewhat subtriangular in L. processionalis distinguenda); and scale-like petiole (somewhat nodiform in $L$. crassicornis, L. tricosa, and apparently also in $L$. strena).

When we compared type specimen images and evaluated morphological variation among L. processionalis group members, we found several characters distinguishing between most species in the group. However, we could not find any significant differences between $L$. butteli and $L$. myops (Fig. 15). Their type specimens (all from Java, Indonesia) present the same head proportions and sculpture; shape of the torular lobes; shape of the clypeal anterior projection; width of the longitudinal protrusion on the clypeal median area; distance from the apex of the antennal scape to the posterior margin of the head; size, shape, and location of compound eyes on the head; sculpture and shape of the mandibles; dorsal outline of the mesosoma in profile; indistinctiveness of


Figure 15. Leptogenys myops; worker caste $\mathbf{A}$ head in full-face view; holotype (CASENT0903952) B dorsal view; holotype (CASENT0903952) C head in full-face view (CASENT0281925) D view in profile; holotype (CASENT0903952). Images by W Ericson (A, B, D) and S Hartman (C); available at AntWeb. org. Scale bars: 0.2 mm .
the metanotal sulcus; shape of the mesonotum in dorsal view; height of the petiole in comparison with the propodeum; shape of the petiolar tergite; body-color; and approximate length, amount, and inclination of standing setae. Consequently, we synonymize Leptogenys butteli (Forel, 1913), syn. nov., with Leptogenys myops (Emery, 1887).

## Transfer from Mesoponera to Bothroponera

In the Afrotropics, the genera Bothroponera Mayr and Mesoponera Emery can be discriminated from other ponerines by a combination of characters in the worker caste (Fisher and Bolton 2016). The anterior portion of the head lacks a dorsolateral carina between the clypeal margin and the anterior margin of the eye. Torular lobes are closely approximated medially. The anterior margin of the eyes is located at or anteriad to the midlength of the head. Mandibles are triangular and inserted on the anterolateral corners of head; masticatory margins bear four or more teeth; basolateral or dorsal faces lack pits of any shape (an oblique dorsolateral sulcus may be present). The dorsoposterior area of the propodeum is devoid of spine-like or tooth-like projections. The mesotibia lacks stout spine-like setae on its dorsal face, and the metatibia bears two distinct spurs on its ventroapical face. The
helcium is positioned ventrad the midlength of the anterior face of abdominal segment IV. Note that contrary to Schmidt and Shattuck (2014), only the basal portion of the masticatory margin of the mandibles is edentate in M. subiridescens; the apical portion is armed with four to seven, but sometimes more, teeth (the count includes denticles).

Bothroponera species have a propodeum with a broad dorsal face, a slit-shaped propodeal spiracle, and a nodiform petiole (Schmidt and Shattuck 2014). The metanotal sulcus is usually obliterated; however, intraspecific variation exists, and the sulcus is weakly impressed in some specimens [as B. ilgii (Forel), specimen CASENT0235600; and B. kruegeri (Forel), CASENT0235604]. Regardless, if present, the sulcus does not interrupt the dorsal outline of the mesosoma in profile. On abdominal segment IV, a strongly impressed constriction separates presclerites from postsclerites, as in B. berthoudi (Forel), specimen CASENT0902470; or moderately so, as in B. silvestrii CASENT0235599. Schmidt and Shattuck (2014) assigned Bothroponera species in two groups. The B. sensu stricto group members share a strongly sculptured body, hypertrophied torular lobes, and a metapleural gland orifice closely skirted medially and posteriorly by a well-developed carina; these characters are absent in the B. sulcata group. Notwithstanding the differential diagnosis given by the authors above, several species of the sensu stricto group [e.g., B. berthoudi, CASENT0902470; B. granosa (Roger), CASENT0250375; and B. laevissima (Arnold), CASENT0902471] have the propodeal dorsum as narrow as those of the sulcata group members.

In Mesoponera, the propodeum is tectiform (i.e., roof-shaped), with its lateral surfaces diverging while sloping ventrad from the noticeably narrow dorsum. In most species, the propodeal dorsal face presents slightly or moderately bulging lateral margins, with the medial area slightly concave posteriorly; the lateral margins may be somewhat parallel to one another (as in M. caffraria, CASENT0915251) or diverge continuously posteriad (as in M. ambigua, CASENT0249194). In a few species, the propodeal dorsum is narrower and transversely convex (as in M. subiridescens, CASENT0003151). The propodeal spiracle is usually round to oval [except for $M$. caffraria and subspecies, M. ingesta (Wheeler), and M. subiridescens; see CASENT0906219]. The petiole is shaped like an upward-pointing wedge in profile, with the anterior and posterior faces of the tergite tapering to a thin dorsal margin. The metanotal sulcus is clearly distinct and usually deeply impressed, and it indents the mesosoma outline in profile. The constriction between the presclerites and postsclerites of abdominal segment IV may be obliterated to moderately impressed. The torular lobes are not hypertrophied. Body is mostly smooth and shiny or densely and uniformly sculptured by fine punctures. These characters are as described here in every Mesoponera species except one.

The species originally described as Pachycondyla (Bothroponera) escherichi Forel, 1910, based on one worker collected in Eritrea (Fig. 16), was recently combined in Mesoponera by Schmidt and Shattuck (2014). However, the species is clearly misassigned. Mesoponera escherichi presents every character that sets Bothroponera and Mesoponera apart from the remaining Ponerinae genera in the Afrotropics, including the absence of a basolateral or dorsal pit on the mandibles (Forel 1910). The shape of the propodeal dorsal face corresponds to that of the B. sulcata group. The propodeal spiracle is slit-


Figure 16. Bothroponera escherichi comb. nov.; holotype, worker (CASENT0907252) A dorsal view B head in full-face view C view in profile. Images by W Ericson; available at AntWeb.org. Scale bars: 0.5 mm .
shaped. The petiolar tergite is unaligned in the image we evaluated; even so, it shows a somewhat nodiform petiole (clearly not cuneiform) that is reasonably accommodated by the variation seen in the Bothroponera. Moreover, Forel (1910) stated that the anterior and posterior faces of the petiole are vertical surfaces. The metanotal sulcus is present but weakly impressed, and does not interrupt the dorsal outline of the mesosoma in profile. The presclerites and postsclerites of abdominal segment IV are separated by a moderately impressed constriction. The torular lobes are not hypertrophied. Sculpture is not strongly impressed: most of the head and the dorsal face of the mesosoma is densely foveolate (i.e., covered by small pits that are wider than punctures); the lateral face of the mesosoma is mostly costulate; the petiole is somewhat shiny; the gaster is slightly punctate, and is shinier than the head and mesosoma. Therefore, M. escherichi becomes Bothroponera escherichi (Forel, 1910), comb. nov., a member of the B. sulcata species group.

## Identification key for the Neotropical genera of Ponerinae based on workers

We did not consider Pachycondyla vieirai Mackay \& Mackay in this key, as it was not examined and the taxon description was uninformative for our purposes. The species was considered incertae sedis in Pachycondyla (Schmidt and Shattuck 2014).

1 Mandibles long and linear in full-face view, inserted at the middle of the anterior margin of the head, their bases closely approximate (Fig. 17A) 2

- Mandibles with variable shape, but always inserted at the anterolateral corners of the head, their bases conspicuously separated (Fig. 17AA) 3


Figure 17. Identification key, couplet 1 A Odontomachus erythrocephalus, worker (CASENT0649085) AA Hypoponera aliena, worker (CASENT0281913). Images by JT Longino and S Hartman, respectively; available at AntWeb.org. Illustrations by Jessica Huppi.

2 Nuchal carina (i.e., carina that separates dorsal from posterior surfaces of the head) and paired dark posterior apophyseal lines converge in a V-shape at the midline of the posterior margin of the head (Fig. 18A, B), and join a sharp mediodorsal sulcus that runs longitudinally on the posterior half of the head. Dorsalmost tooth of apical mandibular series usually truncated. Apex of the petiolar node usually conical or pointed.

Odontomachus

- $\quad$ Nuchal carina forms an uninterrupted curve across the posterodorsal extremity of the head (Fig. 18AA); paired dark apophyseal lines absent (Fig. 18BB); median sulcus absent or ill-defined and shallow on the posterior half of the head. Dorsalmost tooth of apical mandibular series usually acute. Petiolar node with varying shape: subtriangular to scale-like, unarmed to bidentate $\qquad$ Anochetus


Figure 18. Identification key, couplet 2 A, B Odontomachus meinerti, worker (CASENT0178690) AA BB Anochetus diegensis, worker (CASENT0178673). Images by A Nobile; available at AntWeb.org. Illustrations by Jessica Huppi.

3 In full-face view, anterior part of the torular lobes widely separated and usually not confluent; separated by a rounded, truncated, or broadly triangular section of the clypeus (Fig. 19A). The lateral margins of the lobes variously shaped, but only rarely with a pinched-in appearance posteriorly (Fig. 19B).

- In full-face view, anterior section of the torular lobes confluent or closely approximated; separated by a narrow triangular portion of the clypeus or by a very narrow cuticular strip, which extends posteriad between them (Fig. 19AA). The lateral margins of the lobes always with a pinched-in appearance posteriorly (Fig. 19BB)


Figure 19. Identification key, couplet 3 A, B Platythyrea angusta, worker (CASENT0907126) AA, BB Hypoponera aliena, worker (CASENT0281913). Images by W Ericson and S Hartman, respectively; available at AntWeb.org. Illustrations modified from artwork by Jessica Huppi (A, B), and by Jessica Huppi (AA, BB).

4 Mandible triangular, with distinct basal and masticatory margins; masticatory margin edentate or with numerous short teeth (Fig. 20A). Metatibia with 2 distinctly pectinate spurs; the posterior spur usually much larger than the anterior (Fig. 20B). Helcium in profile located approximately at midheight of the anterior face of the first gastral segment (abdominal segment III), so that the first gastral segment does not have a long vertical anterior face in profile. Petiole subrectangular to subcylindrical; the posterior face usually with carinate lateral margins. Dorsal surfaces of the head and the mesosoma without erect/suberect setae. Fine, dense shagreened sculpture, with associated larger punctures $\qquad$ Platythyrea

- Mandible pitchfork-like; with indistinct basal and masticatory margins; armed with 3 noticeably long, curved teeth; the apical tooth so long and curved that it reaches or surpasses the anterolateral corner of the head opposite from its insertion when the mandible is closed (Fig. 20AA). Metatibia with only one spur, pectinate (Fig. 20BB). Helcium in profile located ventrad to the midheight of the anterior face of the first gastral segment (abdominal segment III), so that the first gastral segment has a long vertical anterior face
in profile. Shape of the petiole ranging from a thick, broad scale with sharp lateral margins to a somewhat cuboid node; never subrectangular to subcylindrical with carinate posterolateral margins. Dorsal surfaces of the head and the mesosoma usually with erect/suberect setae, at least partially. Sculpture varying from smooth and shiny to finely shagreened to finely punctate and rugulose


## Thaumatomyrmex

5 Ventral apex of the metatibia with only one spur, which is large and pectinate (Fig. 21A) 6

- Ventral apex of the metatibia with two spurs; the posterior spur always larger and pectinate (Fig. 21AA). 9
6 Dorsal face of the metabasitarsus with stout, spine-like setae amid regular, filiform setae (Fig. 22A); similar spine-like setae also present on mesobasitarsus and mesotibia

Centromyrmex

- Dorsal face of the metabasitarsus vested with filiform setae; stout, spine-like setae absent (Fig. 22AA). Stout, spine-like setae may occur on either mesobasitarsus or mesotibia, but if so, they are absent from the metabasitarsus ...... 7
7 Medial portion of the clypeus projected anteriad, overhanging the anterior clypeal margin in full-face view (Fig. 23A); projection frequently mucronate: anteromedian point with an abrupt, thin, conspicuous prominence (Fig. 23A, mid-image). Mandible subtriangular to falcate (Fig. 23B). Arolium usually well-developed (Fig. 23C)

Simopelta

- Medial portion of the clypeus does not overhang the anterior clypeal margin in full-face view (Fig. 23AA); anterior clypeal margin slightly convex. Mandible triangular (Fig. 23BB). Arolia indistinct (Fig. 23CC) ............................ 8
8 Subpetiolar process in ventrolateral (oblique) view with a pair of angulate projections located posteriorly (Fig. 24A). In profile, subpetiolar process with an anterior translucent fenestra (Fig. 24B), and with a sharp posteroventral angle (Fig. 24C). Maxillary palps with 2 segments Ponera
- $\quad$ Subpetiolar process in ventrolateral (oblique) view without a pair of angulate projections located posteriorly (Fig. 24AA). In profile, subpetiolar process usually without an anterior fenestra (Fig. 24BB); with posteroventral portion rounded to acutely angulate (Fig. 24CC). Maxillary palps with $0-1$ segments .

Hypoponera
Hindlegs usually with pectinate pretarsal claws (Fig. 25A), rarely with only $1-2$ small preapical teeth; if pretarsal claws not pectinate, then mandible with only $1-2$ teeth; if mandibles with $>3$ teeth, then pretarsal claws pectinate. Torular lobes distinctly fail to cover the entire antennal sockets in full-face view (Fig. 25B)

Leptogenys

- Pretarsal claws of hindlegs never pectinate; the claws are simple or with a basal or preapical tooth (Fig. 25AA). Mandible edentate or with variable numbers of teeth; if basal or preapical teeth are present on pretarsal claws, then mandible with 4 or more teeth. The torular lobes may or may not conceal the antennal sockets in full-face view (Fig. 25BB)


Figure 20. Identification key, couplet 4 A Platythyrea angusta, worker (CASENT0907126) AA Thaumatomyrmex zeteki, worker (CASENT0318451) B P. sinuata, worker (CASENT0217573) BB T. zeteki, worker (CASENT0318452). Images by W Ericson, M Esposito, W Ericson, and FA Esteves, respectively; available at AntWeb.org. Illustrations modified from artwork by Jessica Huppi (A), by FA Esteves (B), and Jessica Huppi ( $\mathbf{B}, \mathbf{B B}$ ).


Figure 2I. Identification key, couplet 5 A Simopelta paeminosa, worker (CASENT0217574) AA Dinoponera quadriceps, worker (CASENT0217519). Images by FA Esteves, available at AntWeb.org. Illustrations by Jessica Huppi.


Figure 22. Identification key, couplet 6 A Centromyrmex alfaroi, worker (ANTWEB1032026) AA Simopelta paeminosa, worker (CASENT0217574). Images by JCM Chaul and FA Esteves, respectively; available at AntWeb.org. Illustrations modified from artwork by Jessica Huppi ( $\mathbf{A}$ ), and by Jessica Huppi ( $\mathbf{A A}$ ).


Figure 23. Identification key, couplet 7 A, B left image: Simopelta paeminosa, paratype, worker (CASENT0902468); middle image: S. pergandei, holotype, worker (CASENT0907294) AA, BB Hypoponera alw06, worker (CASENT0173727) C S. paeminosa (CASENT0902468) CC Hypoponera vc01, worker (CASENT0766197). Images by W Ericson, Z Lieberman, A Nobile, and FA Esteves, respectively; available at AntWeb.org. Illustrations by FA Esteves ( $\mathbf{A}, \mathbf{B}$ ), Jessica Huppi ( $\mathbf{A A}, \mathbf{B B}$ ), and modified from artwork by Jessica $\operatorname{Huppi}(\mathbf{C}, \mathbf{C C})$.


Figure 24. Identification key, couplet 8 A, B, C Ponera exotica, worker (CASENT0100692) AA Hypoponera vc01, worker (CASENT0766197) BB, CC H. parva (CASENT0260431). Images by FA Esteves $(\mathbf{A}, \mathbf{B}, \mathbf{C}, \mathbf{A A})$ and $S$ Hartman (BB, CC); available at AntWeb.org. Illustrations by Jessica Huppi. Abbreviations: A2, abdominal segment II; A3, abdominal segment III.

10 Mandibles falcate, elongated, and slender, with long, conspicuous teeth; apical tooth much longer than other teeth (Fig. 26A)
.Belonopelta

- Mandibles triangular or subtriangular, without long, conspicuous teeth (Fig. 26AA). 11
11 Dorsal face of mesotibiae covered with abundant, stout, spine-like setae (Fig. 27A). Prora variable, usually reduced and hardly visible in profile, but never projected ventro-anteriorly as a long, acute prominence (Fig. 28A)......

Cryptopone

- Dorsal face of mesotibiae usually without abundant, stout, spine-like setae (Fig. 27AA). If spine-like setae present along dorsal face of mesotibia (Fig. 27AAA), then prora in profile projected ventro-anteriorly as a long, acute prominence (Fig. 28AA); otherwise prora with variable shape. 12
12 Massive ants (head width greater than 4.0 mm ). Anterior clypeal margin with a pair of large projecting teeth (Fig. 29A)

Dinoponera

- Smaller ants (head width less than 4.0 mm ). Anterior clypeal margin without a pair of large projecting teeth (Fig. 29AA)


Figure 25. Identification key, couplet $9 \mathbf{A}$ Leptogenys pe02, worker (CASENT0372210) AA Dinoponera longipes, worker (CASENT0217518) B L. wheeleri, worker (CASENT0178811) BB D. longipes, worker (CASENT0004663). Images by FA Esteves (top two rows) and A Nobile (bottom two rows); available at AntWeb.org. Illustrations modified from artwork by Jessica Huppi (A, B), by Jessica Huppi (AA), and FA Esteves (BB).


Figure 26. Identification key, couplet 10 A left image: Belonopelta attenuata, worker (ICN100255); middle image: B. deletrix, worker (CASENT0260514) AA Cryptopone cf. guatemalensis (CASENT0646802). Images by JT Longino (top and bottom left) and S Hartman (top middle); available at AntWeb.org. Illustrations by FA Esteves (A), and modified from artwork by Jessica Huppi (AA).

13 In ventral view, petiolar sternite with a posterior spatulate projection folded posteriad over the remaining sternite, so that in profile, the posterior portion of the subpetiolar process presents a long, acute projection strongly directed posteriad (Fig. 30A). Anterior margin of clypeus truncated to emarginated, never entirely convex

Rasopone

- Petiolar sternite usually without a posterior spatulate projection folded posteriad over the remaining sternite (Fig. 30AA). If a posterior spatulate projection is present and the posterior portion of the subpetiolar process is somewhat directed posteriad in profile, then the anterior clypeal margin is convex and angulate; otherwise, the anterior margin of clypeus is variable in shape .. 14
14 Mandible edentate (Fig. 31A). Clypeus in full-face view with a truncate anteromedial projection that overhangs the basal portion of the closed mandibles (Fig. 31B). Ventral face of the hypopygium (abdominal segment VII sternite) longitudinally concave, with posterior region bearing stout, hookshaped setae (Fig. 31C, D); in profile, hook-shaped setae visible. Gaster in profile and in dorsal view without a girdling constriction (presclerites of abdominal segment IV, the second gastral segment, forming an even surface with postsclerites; Fig. 31E)

Corrieopone gen. nov.

- Mandible dentate (with $>4$ teeth/denticles; Fig. 31AA). Clypeus variable in shape, but never with truncate anteromedial projection that overhangs the mandibles (Fig. 31BB). Ventral face of the hypopygium (abdominal segment VII sternite) without longitudinal concavity, and never bearing stout, hookshaped setae (Fig. 31CC, DD). Gaster in profile and in dorsal view with or without a distinct impression between the presclerites and postsclerites of the second gastral segment (abdominal segment IV) that appears as a girdling constriction (Fig. 31EE) 15


Figure 27. Identification key, couplet 11 A Cryptopone gilva, worker (CASENT0006054) AA Dinoponera quadriceps, worker (CASENT0217519) AAA Pseudoponera cognata, worker (CASENT0008155). Images by FA Esteves; available at AntWeb.org. Illustrations modified from artwork by Jessica Huppi.


Figure 28. Identification key, couplet 11 A Cryptopone gilva, worker (CASENT0260411) AA Pseudoponera gilberti, worker (CASENT0828638). Images by S Hartman and FA Esteves, respectively; available at AntWeb.org. Illustrations modified from artwork by Jessica Huppi. Abbreviations: A2, abdominal segment II; A3, abdominal segment III.


Figure 29. Identification key, couplet 12 A Dinoponera quadriceps, worker (CASENT0217519) AA Pachycondyla striata, worker (UFV-LABECOL-000291). Images by S Hartman and JT Longino, respectively; available at AntWeb.org. Illustrations by FA Esteves( $\mathbf{A}$ ), and modified from artwork by Jessica Huppi (AA).


Figure 30. Identification key, couplet 13 A Rasopone panamensis, worker (CASENT0644252) AA Corrieopone nouragues, holotype, worker (CASENT0830464). Images by JT Longino and W Lee, respectively; available at AntWeb.org. Illustrations modified from artwork by Jessica Huppi (A), and by FA Esteves (AA). Abbreviations: A2, abdominal segment II; A3, abdominal segment III.

15 Stridulitrum present on abdominal pretergite IV (Fig. 32A) ..................... 16

- Stridulitrum absent from abdominal pretergite IV (Fig. 32AA)................ 17

16 Propodeal spiracle round or ovoid (Fig. 33A), never slit-shaped, and preocular carina absent (Fig. 33B) ............................................Mayaponera (part)

- Propodeal spiracle usually slit-shaped (Fig. 33AA), but if round, the preocular carina is present (Fig. 33BB)

Neoponera
17 Metapleural gland orifice shaped as a curved slit aperture directed posterodorsally (Fig. 34A). In profile, prora projected ventro-anteriorly as a long, acute prominence (Fig. 34B); in anterior view, prora similar to a soup spoon (or a spatula), transverse on abdominal sternite III. Mandible with 6 or 7 teeth/ denticles. $\qquad$

- Metapleural gland orifice variable, but never shaped as a curved slit aperture (Fig. 34AA). Prora variable, but never projected ventro-anteriorly as a long, acute prominence (Fig. 34BB). Mandibles usually with 9 or more teeth/denticles.


Figure 3 I. Identification key, couplet 14 A,B Corrieopone nouragues, holotype, worker (CASENT0830464) AA, BB Neoponera emiliae (JTLC000015100) C, D, E Corrieopone nouragues, paratypes, worker caste (CASENT0830465; CASENT0645962, insert) CC,DD,EE Neoponerafauveli, worker (CASENT0249142). Images by W Lee (A, B), JT Longino (AA, BB, E), FA Esteves (C, D), and R Perry (CC, DD, EE); available at AntWeb.org. Illustrations by FA Esteves ( $\mathbf{A} \mathbf{- E}$ ), and modified from artwork by Jessica Huppi (AAEE). Abbreviations: A3, abdominal segment III; A4, abdominal segment IV; A7, abdominal segment VII.

18 Propodeal spiracles slit-shaped (Fig. 35A). In full-face view, lateral surfaces of torular lobes with large, smooth, shiny area, which is mostly or entirely glabrous (Fig. 35B). In profile, petiole usually cuboid (Fig. 36A). Hypopygium in profile with spine-like or aristate setae on posteriormost portion (Fig. 36B)

Pachycondyla

- Propodeal spiracles round (Fig. 35AA); never slit-shaped. In full-face view, lateral surfaces of torular lobes covered uniformly by same sculpture and setae (Fig. 35BB). In profile, petiole shaped as an upward-pointing wedge (Fig. 36AA). Hypopygium in profile usually without spine-like or aristate setae on posteriormost portion (Fig. 36BB)

Mayaponera (part)


Figure 32. Identification key, couplet 15 A Neoponera fauveli, worker (CASENT0428712) AA Pachycondyla indet., worker (CASENT0006090). Images by FA Esteves; available at AntWeb.org. Illustrations modified from artwork by Jessica Huppi. Abbreviations: A2, abdominal segment II; A3, abdominal segment III; A4, abdominal segment IV.


Figure 33. Identification key, couplet 16 A Mayaponera constricta (CASENT0643470) AA Neoponera bugabensis (CASENT0217570) B M. constricta (CASENT0845824, left image; CASENT0643470, middle image) BB $N$. bugabensis (CASENT0217570, left and middle images). Images by FA Esteves (A, AA; B, BB, middle images), W Lee (B, left image), and W Ericson (BB, left image); available at AntWeb.org. Illustrations modified from artwork by Jessica Huppi.


Figure 34. Identification key, couplet 17 A Pseudoponera stigma, worker (CASENT0922570) AA, BB Pachycondyla crassinoda, worker (CASENT0830390) B Pseudoponera gilberti, worker (CASENT0828638). Images by FA Esteves; available at AntWeb.org. Illustrations modified from artwork by Jessica Huppi. Abbreviations: A2, abdominal segment II; A3, abdominal segment III.


Figure 35. Identification key, couplet 18 A Pachycondyla crassinoda, worker (CASENT0830390) AA Mayaponera conicula, paratype, worker (CASENT0923099) B P. striata, worker (CASENT0923096) BB $M$. pergandei, worker (CASENT0249156). Images by FA Esteves (A), W Lee (AA, B), and R Perry (BB); available at AntWeb.org. Illustrations modified from artwork by Jessica Huppi.


Figure 36. Identification key, couplet 18 A Pachycondyla crassinoda, worker (CASENT0830390) AA Mayaponera conicula, paratype, worker (CASENT0923099); petiole disarticulated from gaster B P. lattkei, paratype, worker (CASENT0217562) BB M. cernua, worker (CASENT0923098). Images by FA Esteves ( $\mathbf{A}, \mathbf{B}, \mathbf{B B}$ ) and W Lee ( $\mathbf{A} \mathbf{A}$ ); available at AntWeb.org. Illustrations modified from artwork by Jessica Huppi ( $\mathbf{A}$ ), by FA Esteves ( $\mathbf{A} \mathbf{A}$ ), and Jessica Huppi (B, BB). Abbreviations: A3, abdominal segment III; A4, abdominal segment IV; A6, abdominal segment VI; A7, abdominal segment VII.

## Corrieopone, gen. nov.

http://zoobank.org/2B32773A-92E1-41B2-BC74-EC0B8ECD4A0A
Figs 7, 8, 37-44, 52A, 53-57
Type species. Corrieopone nouragues sp. nov., by present designation.
Diagnosis based on workers. Medium-sized, slender Neotropical ants (TL 6.57.1 mm; Fig. 37) with characters of Ponerinae (as in Fisher and Bolton 2016) and Ponerini (as in Schmidt and Shattuck 2014), in addition to the following (asterisks indicate putative autapomorphies):

1. Mandibles triangular, with distinct masticatory and basal margins; inserted at the anterolateral corners of the head (Fig. 38A, E).
2. Mandibles edentate (Fig. 38A, E).
3. Mandible devoid of any pit or sulcus: basolateral and dorsal pits and dorsolateral and dorsomasticatory sulci absent (Fig. 38A-C, E).
4.     * Clypeus complex: In dorsal view, clypeus projected anteromedially as a broad, truncated prominence, overhanging the basal margins of the mandibles, and overlapping the basal portion of the masticatory margins of fully closed mandibles; anterior margin of the clypeal projection approximately as wide as the distance between the lateral arches of the toruli, devoid of stout setae or additional protrusions (Figs 37B, 38A, E). In profile, clypeal projection with a broad anteroventral face (avf, Fig. 39A-D), which extends ventroposteriorly (i.e., obliquely) from the clypeal dorsal face in almost 90 degrees; the ventralmost point of this anteroventral face meets the "true" clypeal ventral face (= surface of the clypeal infold in Boudinot et al. 2021; vf, Fig. 39A-D). In anteroventral view, clypeal anteroventral face subrectangular (avf, Fig. 39D). Median area of the clypeus bulging (Fig. 37A, E); seen in profile, it ascends steeply from the clypeal anterior margin to the torular lobes, with posterior portion slightly convex (Fig. 38B, D).
5. In dorsal view, torular lobes closely approximated.
6. In dorsal view, torular lobes medium- to small-sized: not concealing the lateral arches of the toruli (Fig. 38F). Median and lateral arches of the torulus with discontinuous posterior margins (Fig. 38D).
7. In profile, torular lobes located at the dorsalmost part of a prominence formed by the clypeal median area and the frontal carinae (Fig. 38B).
8. Compound eyes small and located immediately anterior to the midline of the head (Fig. 38A, B, D).
9. Ocelli absent (Fig. 38A).
10. Labrum apically bilobed; with a long, acute cleft at the midpoint of its apical margin. Lobes broadly rounded apicolaterally and unarmed (Fig. 39E).
11. Palpal formula: 4,4 (four maxillary, four labial palpomeres; Fig. 39F).
12. Mesonotum rounded, dome-shaped in profile (Fig. 40A), rounded in dorsal view (Fig. 37A).
13. Notopleural suture distinct (Fig. 40A, B).
14. Metanotal sulcus deeply impressed (Fig. 40A).


Figure 37. Corrieopone nouragues, holotype, worker (CASENT0830464) A body in dorsal view B head in full-face view $\mathbf{C}$ body in profile. Images by W Lee; available at AntWeb.org.
15. Mesopleuron in profile divided into anepisternum and katepisternum (Fig. 40A, B).
16. Metathoracic spiracle concealed by a spiracular lobe (Fig. 40B).
17. Orifice of the metapleural gland round, opening posterolaterally on the metapleuron, with its ventral margin atop the posteriormost portion of the metapleural carina (Fig. 40C, D).
18. Metapleural longitudinal flange absent (Fig. 40C, D).
19. Propodeal dorsum devoid of a median longitudinal groove or impression.
20. Propodeum unarmed: without dorsoposterior projections (Fig. 40A).
21. In profile, propodeal lobe round, not surpassing posteriorly the dorsoposteriormost point of the rim of the propodeal foramen (Fig. 40C).
22. Propodeal spiracle slit-shaped (Fig. 40C).


Figure 38. Corrieopone nouragues, paratype, worker (CASENT0872031) A head in full-face view B head in profile; arrow indicates the torular lobes atop a prominence formed by the clypeal median area and the frontal carinae $\mathbf{C}$ dorsolateral face of the mandible $\mathbf{D}$ clypeus and torulus in profile; the arrowhead indicates the discontinuity between the posterior margins of the median and lateral arches of the torulus $\mathbf{E}$ dorsal face of clypeus and mandibles $\mathbf{F}$ torular lobes in dorsal view; the arrow points to the exposed lateral arch of the torulus. Images by FA Esteves; available at AntWeb.org. Scale bars: $0.5 \mathrm{~mm}(\mathbf{A}, \mathbf{B}) ; 0.3 \mathrm{~mm}(\mathbf{C}, \mathbf{D}, \mathbf{E}, \mathbf{F})$.
23. Mesosternal process bidentate; metasternal process bilobate, long (Fig. 40F).
24. Metacoxal cavities open; cavities tightly encircled by cuticle, but cuticular annulus not fused (Fig. 40E; see comment in the following section).
25. Calcar of strigil with a basoventral lamella (Fig. 41A, B).
26. Probasitarsus with anterior and ventral faces densely vested with spatulatecostate setae (Fig. 41A), except for the shorter, spatulate-bicuspid setae present on the area immediately anterior to the comb of strigil (Fig. 41B).
27. Row of stout, spine-like setae present on the posterior face of the probasitarsal notch, parallel to the comb of strigil (Fig. 41B).
28. Two well-developed mesotibial spurs: anterior spur simple, posterior spur serrate (Fig. 41C, D).


Figure 39. Corrieopone nouragues, paratype, worker caste $\mathbf{A}$ clypeus in profile; mandible removed $\mathbf{B}$ anterior part of the head in oblique anterior view; mandibles and maxilla-labial complex removed $\mathbf{C}$ clypeus in oblique anterior view; mandibles and maxilla-labial complex removed $\mathbf{D}$ anterior part of the head in ventral view; mandibles and maxilla-labial complex removed $\mathbf{E}$ outer face of the labrum; basal towards the top of the image $\mathbf{F}$ outer face of the maxillolabial complex; basal towards the bottom of the image; Roman numerals indicate the count of maxillary and labial palpomeres. Specimens imaged: CASENT0923158 (E, F); CASENT0872031 (A-D). Images by FA Esteves; available at AntWeb.org. Abbreviations: avf, clypeal anteroventral face; df, clypeal dorsal face; $\mathbf{v f}$, clypeal ventral face. Scale bars: 0.1 mm .
29. Apparent metatibial gland cuticular patch present on the apicoposterior face of the metatibia, next to the posterior metatibial spur (Fig. 41E).
30. Two well-developed metatibial spurs: anterior spur simple, posterior spur pectinate (Fig. 41F).


Figure 40. Corrieopone nouragues, paratypes, worker caste $\mathbf{A}$ mesosoma in profile $\mathbf{B}$ mesopleuron and part of metapleuron and propodeum in profile $\mathbf{C}$ katepisternum, metapleuron, and propodeum in profile D metapleural gland orifice in posterolateral view $\mathbf{E}$ metapleuron (and posterior part of the mesopleuron) in ventral view; arrowhead indicates the unfused annulus around the metacoxal cavity $\mathbf{F}$ mesosternal and metasternal processes in posterolateral view, highlighted by arrowhead and star, respectively. Specimens imaged: CASENT0872031 (A); CASENT0923158 (B-F). Images by FA Esteves; available at AntWeb. org. Scale bars: 0.2 mm .
31. Stout, spine-like setae absent from the dorsal face of mid-and hindlegs (Fig. 42A, B).
32. Ventral faces of the second, third, and fourth tarsomeres of fore-, mid-, and hindlegs with a paired row of stout, spine-like setae skirting the midline of each segment (Fig. 42C-E).


Figure 41. Corrieopone nouragues, worker caste $\mathbf{A}$ protibia and protarsus in anterior view $\mathbf{B}$ protibial apex and probasitarsus in posteroventral view $\mathbf{C}$ mesotibial apex and basal portion of the mesobasitarsus in ventral view $\mathbf{D}$ mesotibial apex and basal portion of the mesobasitarsus in posterior view $\mathbf{E}$ metatibial apex in posterior view; note the lighter, colliculate cuticular patch next to the posterior metatibial spur $\mathbf{F}$ metatibial spurs in anterior view. Specimens imaged: holotype CASENT0830464 (E); paratype CASENT0923158 (A, C, D, F); paratype CASENT0872031 (B). Images by FA Esteves (A-D, F) and M Esposito (E); available at AntWeb.org. Scale bars: 0.1 mm .
33. Pro-, meso-, and metapretarsi with simple claws (Fig. 42C-F).
34. Arolia indistinct (Fig. 42F).
35. Petiole sessile, with high, unarmed, conic, scale-like node (i.e., tergite narrow in profile and dorsal view; Figs 37A, C, 43A).
36. Petiolar tergite with anteroventral lateral carina (= lateral, dorsoventral carina in Ward 1990, character 29); posteroventral portion of tergite strigate (Fig. 43A).


Figure 42. Corrieopone nouragues, paratypes, worker caste $\mathbf{A}$ midleg (minus coxa) in posterior view $\mathbf{B}$ hindleg (minus coxa) in anterior view $\mathbf{C}$ apex of the foreleg in anteroventral view $\mathbf{D}$ apex of the midleg in anteroventral view $\mathbf{E}$ apex of the hindleg in anteroventral view $\mathbf{F}$ apicalmost tarsomere and pretarsus of the foreleg in anteroventral view. Specimens imaged: CASENT0923158 (A-C, E, F); CASENT0872031 (B). Images by FA Esteves; available at AntWeb.org. Scale bars: 0.1 mm .
37. Petiolar laterotergite distinct (Fig. 43A).
38. Proprioceptor zone on anterior disc of petiolar sternite shaped as a large circular area (Fig. 43B).


Figure 43. Corrieopone nouragues, paratypes, worker caste $\mathbf{A}$ petiole in profile; arrowhead indicates the anteroventral lateral carina of the petiolar tergite $\mathbf{B}$ petiolar sternite in postlateroventral view; arrow indicates the proprioceptor zone on the anterior disc of the sternite $\mathbf{C}$ gaster in profile $\mathbf{D}$ helcium and prora in anterior view $\mathbf{E}$ the posterior portion of the abdominal tergite III and the anterior portion of the abdominal tergite IV in dorsal view; arrowhead indicates the stridulitrum $\mathbf{F}$ stridulitrum on abdominal pretergite IV. Specimens imaged: CASENT0923158 (A, B, D); CASENT0872031 (C, E, F). Images by FA Esteves; available at AntWeb.org. Scale bars: 0.1 mm .
39. Petiolar sternite without posterior spatulate projection (Fig. 43B); articulation with helcium visible in ventral view.
40. Helcium infra-axial: positioned ventrad the midheight of the anterior face of abdominal segment III (Fig. 43C).
41. Prora present as a lip-shaped, transverse projection on the anterior portion of the abdominal poststernite III (Fig. 43C, D); projection not extending anteriorly to the area between the ventroposterior margins of helcial tergite (Fig. 43D).
42. Abdominal segment IV tubular: tergite and sternite with similar lengths; tergite not arched (Fig. 43C).
43. Presclerites of abdominal segment IV forming an even surface with postsclerites: girdling constriction absent (Fig. 43C).
44. Stridulitrum present on abdominal pretergite IV, small (Fig. 43E, F).
45. Pygidium devoid of stout, spine-like setae or spine-like microtrichia; dorsal face convex.


Figure 44. Corrieopone nouragues, worker caste $\mathbf{A}$ the concave ventral face of the hypopygium armed with stout, hook-shaped setae (see arrowheads) on its posteriormost portion B hypopygium in ventrolateral view $\mathbf{C}$ close-up of the median carina (see arrowhead) and hook-shaped setae of the hypopygium in ventrolateral view $\mathbf{D}$ hook-shaped setae of the hypopygium in posteroventral view. Specimens imaged: holotype CASENT0830464 (A); paratype CASENT0923158 (B-D). Images by FA Esteves; available at AntWeb.org. Scale bars: 0.1 mm .
46. * Ventral face of hypopygium longitudinally concave (Fig. 44A, B). Longitudinal carina present at midline of concavity's posterior portion, skirted laterally by stout, hook-shaped setae (Fig. 44C, D).
47. Hypopygium posterolateral region without stout, spine-like setae or spine-like microtrichia.

## Comments on worker characters

The enumeration below corresponds to character numbers presented above.

1. The mandibles articulate with the anterolateral corners of the head in virtually all Ponerinae and are either triangular or subtriangular in most genera $(N=29)$. Other mandibular shapes with little intrageneric variation are the oblique (Boloponera, Buniapone, Dinoponera, Iroponera, Plectroctena, and Promyopias), pitchfork-like (Belonopelta, Emeryopone, Thaumatomyrmex), elongate sub-oblique (Streblognathus), and scythe-shaped (Harpegnathos). The shape of the mandibles varies from triangular to elongate-triangular in Centromyrmex; from triangular to subtriangular, to oblique in Cryptopone; from subtriangular to oblique, to falcate, to bizarre forms in-between in Leptogenys and Myopias; from sub-oblique to falcate in Psalidomyrmex; and from subtriangular to oblique in Simopelta. Anochetus and Odontomachus are the only ponerines in which the mandibles insert near the midline of the anterior margin of the head.
2. The mandible of Corrieopone is completely edentate (i.e., devoid of any teeth, denticle, or projected apex). To our knowledge, this condition is virtually absent in other Ponerinae, apart from some species of Leptogenys (see Arimoto and Yamane 2018) and Platythyrea (see Fisher and Bolton 2016).
3. Ponerine may present mandibles ornamented with pits and sulci, which are relatively good diagnostic characters to genera.
3.1. The basolateral pit (= basal pit in Fisher and Bolton 2016) is a round to oblong impression on the basal portion of the lateral face of the mandible. It occurs in most Brachyponera species (excluding species from Borneo, Bali, Krakatau, and Sumatra, according to Yamane 2007), Cryptopone [except for C. guianensis and, based on its original description, C. mirabilis (Mackay \& Mackay), as far as we know], and Euponera (see Schmidt and Shattuck 2014). The pit is also present in Fisheropone ambigua (Fig. 45A, B), which disagrees with Schmidt and Shattuck (2014) and Fisher and Bolton (2016).
3.2. The dorsal pit resembles the basolateral pit, although more elongated and impressed on the dorsal face of the mandible. In Ponerinae, it is only present in Dolioponera, Euponera fossigera Mayr (see Mayr 1901), Hagensia, and Iroponera (Fig. 45C; contrary to Schmidt and Shattuck 2014).
3.3. The dorsolateral sulcus runs obliquely along the mandible, from the basal portion of the dorsal face towards the lateral face. It is widespread among the Ponerinae and may be shallowly or deeply impressed, restricted to the basal portion of the mandible, or present along almost the entire lateral face of the mandible. The sulcus is consistently present in species of Asphinctopone, Boloponera, Buniapone, Centromyrmex, most (perhaps all) Ectomomyrmex, Feroponera, Loboponera, Myopias, Odontoponera, Paltothyreus, Phrynoponera, Plectroctena, Promyopias, Psalidomyrmex, Pseudoponera, and Streblogna-


Figure 45. A left mandible of Fisheropone ambigua in dorsal view; worker (CASENT0906209); arrow indicates the basolateral pit B left mandible of Fisheropone ambigua in postdorsolateral view (CASENT0906209); arrow indicates the basolateral pit $\mathbf{C}$ anterior part of the head of Iroponera odax in dorsal view; worker (ANTWEB1008537); arrow indicates the dorsal pit D head of Rasopone panamensis in profile; worker (CASENT0644543); arrow indicates the dorsolateral sulcus $\mathbf{E}$ anterior portion of the clypeus and posterior portion of the mandible of Austroponera castanea in dorsolateral view; worker (CASENT0097796); arrow indicates the short and shallow dorsolateral sulcus $\mathbf{F}$ the expanded maxillolabial complex of Dolioponera fustigera in ventral view; worker (CASENT0411307); Roman numerals indicate the count of maxillary and labial palpomeres $\mathbf{G}$ apical portion of the outer face of the left maxilla of Fisheropone ambigua (CASENT0906209); Roman numerals indicate the count of maxillary palpomeres $\mathbf{H}$ outer face of the labium of Fisheropone ambigua (CASENT0906209); Roman numerals indicate the count of labial palpomeres I apical portion of the outer face of the right maxilla of Loboponera obeliscata; worker (ANTWEB1008545); Roman numerals indicate the count of maxillary palpomeres $\boldsymbol{J}$ outer face of the labium and left maxilla of Loboponera obeliscata (ANTWEB1008545); Roman numerals indicate the count of labial palpomeres $\mathbf{K}$ apical portion of the outer face of the left maxilla of Boloponera ikemkha; paratype, ergatoid queen (CASENT0254321); Roman numerals indicate the count of maxillary palpomeres $\mathbf{L}$ outer face of the labium and right maxilla of Boloponera ikemkha (CASENT0254321); Roman numerals indicate the count of labial palpomeres. Images by FA Esteves ( $\mathbf{A}, \mathbf{B}, \mathbf{E}-\mathbf{H}, \mathbf{K}, \mathbf{L})$, RA $\operatorname{Keller}(\mathbf{C}, \mathbf{I}, \mathbf{J})$, and JT Longino (D); available at AntWeb.org. Scale bars: $0.02 \mathrm{~mm}(\mathbf{A}-\mathbf{C}, \mathbf{F}, \mathbf{G}, \mathbf{I}, \mathbf{J}-\mathbf{L}) ; 0.2 \mathrm{~mm}(\mathbf{D}, \mathbf{E}) ; 0.01 \mathrm{~mm}(\mathbf{H})$.
thus (see Suppl. material 3: Table S3; Schmidt and Shattuck 2014; Fisher and Bolton 2016). On the other hand, the presence of this character varies among species of other genera, such as Bothroponera (present in B. crassa, absent in B. pachyderma), Dinoponera (weakly present in D. lucida; absent in D. longipes), Leptogenys (see Bolton 1975), Mesoponera (only present in M. subiridescens), Neoponera [present in N. fauveli (Emery), weakly impressed in N. apicalis, absent in N. fisheri], Platythyrea (present in P. punctata, absent in P. turneri), and Pseudoneoponera (present in P. porcata, absent in P. denticulata). Contrary to Schmidt and Shattuck (2014), the sulcus is distinct in all Rasopone species examined here (Fig. 45D); although short and constrained to the basal region of the mandibles, it is also present in Austroponera castanea (Fig. 45E). Among the Pachycondyla species examined, the sulcus is a shallow and short basal impression save in P. lenis, where it is absent. In Euponera sikorae (and in all other Malagasy species, according to Rakotonirina and Fisher 2013), the lateral face of the mandible bears a longitudinal sulcus that runs apicad from the lateral margin of the dorsal mandibular articulation. Whether this sulcus is present in Euponera species occurring in other bioregions remains unknown, but it is absent in at least two Afrotropical species, E. brunoi and E. sjostedti.
3.4. Plectroctena species present a sulcus that skirts the mandibular masticatory margin dorsally (Fisher and Bolton 2016), which we refer to as the dorsomasticatory sulcus.

Among the specimens examined, the following taxa present mandibles devoid of any pit or sulcus, like Corrieopone: Anochetus angolensis, A. emarginatus, Belonopelta deletrix, Bothroponera cariosa, B. pachyderma, B. talpa, Cryptopone guianensis, Diacamma ceylonense, Dinoponera longipes, Emeryopone buttelreepeni, Harpegnathos saltator, Hypoponera punctatissima, Mayaponera, Megaponera analis, Mesoponera ambigua, M. australis, M. caffraria, M. elisae rotundata, M. melanaria macra, M. papuana, M. rubra, Neoponera commutata, N. fisheri, N. laevigata, N. luteola, N. verenae, N. villosa, Odontomachus bauri, Ophthalmopone berthoudi, Pachycondyla lenis, Parvaponera darwinii madecassa, Platythyrea turneri, Ponera alpha, P. pennsylvanica, Simopelta oculata, S. transversa, Thaumatomyrmex fraxini, and T. zeteki.
6. We assessed the size of the torular lobes across taxa examined according to the degree of connection between median and lateral arches of torulus (as in Keller 2011, character 08). Among material examined, the lobes are anteriorly and posteriorly continuous with the lateral torular arches in taxa whose antennal sockets are largely exposed in full-face view (Belonopelta, Leptogenys, and Ophthalmopone). On the other extreme, hypertrophied lobes are anteriorly and posteriorly discontinuous with the lateral arches of the torulus (as in Boloponera, Bothroponera sensu stricto group, Loboponera, Platythyrea punctata, Plectroctena, and Psalidomyrmex). Like Corrieopone, most ponerines present intermediate-sized torular lobes that are anteriorly continuous and posteriorly discontinuous with respective lateral arches. In this latter state, the lobes may conceal entirely or partially the lateral arches of the torulus.

For the record, the torular lobes in Bothroponera sulcata species-group members (sensu Schmidt and Shattuck 2014) resemble those of Corrieopone, except for B. henryi (Donisthorpe) (see specimen CASENT0902482) and B. zumpti Santschi (see CASENT0922370), where the lobes are hypertrophied as in the Bothroponera sensu stricto group.
9. Ocelli are invariably present on Harpegnathos workers; it is absent in the worker caste of other Ponerinae, apart from occasional workers.
11. A count of four maxillary and four labial palpomeres is consistently present in Buniapone, Dinoponera, Hagensia, Harpegnathos, Mayaponera (note that we did not examine M. longidentata), Megaponera, Odontoponera, Ophthalmopone, Paltothyreus, Phrynoponera, Promyopias, Streblognathus (see Suppl. material 3: Table S3; Bolton 2003; Fisher and Bolton 2016). We are hesitant to affirm that this is also the case in Neoponera, Pachycondyla, Pseudoneoponera, and Rasopone. These characters usually have been neglected in taxonomic reviews and descriptions of new species, and we did not examine every species in these genera. Yet, in every species we did examine (Neoponera aenescens, $N$. apicalis, $N$. bugabensis, $N$. carinulata, $N$. commutata, $N$. crenata, $N$. curiosa, $N$. dismarginata, $N$. fisheri, $N$. foetida, $N$. globularia, $N$. insignis, $N$. inversa, N. laevigata, N. luteola, N. moesta, N. obscuricornis, N. schoedli, N. striadinodis, N. unidentata, N. verenae, N. villosa, Pachycondyla crassinoda, P. harpax, P. impressa, P. lattkei, P. lenis, P. procidua, P. striata, Pseudoneoponera porcata, P. tridentata, Rasopone costaricensis, R. cryptergates, R. cubitalis, R. guatemalensis, R. panamensis, R. pluviselva, and $R$. politognatha), the palpal formula was 4,4 . The count is also 4,4 in some species of genera with variable palpal formulae, such as Anochetus, Bothroponera, Leptogenys, Mesoponera, and Myopias (see Suppl. material 3: Table S3; Willey and Brown 1983; Bolton 2003; Fisher and Bolton 2016).

While conducting this study, we noticed that the palpal formula of some taxa was incorrectly reported or missing in the pertinent literature. Thus we correct or update the record here. Contrary to Fisher and Bolton (2016), the palpal formula in Dolioponera fustigera is 3,3 (not 2,2, as previously stated; Fig. 45F), and 3,3 in Fisheropone ambigua (not 3,2; Fig. 45G, H). Contrary to Bolton (2003), Keller (2011), and Fisher and Bolton (2016), the palpal formula in Loboponera obeliscata is 2,3 (not 2,2; Fig. 45I, $J$ ); the formula in L. vigilans was correctly reported as 2,2 by these authors (verified on specimen CASENT0003102). We also report for the first time the palpal formula in Boloponera ikemkha (2,3; Fig. 45K, L), Cryptopone hartwigi (3,3; Fig. 46A, B), and Simopelta transversa (2,2; Fig. 46C). The specimen of Thaumatomyrmex atrox examined by Keller (2011) was later determined to be T. fraxini by D'Esquivel et al. (2017) note that the specimen is not in the list of material examined by the latter authors, but some SEM images taken by Keller (2011; specimen ANTWEB1008597) were used to illustrate the new taxon description. One detail not mentioned by D'Esquivel et al. (2017) was the palpal formula of T. fraxini, which is 3,3 (Fig. 46D; previously reported for T. atrox by Keller 2011). Finally, although Fisher and Bolton (2016) stated that the palpal formula in Parvaponera darwinii madecassa is unknown, Brown (1963) provided the correct count, which is 4,3 (Fig. 46E).

12-14. In Corrieopone, the mesonotum is dome-shaped in profile, with a round dorsal margin that is discontinuous with the outline of the pronotum (i.e., it is slightly higher than the pronotum). The promesonotum is much higher than the propodeum, and a deeply impressed metanotal sulcus separates the two. A distinct notopleural suture delimits the mesonotum from the mesopleuron. In dorsal view, the mesonotum is
round. Taxa that bear some resemblance to Corrieopone in this combination of characters are: several Anochetus species (e.g., A. altisquamis Mayr, specimen CASENT0915154; A. armstrongi McAreavey, CASENT0902449; A. brevis Brown, CASENT0902439), Asphinctopone, Austroponera [except A. rufonigra (Clark), CASENT0249178], Brachyponera, Euponera sikorae, Fisheropone ambigua, Hagensia, some Hypoponera [e.g., H. foreli (Mayr), CASENT0173714; H. herbertonensis (Forel), CASENT0907320; H. mesoponeroides (Radchenko), CASENT0917250], some Leptogenys (e.g., L. borivava Rakotonirina \& Fisher, CASENT0430091; L. ixta, L. peruana, L. sonora), Mayaponera, Megaponera analis (dome-shaped mesonotum in larger specimens, as CASENT0781129), most Mesoponera (e.g., M. ambigua, M. australis, M. caffraria, M. elisae rotundata, M. melanaria, M. papuana, M. rubra, M. subiridescens), several Myopias [e.g., M. castaneicola (Donisthorpe), CASENT0902520; M. chapmani Willey \& Brown, CASENT0902533; M. latinoda (Emery), CASENT0270592], several Neoponera (e.g., N. apicalis, N. aenescens, N. fisheri, N. schoedli, N. villosa), some Odontomachus (e.g., O. bauri; O. laticeps Roger, CASENT0904008; O. spissus Kempf, CASENT0281868), Odontoponera transversa, Ophthalmopone, Rasopone rupinicola, R. cubitalis, and Streblognathus.
16. The spiracular lobe is present in most ponerine genera; it is present in Boloponera ikemkha (CASENT0254321) and B. vicans (CASENT0401737), contrary to Fisher and Bolton (2016).

The lobe is absent in Dolioponera, Fisheropone, some Loboponera species, the Afrotropical and Malagasy Hypoponera, H. punctatissima, Simopelta oculata, S. transversa, and Thaumatomyrmex fraxini (Suppl. material 3: Table S3; Fisher and Bolton 2016). Interestingly, while the lobe is absent in the Afrotropical Cryptopone (according to Fisher and Bolton 2016; here confirmed on C. hartwigi), it is present in the Neotropical C. gilva (ANTWEB1008514), C. guianensis (ANTWEB1008565), C. holmgreni (ECOFOG-IT14-0276-07), and C. pauli (CASENT0637806).
18. The metapleural longitudinal flange is a carina that extends along the metapleuron in profile, with its posterior end immediately dorsad the metapleural gland orifice. When well-developed, it projects laterad or ventrolaterad and may overhang the gland orifice (as defined by Keller 2011: character 62). In Simopelta species (ANTWEB1008589), this flange is strongly projected ventrolaterad and overlaps the gland orifice.
19. The propodeal dorsum presents a well-delimited, narrow, median longitudinal groove in Psalidomyrmex (Fisher and Bolton 2016; ANTWEB1008585). In addition, a vestigial longitudinal groove may be present on the propodeal dorsal face of specimens of the Plectroctena minor group (see Bolton and Brown 2002). The propodeal dorsum is transversely concave along its entire length, or only posteriorly, in Hagensia, Mayaponera (CASENT0249137), several Mesoponera (CASENT0249194), and Pseudoponera (CASENT0923115).
20. Most ponerine genera present an unarmed propodeum. In profile, the propodeal dorsoposterior corner bears acute projections in several Anochetus species (e.g., CASENT0902431, CASENT0815182, CASENT0746783), Phrynoponera (CASENT0178230), Streblognathus (ANTWEB1008591), some Platythyrea (CASENT0281867, CASENT0900569, CASENT0903799), and Pseu-


Figure 46. A outer face of the left maxilla of Cryptopone hartwigi; worker (CASENT0251956); Roman numerals indicate the count of maxillary palpomeres $\mathbf{B}$ outer face of the labium of Cryptopone hartwigi (CASENT0251956); Roman numerals indicate the count of labial palpomeres $\mathbf{C}$ expanded maxillolabial complex of Simopelta transversa in ventral view; worker (ANTWEB1008589); Roman numerals indicate the count of maxillary and labial palpomeres $\mathbf{D}$ maxillolabial complex of Thaumatomyrmex fraxini in ventroapical view; worker (ANTWEB1008597); Roman numerals indicate the count of maxillary and labial palpomeres E expanded maxillolabial complex of Parvaponera darwinii madecassa in ventral view; worker (CASENT0389498); Roman numerals indicate the count of maxillary and labial palpomeres $\mathbf{F}$ right metacoxal cavity of Platythyrea cribrinodis in ventral view; worker (CASENT0778160); arrowhead indicates the fused annular cuticle encircling the cavity externally $\mathbf{G}$ right metacoxal cavity of Myopias darioi in ventral view; paratype, worker (CASENT0810080); arrowhead indicates the fused annular cuticle encircling the cavity externally $\mathbf{H}$ right metacoxal cavity of Phrynoponera pulchella in ventral view; worker (CASENT0217034); arrowhead indicates the annular gap around the cavity $\mathbf{F}$ protibial apex and basal region of the probasitarsus of Loboponera obeliscata in posterior view; worker (ANTWEB1008545); arrowhead indicates the minute basoventral lamella of the calcar. Images by FA Esteves $(\mathbf{A}, \mathbf{B}, \mathbf{E}-\mathbf{H})$ and RA Keller (C, D, I); available at AntWeb.org. Scale bars: $0.02 \mathrm{~mm}(\mathbf{A}, \mathbf{B}, \mathbf{D}) ; 0.03 \mathrm{~mm}(\mathbf{C}) ; 0.04 \mathrm{~mm}(\mathbf{E}-\mathbf{I})$.
doneoponera bispinosa (Smith). Also, some species of the Loboponera vigilans group (CASENT0003111), Plectroctena minor group (CASENT0915285), and P. mandibularis group (CASENT0102947) present the lateral margin of the propodeal declivity with a lamella that is dorsally toothed; in other species of the L. vigilans group, the lateral margin of the propodeal declivity is toothed dorsally, but the lamella is absent (CASENT0003098; see Bolton and Brown 2002).
22. In general, the shape of the propodeal spiracle is constant within genus in Ponerinae. A slit-shaped spiracle (i.e., external atrial opening $>2 \times$ longer than wide; as in Keller 2011, character 65) is present in Asphinctopone, Austroponera, Bothroponera, Buniapone, Corrieopone, Diacamma, Dinoponera, Ectomomyrmex, Euponera, Feroponera, Fisheropone, Hagensia, Harpegnathos, Megaponera, Odontoponera, Ophthalmopone, Pachycondyla, Paltothyreus, Phrynoponera, Pseudoneoponera, and Streblognathus. A round to oval spiracle occurs in Belonopelta, Boloponera, Dolioponera, Emeryopone, Hypoponera, Iroponera, Loboponera, Mayaponera, Myopias, Odontomachus, Plectroctena, Ponera, Promyopias, Psalidomyrmex, Rasopone, Simopelta, and Thaumatomyrmex. The propodeal spiracle is round to oval in most Brachyponera species but varies from oval to slit-shaped in B. atrata and B. sennaarensis.
24. We classified the metacoxal cavities as closed or open by integrating the definition given by Keller (2011: character 69) with that in Fisher and Bolton (2016; see also Bolton 2003: character 41). If closed, the medial surface of the metacoxal acetabulum does not have a fenestra, and thus, the metacoxal cavity is not connected internally with the propodeal foramen; the annulus externally encircling the cavity is fused. If open, the internal medial surface of the cavity is fenestrate and connects with the propodeal foramen, and the annulus is unfused. In this case, the annulus may encircle the cavity with its free ends overlapping next to the propodeal foramen; or there may be a gap in the annulus.

An unfused cuticular annulus tightly encircles the metacoxal cavities in most Ponerinae we dissected. This condition occurs in Phrynoponera pulchella specimens from Kenya (CASENT0178203, CASENT0178204, CASENT0217125), which is in accord with Bolton and Fisher (2008a). However, a specimen from Tanzania possesses an annular gap in both metacoxal cavities (Fig. 46H); we are uncertain if those were natural or dissection artifacts, as unfortunately, only one specimen from that population was available to us. Metacoxal cavities also present an annular gap in Phrynoponera transversa and Platythyrea punctata. On the other hand, the annulus is fused and uninterrupted in Harpegnathos saltator, Platythyrea cribrinodis (Fig. 46F; contrary to Fisher and Bolton 2016), and Myopias darioi (Fig. 46G).
25. The calcar of strigil presents a basoventral lamella in most Ponerinae evaluated (as in Keller 2011, character 74). For clarification, we consider the calcar to present a small lamella in Loboponera obeliscata (Fig. 46I) and that it is entirely pectinate in Platythyrea punctata and P. turneri (specimens ANTWEB1008574 and ANTWEB1008575, respectively), which is contrary to Keller (2011). The lamella is also absent in Boloponera, Brachyponera sennaarensis, Diacamma ceylonense, Dolioponera fustigera, Emeryopone buttelreepeni, Harpegnathos saltator, Hypoponera punctatissima, Leptogenys ixta, L. peruana, L. pucuna, L. sonora, L. wheeleri, Loboponera vigilans, Mesoponera ambigua, M. elisae
rotundata, Myopias darioi, Platythyrea cribrinodis, Ponera alpha, P. pennsylvanica, Promyopias silvestrii, Simopelta oculata, S. transversa, Thaumatomyrmex fraxini, and T. zeteki.
27. A row of stout, spine-like setae occurs along the posterior face of probasitarsal notch, parallel to the comb of strigil, in most Ponerinae taxa examined. We adopted the definition of "row" analogous to that of a line, whose existence requires at least two points in space. Thus, the row is present if two or more spine-like setae are aligned longitudinally along the posterior face of the probasitarsal notch; less than two setae make the row absent. Among material examined, the row is absent on the posterior face of the probasitarsal notch in Dolioponera fustigera, Leptogenys, Myopias darioi, and Thaumatomyrmex fraxini.
28. Most Ponerinae taxa present two mesotibial spurs. Among taxa examined, the anterior spur is simple, and the posterior spur is serrate in Brachyponera chinensis, B. croceicornis, B. lutea, B. luteipes, B. obscurans, Dinoponera longipes, D. lucida, Hagensia havilandi marleyi, Harpegnathos saltator, Leptogenys peruana, L. pucuna, and Ophthalmopone berthoudi.

For the record, only one mesotibial spur is visible under a stereomicroscope in Asphinctopone silvestrii and Fisheropone ambigua; however, SEM images reveal that a vestigial anterior spur is present in both taxa (Fig. 47C, E); A. differens also seems to bear a minute anterior spur on the mesotibia.
29. The metatibial gland has been confirmed in Bothroponera tesseronoda (Emery), several species of Diacamma Mayr, Harpegnathos saltator, Neoponera crenata, N. marginata (Roger), Paltothyreus tarsatus, Pseudoneoponera rufipes (Jerdon), and P. tridentata (see Hölldobler et al. 1996; Billen 2009). The gland is associated with a cuticular pore plate, which may be covered by a brush of stouter, distinctly-shaped setae (as in Diacamma and Paltothyreus tarsatus), or it may be an oblong, glabrous, distinctly colored, smooth, or distinctly sculptured cuticular patch (Hölldobler et al. 1996). Here, we list taxa that present the latter condition, which is what is seen in Corrieopone. Among the taxa examined, the cuticular patch is present on the apicoposterior face of the metatibia in Bothroponera crassa, B. silvestrii, Brachyponera croceicornis, B. lutea, B. luteipes, B. obscurans, B. sennaarensis, Cryptopone hartwigi, Ectomomyrmex javanus, Emeryopone buttelreepeni, Euponera sikorae, E. sjostedti, Feroponera ferox, Hagensia havilandi marleyi, Mesoponera caffraria, Pseudoneoponera porcata, and P. tridentata.
30. Most Ponerinae taxa present two metatibial spurs. Here, all specimens examined present a pectinate posterior spur, and in the majority, the anterior spur is simple, as in Corrieopone (see Suppl. material 3: Table S3).

For the record, Schmidt and Shattuck (2014: 185) and Fernandes and Delabie (2019: 411) stated that Cryptopone species present one metatibial spur (excluding C. guianensis, and in the case of the latter authors, also C. pauli). Those assertions are puzzling, as several Cryptopone species have two spurs on the metatibia [e.g., C. arabica Collingwood \& Agosti, C. gilva, C. holmgreni (Wheeler), C. ochracea Mayr; see Fig. 47A; Kempf 1961b; Bernard 1967; Collingwood and Agosti 1996]. Belonopelta deletrix has two spurs on the metatibia (Fig. 47B), as correctly stated by Baroni Urbani (1975), but contrary to Schmidt and Shattuck (2014). Asphinctopone silvestrii presents


Figure 47. A apex of the metatibia and basal region of the metabasitarsus of Cryptopone gilva in anterior view; worker (ANTWEB1008514); arrows highlight the metatibial spurs B apex of the metatibia and basal region of the metabasitarsus of Belonopelta deletrix in anterior view; worker (ANTWEB1008507); arrows highlight the metatibial spurs $\mathbf{C}$ ventroapical portion of the mesotibia of Asphinctopone silvestrii in anteroventral view; worker (CASENT0824505); arrows highlight the minute anterior and much larger posterior spurs $\mathbf{D}$ ventroapical portion of the metatibia of Asphinctopone silvestrii in anteroventral view (CASENT0824505); arrows highlight the minute anterior and much larger posterior spurs $\mathbf{E}$ ventroapical portion of the mesotibia of Fisheropone ambigua in ventral view; worker (CASENT0906216); arrows highlight the vestigial anterior and well-developed posterior spurs $\mathbf{F}$ ventroapical portion of the metatibia of Fisheropone ambigua in oblique anteroventral view (CASENT0906216); arrows highlight a fovea at the place the anterior spur would be located and the large posterior spur. Images by RA Keller (A, B) and FA Esteves (C-F); available at AntWeb.org. Scale bars: $0.03 \mathrm{~mm}(\mathbf{A}, \mathbf{B}) ; 0.004 \mathrm{~mm}(\mathbf{C}-\mathbf{F})$.
a minute additional metatibial spur (Fig. 47D), anterior to the much larger pectinate spur; A. differens seems to present the same. The metatibia of Fisheropone ambigua, which bears a single spur, long and pectinate, also presents a fovea where the anterior spur would be located (Fig. 47F).
31. Taxa without stout, spine-like setae on the dorsal face of the mid- and hindlegs are Anochetus angolensis, A. emarginatus, Asphinctopone differens, As. silvestrii, Belonopelta deletrix, Boloponera ikemkha, B. vicans, Bothroponera silvestrii, Brachyponera chinensis, B. croceicornis, B. luteipes, B. obscurans, B. sennaarensis, Diacamma ceylonense, Dolioponera fustigera, Emeryopone buttelreepeni, Euponera sikorae, E. sjostedti, Hagensia havilandi marleyi, Harpegnathos saltator, Iroponera odax, Leptogenys ixta, L. peruana, L. pucuna, L. sonora, L. wheeleri, Loboponera obeliscata, L. vigilans, Mayaponera conicula, Myopias darioi, M. maligna, Neoponera bugabensis, $N$. carinulata, $N$. cavinodis, $N$. crenata, $N$. dismarginata, $N$. fiebrigi cf., N. fisheri, N. foetida, N. globularia, N. insignis, N. inversa, N. luteola, N. moesta, N. obscuricornis, N. striadinodis, N. unidentata, N. villosa, Odontomachus bauri,

Odontoponera transversa, Ophthalmopone berthoudi, Platythyrea cribrinodis, P. punctata, P. turneri, Simopelta oculata, S. transversa, Thaumatomyrmex fraxini, and T. zeteki.

Contrary to Fisher and Bolton (2016), Fisheropone ambigua presents stout, spinelike setae on the dorsal face of the mesobasitarsus (Fig. 48A).
33. We categorized the shape of pro-, meso-, and metapretarsal claws according to the presence, location, and number of acute projections on their inner margins (modified from Keller 2011, character 82). A simple pretarsal claw is devoid of prominences (Fig. 48B) or presents a blunt basal angle or rounded swell (Fig. 48C), and is found in the following taxa: Anochetus angolensis, A. emarginatus, Asphinctopone differens, A. silvestrii, Austroponera castanea, Belonopelta deletrix, Boloponera ikemkha, B. vicans, Bothroponera crassa, B. silvestrii, Brachyponera chinensis, B. croceicornis, B. lutea, B. luteipes, B. obscurans, B. sennaarensis, Centromyrmex brachycola, C. decessor, C. ereptor, C. raptor, Cryptopone gilva, C. guianensis, C. hartwigi, Diacamma ceylonense, Dolioponera fustigera, Ectomomyrmex javanus, Emeryopone buttelreepeni, Euponera brunoi, E. sikorae, E. sjostedti, Feroponera ferox, Fisheropone ambigua, Hypoponera punctatissima, Iroponera odax, Loboponera obeliscata, L. vigilans, Mayaponera becculata, M. cernua, M. conicula, M. constricta, Mesoponera ambigua, M. australis, M. caffraria, M. elisae rotundata, M. melanaria macra, M. papuana, M. rubra, M. subiridescens, Myopias darioi, M. maligna, Neoponera aenescens, $N$. apicalis, N. bugabensis, N. cavinodis, N. crenata, N. dismarginata, N. eleonorae, N. fiebrigi cf., N. fisheri, N. foetida, N. globularia, N. insignis, N. inversa, N. laevigata, N. luteola, N. obscuricornis, N. schoedli, N. striadinodis, $N$. unidentata, $N$. verenae, N. villosa, Odontomachus bauri, Odontoponera transversa, Pachycondyla lattkei, Parvaponera darwinii madecassa, Plectroctena strigosa, Ponera alpha, P. pennsylvanica, Psalidomyrmex procerus, Pseudoneoponera porcata, Pseudoponera gilberti, P. stigma, Rasopone costaricensis, R. cryptergates, R. cubitalis, R. guatemalensis, R. panamensis, R. pluviselva, R. politognatha, Simopelta oculata, S. transversa, and Streblognathus peetersi.

A claw with a basal tooth bears an acute prominence on the basal third of its inner margin (Fig. 12B). The basal tooth overhangs the outline of the inner margin of the claw and may bear several small, acute projections (Fig. 48D). This type of claw occurs in Bothroponera cariosa, B. pachyderma, B. talpa, Mayaponera arbuaca, M. pergandei, Megaponera analis, Neoponera carinulata, N. commutata, N. moesta, Ophthalmopone berthoudi, Pachycondyla crassinoda, P. harpax, P. impressa, P. lenis, P. procidua, P. striata, Phrynoponera pulchella, P. transversa, and Pseudoneoponera tridentata. A claw with a preapical tooth has an acute projection rising from the apical two-thirds of its inner margin (Fig. 48E). It is present in Dinoponera longipes, D. lucida, Hagensia havilandi marleyi, Harpegnathos saltator, Paltothyreus tarsatus, Platythyrea cribrinodis, P. punctata, P. turneri, Thaumatomyrmex fraxini, and T. zeteki. Pectinate pretarsal claws are shaped like a comb and are unique to Leptogenys among Ponerinae (Fig. 48F).

Finally, we found shape variations among the claws of the fore-, mid-, and hindlegs only in Buniapone amblyops and Promyopias silvestrii. These taxa present a propretarsal claw with a long basal tooth, while their meso- and metapretarsi claws are simple (Fig. 48G-I). This condition contradicts Bolton and Fisher (2008b) and Fisher and Bolton (2016), who stated the claws are simple in P. silvestrii.


Figure 48. A mesobasitarsus of Fisheropone ambigua in anterior view; worker (CASENT0906209); arrows indicate the stout, spine-like setae on the dorsal face of the tarsomere B pretarsus of Mayaponera constricta in ventral view; worker (CASENT0260254) C pretarsus of Simopelta transversa in oblique apicolateral view; worker (ANTWEB1008589) D pretarsus of Bothroponera pachyderma in oblique dorsal view; worker (ANTWEB1008567) E pretarsus of Hagensia havilandi marleyi in apicolateral view; worker (ANTWEB1008566) F pretarsus of Leptogenys wheeleri in ventral view; worker (ANTWEB1008541) G propretarsus of Promyopias silvestrii in dorsolateral view; worker (CASENT00178751) H metapretarsus of Promyopias silvestrii in apical view (CASENT00178751) I propretarsus of Buniapone amblyops in apicolateral view; worker (CASENT0384973); insert: metapretarsus of Buniapone amblyops in dorsoapical view (CASENT0384973). Images by FA Esteves (A, B, G-I) and RA Keller (C-F); available at AntWeb. org. Scale bars: $0.04 \mathrm{~mm}(\mathbf{A}, \mathbf{B}, \mathbf{F}) ; 0.02 \mathrm{~mm}(\mathbf{C}-\mathbf{E}, \mathbf{G}-\mathbf{I})$.
34. The arolium was absent or reduced to a membranous cuticular flap between the pretarsal claws of most taxa examined (Figs 12B, 48D-I). Exceptions were Belonopelta deletrix, Diacamma ceylonense, Harpegnathos saltator, Iroponera odax, Mayaponera, Neoponera, Parvaponera darwinii madecassa, Platythyrea cribrinodis, P. punctata, P. turneri, Rasopone guatemalensis, Simopelta oculata, S. transversa (Fig. 48C), Thaumatomyrmex fraxini, and T. zeteki.

Contrary to Schmidt and Shattuck (2014), we consider that Mayaponera constricta has indistinct arolia like all its congeners (Fig. 48B). In this species, the arolium is reduced to a cuticular flap, although more developed than what we classified as indistinct in other taxa. However, this feature was not noticeable under our stereo microscope.
35. The petiolar node of Corrieopone lacks a spine-like or any other acute projection and is narrow in profile, with its anterior and posterior surfaces tapering to an insignificant dorsal surface.

Ponerine taxa with an unarmed scale-like petiole are: several Anochetus (see CASENT0915154); Asphinctopone(CASENT0915481); Austroponera (FOCOL0965); Brachyponera (CASENT0915660); Buniapone (CASENT0903944); some Cryptopone (ANTWEB1008000); some Ectomomyrmex (CASENT0907270); Euponera fossigera species group [viz.: E. brunoi, E. fossigera Mayr (SAM-HYM-C002649B), E. malayana (Wheeler), E. sharpi Forel, E. wroughtonii Forel, E. wroughtonii crudelis Forel, and probably also E. sakishimensis (Terayama)]; Fisheropone (CASENT0906215); Hagensia (CASENT0256487); several Hypoponera (CASENT0281911); some Leptogenys (CASENT0902609); Mayaponera (USNMENT00442104); Mesoponera (CASENT0249169); some Neoponera (ANTWEB1014009); very few Odontomachus (CASENT0281868); very few Pachycondyla (UFV-LABECOL-000002); some Parvaponera (CASENT0915276); some Ponera (CASENT0235336); and Pseudoponera [CASENT0902509; except P. pachynoda (Clark), ANTWEB1008183].
36. The strigation on the posteroventral portion of the petiolar tergite of Corrieopone resembles that of Asphinctopone (CASENT0178221).
39. The spatulate projection rises from the posterior portion of the petiolar sternite and extends posteriad, overlapping either partially or entirely the remaining sternite. This definition departs from Keller (2011, character 111) in not requiring (a) the projection to extend beyond the posterior margin of the petiolar sternite and (b) close proximity between projection and remaining sternite. The reason is the character varies continuously, which invalidates both requirements. The variation comprises projections that are long and conceal the helcial sternite almost completely to partially (as in Phrynoponera pulchella, specimen ANTWEB1008573, and Platythyrea punctata, respectively; Fig. 49A); those that reach or almost reach the posterior margin of the petiolar sternite (as in Platythyrea turneri, Fig. 49B); and those even shorter (as in Asphinctopone silvestrii; Fig. 49C). In addition, projections may tightly envelop the remaining sternite (as in Phrynoponera pulchella and Platythyrea punctata, ANTWEB1008574; Fig. 49D); or a slight gap may be present [as in Platythyrea turneri, specimen ANTWEB1008575, and Phrynoponera gabonensis (André); Fig. 49E]; or the gap may be slightly broader (as in Rasopone jtl030; Fig. 49F) to much broader (as in Streblognathus peetersi; Fig. 49G).

Thus, according to our definition, the spatulate projection of the petiolar sternite is present in Austroponera, Asphinctopone, Brachyponera, Megaponera analis, Ophthalmopone, Phrynoponera, Platythyrea, Rasopone, and Streblognathus; see also Fisher and Bolton (2016) for the description of the sternite in some of these taxa. The petiolar sternite in Belonopelta deletrix, immediately anterior to its posterior margin, is pro-
jected ventrad and slightly posteriad; the same seems to happen in Thaumatomyrmex fraxini. According to the species redescription given by Mackay and Mackay (2010), Neoponera magnifica may also present the character.
40. Like Corrieopone, most ponerine genera present an infra-axial helcium (i.e., positioned ventrad the midheight of the anterior face of abdominal segment III; see Keller 2011, character 114). A few taxa examined present an axial helcium (i.e., positioned at midheight of the anterior face of abdominal segment III): Boloponera, Buniapone, Centromyrmex, Cryptopone (except C. guianensis), Dolioponera, Feroponera, Iroponera, Platythyrea, and Promyopias.
41. As far as we know, the prora is present and well-developed in most Ponerinae. It is usually indistinct in Platythyrea, but contrary to Fisher and Bolton (2016), it is well-developed in some species [as in P. lamellosa (Roger); Fig. 49H], and weakly projected but still visible in others [as in P. pilosula (Smith); Fig. 49I]. According to Schmidt and Shattuck (2014), the prora is absent in Iroponera, but SEM images of I. odax reveal that it is present, albeit weakly projected (Fig. 49J). According to Fisher and Bolton (2016), the trait is absent in Dolioponera, which agrees with what we saw in most specimens. However, SEM images of specimen ANTWEB1008521 show a weak projection on the anterior face of abdominal sternite III; we are unsure whether that detail is natural or an image artifact.

We consider the prora present in Mayaponera and Rasopone, in disagreement with Longino and Branstetter (2020). The trait, although small, is distinct in the profile view of $M$. becculata, $M$. conicula, $M$. constricta, and $R$. panamensis. In the first two species and $R$. panamensis, the prora rises from the anterior portion of the abdominal poststernite III (see specimens CASENT0249130, CASENT0644252). In M. constricta, it projects from the area in between the ventral margins of the helcial tergite arch and the anterior portion of the abdominal poststernite III (Fig. 49K) and resembles the condition seen in Dinoponera, Pachycondyla, and Streblognathus (ANTWEB1014000, CASENT0249148, and Fig. 49G, respectively). In M. arhuaca, M. cernua, M. pergandei, and other Rasopone species, the prora is indistinct in the profile view of undissected specimens, for it is a minute prominence located in the area between the ventral margins of the helcial tergite (as in Brachyponera, Fig. 49L), and may be fused entirely with it (as described for Phrynoponera by Bolton and Fisher 2008a).
43. We considered the girdling constriction present if the surface between pre- and postsclerites of abdominal segment IV was interrupted by a shallow or deep impression on the integument. A line, if present, only constituted a constriction if the integument was depressed.

Among taxa examined, the constriction is absent in Asphinctopone, Brachyponera sennaarensis (weakly impressed in other species), Corrieopone, Mesoponera (except M. caffraria), Odontomachus, Odontoponera, Simopelta, Streblognathus, and Thaumatomyrmex fraxini.
44. The stridulitrum is consistently present on abdominal pretergite IV in Austroponera, Belonopelta, Dinoponera, Harpegnathos, Megaponera, Neoponera, Odontoponera, Ophthalmopone, Streblognathus, and Thaumatomyrmex. The trait's oc-


Figure 49. A petiole of Platythyrea punctata in ventral view; worker (ANTWEB1008574) B petiole of Platythyrea turneri in ventral view; worker (ANTWEB1008575) C petiole of Asphinctopone silvestrii in ventral view; worker (CASENT0824505) D petiole of Phrynoponera pulchella in profile; worker (ANTWEB1008573) E petiole of Phrynoponera gabonensis in profile; worker (CASENT0250060) F petiole of Rasopone jt1030 in profile; worker (CASENT0633075) G petiole of Streblognathus peetersi in profile; worker (CASENT0258947) H petiole and abdominal segment IV of Platythyrea lamellosa in profile; worker (CASENT0252018); arrow indicates the prora I petiole and abdominal segment IV of Platythyrea pillosula in profile; worker (CASENT0260481); arrows indicate the prora $\mathbf{J}$ petiole and anterior portion of the abdominal segment IV of Iroponera odax in ventral view; worker (ANTWEB1008537); arrow indicates the prora $\mathbf{K}$ petiole and anterior portion of the abdominal segment IV of Mayaponera constricta in profile; worker (CASENT0643469); arrow indicates the prora $\mathbf{L}$ posterior portion of the petiole and the helcium of Brachyponera croceicornis in ventral view; worker (ANTWEB1008564); arrow indicates the minute prora projected from the area in between the ventral margins of the helcium tergite. Images by RA Keller (A, B, D, J, L), FA Esteves ( $\mathbf{C}, \mathbf{K}$ ), B Reynolds ( $\mathbf{E}, \mathbf{H}$ ), JT Longino ( $\mathbf{F}$ ), M Esposito ( $\mathbf{G}$ ), and W Ericson (I); available at AntWeb.org. Scale bars: 0.2 mm .


Figure 50. A abdominal pretergite IV of Brachyponera obscuransin dorsal view; worker (CASENT0059638) B abdominal pretergite IV of Mesoponera subiridescens in dorsal view; worker (CASENT0906219) C stridulitrum of Mesoponera subiridescens in dorsal view (CASENT0906219) D medial area of the abdominal pretergite IV of Euponera sikorae; worker (CASENT0497109) E close-up of the medial area of the abdominal pretergite IV of Euponera sikorae; worker (CASENT0134370) F medial area of the abdominal pretergite IV of Mesoponera caffraria; worker (CASENT0408614) G medial area of the abdominal pretergite IV of Mayaponera pergandei; male (CASENT0317474) $\mathbf{H}$ abdominal pretergite IV of Mayaponera pergandei in dorsal view; worker (CASENT0845437) I close-up of the medial area of the abdominal pretergite IV of Mayaponera pergandei; worker (CASENT0845437). Images by FA Esteves; available at AntWeb.org. Scale bars: $0.06 \mathrm{~mm}(\mathbf{A}, \mathbf{F}, \mathbf{G}, \mathbf{I}) ; 0.09 \mathrm{~mm}(\mathbf{B}) ; 0.04 \mathrm{~mm}(\mathbf{C}) ; 0.2 \mathrm{~mm}(\mathbf{D}, \mathbf{E}) ; 0.08 \mathrm{~mm}(\mathbf{H})$.
currence is variable in Anochetus, Bothroponera (present in members of the sulcata group), Brachyponera, Hypoponera, Mayaponera (only present in M. constricta), Mesoponera, Myopias, Odontomachus, Phrynoponera (only present in P. pulchella), and Ponera (Suppl. material 3: Table S3; see also Markl 1973; Schmidt and Shattuck 2014; Fisher and Bolton 2016). Schmidt and Shattuck (2014) state that it is universally present in Leptogenys and Platythyrea. However, Markl (1973), after a


Figure 5I. Posterior portion of abdominal segment IV in profile (only the hypopygium is seen in image C); in each image, a sample of stout, spine-like seta (or microtrichia, in image I) is highlighted in orange A Brachyponera chinensis; worker (CASENT0104738) B Buniapone amblyops; worker (CASENT0384786) C Mayaponera becculata; worker (CASENT0428720) D Mesoponera melanaria macra; worker (CASENT0159302) E Myopias darioi; worker (CASENT0810080) F Parvaponera darwinii madecassa; worker (CASENT0389498) G Promyopias silvestrii; worker (CASENT00178751) H Rasopone panamensis; worker (CASENT0644252) I Thaumatomyrmex fraxini; worker (ANTWEB1008597). Images by FA Esteves ( $\mathbf{A} \mathbf{- H}$ ) and RA Keller (I); available at AntWeb.org. Scale bars: 0.04 mm .
comprehensive taxa examination, affirmed that the occurrence of the trait is variable among species in both genera.

Contrary to Fisher and Bolton (2016), the stridulitrum is present in Brachyponera obscurans (Fig. 50A) and at least two Afrotropical and Malagasy species of Mesoponera (Fig. 50B, C). For the record, it is absent in Boloponera ikemkba. In addition, the midline of the fourth abdominal pretergite is strigulate and dissimilar to the surrounding sculpture in Euponera sikorae (Fig. 50D, E) and Mesoponera caffraria (Fig. 50F); the ridges are set farther apart and form a narrower area in the latter species. However,
whether they have a stridulatory function in those species is unclear. Finally, the stridulitrum is present in males of Mayaponera pergandei (Fig. 50G; contrary to Mackay and Mackay 2010) but absent in the worker caste (Fig. 50H, I). Assuming that W. P. Mackay correctly determined the two males we examined, this intercaste variation is interesting because (1) it has not been observed in ants (see Markl 1973), and (2) it suggests that this trait has a reproductive function in $M$. pergandei.
47. The hypopygium (abdominal sternite VII) is armed with spine-like setae that flank the sting in some Ponerinae. According to previous studies, the setae are present in Dinoponera, Ophthalmopone, Pachycondyla, Paltothyreus, some Leptogenys, and few Ponera (Schmidt and Shattuck 2014; Fisher and Bolton 2016). However, in Pachycondyla, we found that the hypopygial setae may be spine-like or aristate (Figs 10C-F, 12C, D; see details in the preceding subsection "Transfers between Neoponera to Pachycondyla").

Hypopygial spine-like setae occur in several Brachyponera (Fig. 51A), Buniapone amblyops (Fig. 51B), Mayaponera becculata (Fig. 51C), some Mesoponera (Fig. 51D), Myopias darioi (Fig. 51E), M. maligna, some Neoponera (Fig. 10A, B; see subsection "Transfers between Neoponera to Pachycondyla"), Parvaponera darwinii madecassa (Fig. 51F), Promyopias silvestrii (Fig. 51G), and Rasopone panamensis (Fig. 51H; see also Suppl. material 3: Table S3). In Thaumatomyrmex fraxini and T. zeteki, the hypopygium vestiture is composed of minute spine-like microtrichia (Fig. 51I).

## Comparison with similar genera

Corrieopone gen. nov. is morphologically distinctive and unlikely to be confused with any other Ponerinae genus. To our knowledge, no other Ponerinae has a clypeal medial area that projects anteriorly as a broad, truncated prominence that overlaps the mandibles dorsally and presents a broad anteroventral face, which is subrectangular in ventro-anterior view (Figs 37B, 38A-E, 39A-D). In addition, no other ant presents the ventral face of the hypopygium with a longitudinal concavity that posteriorly bears a longitudinal carina and stout, hook-shaped setae.

Despite these features, workers of Corrieopone superficially resemble Asphinctopone, Brachyponera, some Hagensia, some Mayaponera, and most Mesoponera species, because they share the following characters: the eyes are located on the anterior part of the head; the mesonotum is dome-shaped; the metanotal sulcus is deeply impressed; the notopleural suture is conspicuous; the metathoracic spiracles are concealed by a cuticular lobe; the dorsoposterior area of the propodeum lacks spines or tubercles; the petiole is unarmed and scale-like; the constriction between the presclerites and postsclerites of abdominal segment IV is shallowly impressed or absent (Fig. 52A-F).

However, Corrieopone differs from these genera in many characters (Tables 2, S3), including those of the mandibles (only in Corrieopone it is edentate), clypeus (only Corrieopone has the clypeal anterior projection shaped as described above), and hypopygium (only Corrieopone has a hypopygium as described above). In Corrieopone, (1) the mandible is devoid of a dorsolateral sulcus or basolateral or dorsal pits (the mandible presents a dorsal pit in Hagensia, a basolateral pit in Brachypon-


Figure 52. A Corrieopone nouragues, holotype, worker (CASENT0830464) B Asphinctopone silvestrii; worker (CASENT0406793) C Brachyponera arcuata; syntype, worker (CASENT0916828) D Hagensia peringueyi; worker (CASENT0256487) E Mayaponera constricta; worker (CASENT0249137) F Mesoponera cf. ambigua; worker (CASENT0629612). Images by W Lee (A), A Nobile (B), K Martynova (C), B Reynolds (D), R Perry (E), and JT Longino (F); available at AntWeb.org. Scale bars: 1 mm .
era, and a dorsolateral sulcus in Asphinctopone and Mesoponera subiridescens); (2) the palpal formula is 4,4 (it is 3,3 in Asphinctopone, Brachyponera, and Mesoponera ambigua); (3) the mesopleuron is distinctively divided into anepisternum and katepisternum (it is undivided in Mesoponera, excluding M. subiridescens, and Mayaponera, excluding M. conicula); (4) the propodeal spiracle is slit-shaped (it is round to oval in Brachyponera save B. sennaarensis, Mayaponera, and Mesoponera, excluding

Table 2. Character matrix for ponerine genera that superficially resemble Corrieopone.

| Genus | 2 | 3.1 | 3.2 | 3.3 | 4 | 11 | 15 | 22 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Asphinctopone | 1 | 0 | 0 | 1 | 0 | 3,3 | 1 | 0 |
| Brachyponera | 1 | $1^{\text {a }}$ | 0 | 0 | 0 | 3,3 | 0,1 | $1^{\text {f }}$ |
| Corrieopone | 0 | 0 | 0 | 0 | 1 | 4,4 | 1 | 0 |
| Hagensia | 1 | 0 | 1 | 0 | 0 | 4,4 | 1 | 0 |
| Mayaponera | 1 | 0 | 0 | 0 | 0 | 4,4 | $0^{\text {d }}$ | 1 |
| Mesoponera | 1 | 0 | 0 | $0^{\text {b }}$ | 0 | 3,3; 4,3; 4,4 ${ }^{\text {c }}$ | $0,1{ }^{\text {e }}$ | $1^{8}$ |
| Genus | 29 | 30 | 33 | 39 | 41 | 44 | 46 | 47 |
| Asphinctopone | 0 | 1 | 0 | 1 | 1 | 0 | 0 | 0 |
| Brachyponera | $1^{\text {h }}$ | 2 | 0 | 1 | 2 | 0,1 ${ }^{\text {j }}$ | 0 | 1 |
| Corrieopone | 1 | 2 | 0 | 0 | 1 | 1 | 1 | 0 |
| Hagensia | 1 | 2 | 2 | 0 | 1 | 0 | 0 | 0 |
| Mayaponera | 0 | 2 | 0,1 | 0 | 1,2 | $0^{\text {k }}$ | 0 | $0^{\text {m }}$ |
| Mesoponera | $0^{\text {i }}$ | 2 | 0 | 0 | 1 | $0,1^{1}$ | 0 | $1^{\text {n }}$ |

Character numbers correspond to characters used in genus diagnosis (see genus diagnosis and "Comments on worker characters" for character definition):

2 Mandibles, dentition: (0) edentate, (1) dentate.
3.1 Mandibles, basolateral pit: (0) absent; (1) present (as in Fig. 45A, B).
3.2 Mandibles, dorsal pit: (0) absent; (1) present (as in Fig. 45C).
3.3 Mandibles, dorsolateral sulcus: (0) absent; (1) present (as in Fig. 45D).

4 Clypeus, anteroventral face: (0) absent; (1) present (avf, Fig. 39A-D).
11 Palpal formula: number of maxillary palps, number of labial palps.
15 Mesopleuron, division into anepisternum and katepisternum: (0) absent, (1) present.
22 Propodeal spiracle, shape: (0) slit-shaped, (1) round to oval.
29 Metatibia, apparent metatibial gland cuticular patch: (0) absent; (1) present (as in Fig. 41E).
30 Metatibia, number of well-developed spurs (i.e., number of spurs seen under a stereomicroscope).
33 Pretarsal claws, shape: (0) simple (as in Fig. 48B, C); (1) basal tooth (as in Figs 12B, 48D); (2) pre-apical tooth (as in Fig.
48E).
39 Petiolar poststernite, posterior spatulate projection: (0) absent; (1) present (as in Fig. 49A-G).
41 Abdominal poststernite III, prora: (1) present and conspicuous; (2) present (Fig. 49L), but indistinct in the profile of undissected specimens.

44 Abdominal pretergite IV, stridulitrum: (0) absent, (1) present.
46 Abdominal sternite VII (hypopygium), ventral face: (0) convex or flat; (1) distinctively concave longitudinally (Fig. 44).
47 Abdominal sternite VII (hypopygium), stout, spine-like setae on posterolateral portion: (0) absent, (1) present.
Superscripts:
${ }^{a}$ Absent in species from Borneo, Bali, Krakatau, and Sumatra (Yamane 2007).
${ }^{\mathrm{b}}$ Present in Mesoponera subiridescens.
c The palpal formula is 4,4 in all species in which the character was assessed save Mesoponera ambigua, whose formula is 3,3 . According to Fisher and Bolton (2016), Afrotropical and Malagasy species may present palpal formula 3,3; 4,3; 4,4.
${ }^{\text {d }}$ Present in Mayaponera conicula.
${ }^{e}$ Present in Mesoponera subiridescens and very shallowly impressed in some M. melanaria macra specimens and Indomalayan and Australasian species.
${ }^{\mathrm{f}}$ Oval to slit-shaped in Brachyponera sennaarensis among species examined; slit-shaped in images of B. sennaarensis subspecies on AntWeb.org. According to Schmidt and Shattuck (2014: 79), the character is round in all other species.
${ }^{\mathrm{g}}$ Slit-shaped in Mesoponera caffraria and subspecies, M. ingesta, and M. subiridescens.
${ }^{\text {h }}$ Among species examined $(N=6)$, the character is absent only in Brachyponera chinensis. It is visible in images of B. arcuata, B. atrata, and B. sennaarensis ruginota on AntWeb.org.
${ }^{\text {i }}$ Among species examined $(N=8)$, the character is present only in Mesoponera caffraria; it is visible in images of specimens identified as M. ingesta on AntWeb.org.
${ }^{\text {j }}$ Among species examined $(N=6)$, the stridulitrum is absent only in Brachyponera sennaarensis, but see Schmidt and Shattuck (2014: 79).
${ }^{\text {k }}$ Present in Mayaponera constricta.
${ }^{1}$ Among species examined $(N=8)$, the stridulitrum is only absent in Mesoponera ambigua. The character was not assessed in M. elisae rotundata; it is unclear in M. caffraria; but see Schmidt and Shattuck (2014: 109).
${ }^{m}$ Among species examined $(N=8)$, the character is only present in Brachyponera becculata.
${ }^{n}$ Among material examined, the character is absent in the Afrotropical species.
M. caffraria and subspecies, M. ingesta, and M. subiridescens); (5) the apicoposterior area of the metatibia presents an apparent metatibial gland cuticular patch (it is absent in Asphinctopone, Brachyponera chinensis, Mayaponera, and Mesoponera save M. caffraria, and likely its subspecies, and $M$. ingesta); (6) meso- and metatibiae bear two well-developed spurs (Asphinctopone only has one well-developed spur on each tibia); (7) pretarsal claws are simple (the claws present a preapical tooth in Ha gensia); (8) the posterior portion of the petiole sternite is not projected (it presents a spatulate projection that folds posteriad over the remaining sternite in Asphinctopone and Brachyponera); (9) the prora is conspicuous in profile (it is indistinct in profile of undissected specimens in Brachyponera, and some Mayaponera species); (10) the pretergite of abdominal segment IV presents a stridulitrum (the stridulitrum is absent in Asphinctopone, Brachyponera sennaarensis, Hagensia, Mayaponera save M. constricta, and Mesoponera ambigua); and (11) the posterolateral portion of the hypopygium lacks stout, spine-like setae (the setae are present in Mayaponera becculata, and Mesoponera save species occurring in the afrotropics). Moreover, Asphinctopone has an oblong, smooth, bright, yellowish cuticular patch on the dorsal face of its subtriangular mandibles (Hawkes 2010); the cuticular patch is absent from the triangular mandibles of Corrieopone.

Preliminary phylogenetic analysis based on molecular data inferred a sister relationship between Corrieopone and Asphinctopone (The Ants of the World Project, unpublished data); however, analysis of a more comprehensive dataset is still ongoing and upcoming results may challenge this hypothesis.

## Other castes and stages of development

Queen (Fig. 53A-F): Winged. Similar to the worker caste but for the slightly greater body length (HL: 1.65; TL: 8.48), larger compound eyes, presence of ocelli, differences in the mesosoma due to the presence of wings, and darker color. Clypeus as described in the definition of the genus. Parapsidal lines present on the mesoscutum (Fig. 53D). In profile, mesoscutum bulging; higher than the pronotum; slightly higher than the mesoscutellum (Fig. 53C). In profile, scutoscutellar sulcus indenting the dorsal outline of the mesoscutum and mesoscutellar-axillar complex. Mesoscutellum dome-shaped in profile; higher than the metanotum and the propodeum (Fig. 53C). Mesopleuron in profile divided into anepisternum and katepisternum. In profile, metapleuron divided


Figure 53. Corrieopone nouragues, paratype, dealated queen (CASENT0923157) A head in full-face view $\mathbf{B}$ body in profile $\mathbf{C}$ mesosoma in profile $\mathbf{D}$ body in dorsal view $\mathbf{E}$ metatibial apex and basal portion of the metabasitarsus in posterior view $\mathbf{F}$ medial area of abdominal segments III and IV in dorsal view. Images by M Esposito ( $\mathbf{A}, \mathbf{B}, \mathbf{D}, \mathbf{F}$ ) and FA Esteves ( $\mathbf{C}, \mathbf{E}$ ); available at AntWeb.org. Numbers: 1, mesoscutum; 2, indentation of the scutoscutellar sulcus; 3, mesoscutellum; 4, metanotum; 5, anepisternum; $\mathbf{6}$, katepisternum; $\mathbf{7}$, upper section of the metapleuron; $\mathbf{8}$, lower section of the metapleuron; $\mathbf{9}$, parapsidal line. Scale bars: $1 \mathrm{~mm}(\mathbf{A}, \mathbf{B}, \mathbf{D}) ; 0.2 \mathrm{~mm}(\mathbf{C}, \mathbf{E}, \mathbf{F})$.
into upper and lower sections; upper metapleuron separated from propodeum by sulcus; lower metapleuron indistinct from propodeum (Fig. 53C). Propodeal spiracle slitshaped. Legs as in the worker caste, including the presence of the metatibial gland cuticular patch on the apicoposterior face of the metatibia (Fig. 53E). Petiole and gaster
shaped as in the worker caste (Fig. 53B). Prora present; lip-shaped. Stridulitrum small, as in workers (Fig. 53F). Hypopygium as described in the definition of the genus.

## Male: Unknown.

Larva: Unknown.
Etymology. The genus name Corrieopone is feminine. It is a tribute to Dr. Corrie Saux Moreau, a myrmecologist, professor of Arthropod Biosystematics and Biodiversity at Cornell University, and director and curator of the Cornell University Insect Collection, in Ithaca, N.Y. A flagbearer for gender inclusion and diversity in our field, audacious (see Wilson 2013, chapter 13), upbeat, and the owner of a contagious laugh, Corrie inspires everybody with her trailblazing, glass ceiling- and stereotype-breaking approach to science. Moreover, the discovery of Corrieopone only occurred because, back in 2017, Corrie Moreau and Christophe Duplais spurred us to house Ant Course 2018 at Nouragues Research Station in French Guiana. The suffix "pone" is derived from the subfamily name Ponerinae.

Distribution and ecology. To date, Corrieopone is only known from the Natural Reserve of Nouragues in French Guiana (Figs 1, 2). Specimens were discovered in leaf litter at the base of an Astrocaryum palm in an area referred to as Petit Plateau, which is covered by oldgrowth lowland terra-firme equatorial evergreen forest (Fig. 2F; Poncy et al. 2001). The forest seemed undisturbed, and according to Charles-Dominique (2001), anthropogenic impact has been limited since settlements disappeared from the area in the eighteenth century.

## Corrieopone nouragues, sp. nov.

http://zoobank.org/A82C546A-D3B5-421C-BF00-BFF998F86F02
Figs 7, 8, 37-44, 52A, 53-57
Type locality. French Guiana: Cayenne, Nouragues Natural Reserve, Nouragues Research Station, Inselberg Station, Petit Plateau grid; $4.08354^{\circ} \mathrm{N}, 52.68368^{\circ} \mathrm{W}, \pm 5 \mathrm{~m}$; alt. 145 m ; old-growth lowland terra-firme equatorial evergreen forest; in leaf litter at the base of an Astrocaryum palm (Arecaceae).

Type specimens. Holotype: worker, pinned; "French Guiana: Rés. Nouragues, Inselberg Stn; $4.08354^{\circ} \mathrm{N},-52.68368^{\circ} \mathrm{W}, \pm 5 \mathrm{~m} ; 145 \mathrm{~m}$; rainforest; hand collection; 29 Aug. 2018; B.L. Fisher, M. Fichaux leg.; BLF41460"; CASC, CASENT0830464. Paratypes: French Guiana • 1 worker; same data as for holotype; JTLC, CASENT0645962 - 1 worker; same data as for holotype; MNHN, CASENT0872028 • 1 worker; same data as for holotype; MZSP, CASENT0872029 • 3 workers; same data as for holotype; CASC, CASENT0830465, CASENT0923158, CASENT0872031•1 dealated queen; same data as for holotype; CASC, CASENT0923157.

Worker description. Measurements ( $N=4$; holotype values within parentheses): HL: 1.54-1.57 (1.57); HW: 1.33-1.41 (1.41); SL: 1.59-1.67 (1.67); WL: 2.14-2.26 (2.26); TL: 6.54-7.10 (7.06); CI: 87-90 (89); SI: 119-120 (119).

Medium-sized, slender ants (TL $6.54-7.1 \mathrm{~mm}$ ) with characters as described for Corrieopone and the following:


Figure 54. Antenna of Corrieopone nouragues; paratypes, worker caste $\mathbf{A}$ right antenna in ventral view (CASENT0923158) B lateral face of the bulbus, bulbus neck, and basal portion of the main shaft of the scape; arrow indicates the notched lateral margin of the bulbus (CASENT0872031) C bulbus (arrowhead) and bulbus neck (arrow) in anterior view, with the lateral side facing the right of the image (CASENT0923158) D bulbus (indicated by arrowhead) and bulbus neck (indicated by arrow) in dorsal view with lateral side facing the right of the image (CASENT0923158). Images by FA Esteves; available at AntWeb.org. Scale bars: 0.1 mm .

Color: reddish brown; lateral surfaces of clypeus and appendages slightly lighter; apex of gaster yellowish (Fig. 37).

General vestiture: Head, mesosoma, petiole, and gaster vested with densely arranged, short, subdecumbent to suberect filiform setae, and with much sparser, longer, suberect to erect filiform setae; posterior region of abdominal segment VII with abundant longer setae (Figs 38A, B, 40A, 43A, C). Dorsal face of mandible with sparse, appressed, filiform setae, grading laterad to buoyant, flexuous, longer filiform setae (Fig. 38A, E). Legs dressed with densely arranged, short, subdecumbent to suberect filiform setae.

Sculpture: Head in dorsal view mostly punctate (Fig. 8D). Mandibles sparsely punctate, with shallow fossulae skirting the apical two-thirds of the dorsal face of the masticatory margin (Fig. 8C). Clypeus mostly smooth; region adjacent to median area weakly costulate. Pronotum weakly punctate dorsally, grading to costulate laterally (Fig. 8B). Mesonotum weakly punctate. Anepisternum mostly smooth to sparsely


Figure 55. Labrum and maxillolabial complex of Corrieopone nouragues, paratypes, worker caste $\mathbf{A}$ outer face of the labrum in ventrolateral view (CASENT0872031); arrow indicates transverse protrusion; arrowhead indicate median carina B outer face of the labrum in apical view (CASENT0923158); arrow indicates transverse protrusion; arrowhead indicate weak median carina $\mathbf{C}$ outer face of the maxillolabial complex (CASENT0923158); basal towards the bottom of the image; Roman numerals indicate the count of maxillary and labial palpomeres $\mathbf{D}$ outer face of the galea (CASENT0923158); galeal comb in orange; arrows indicate elliptic seta on the galeal crown; Roman numerals indicate maxillary palpomeres. Images by FA Esteves; available at AntWeb.org. Scale bars: 0.1 mm .
costulate (Fig. 40B). Katepisternum costulate dorsally, grading to obliquely strigulate ventrally (Fig. 40B). In profile, metapleuron obliquely strigulate (Fig. 40C). Propodeum strigulate-punctate dorsally; obliquely strigulate laterally (Fig. 40C); declivitous face strigulate laterally, mostly smooth medially (Fig. 8F). Petiolar tergite anterior face mostly smooth; lateral face mostly smooth, sparsely costulate, or confused rugose (Figs 8E, 43A); posterior face mostly smooth dorsally, grading to strigate ventrally (Figs 8F, 43A). Gaster weakly punctate; mostly smooth (Fig. 43C).

Head: Head slightly longer than broad (CI: 87-90). In dorsal view, posterior margin of the head slightly concave medially (Figs 37B, 38A). In profile, posteroventral curve of the head without a projecting flange (Figs 37C, 38B). Mandibles triangular, edentate, devoid of basolateral and dorsal pits or dorsolateral and dorsomasticatory sulci (Figs 37B, 38A, E). Basal three-fourths of the mandibular masticatory margin skirted ventrally by two rows of long, flexuous, helicoid setae, with a row of filiform setae present in between. Mandible with a row of filiform setae running obliquely


Figure 56. Leg of Corrieopone nouragues; paratypes, worker caste $\mathbf{A}$ calcar of strigil in anterior view (CASENT0872031) B probasitarsus in ventroanterior view (CASENT0923158) $\mathbf{C}$ probasitarsal notch in posterior view (CASENT0923158); arrow indicates longitudinal sulcus next to comb of strigil $\mathbf{D}$ ventral area of the mesotibia apical half and mesobasitarsus (CASENT0872031). Images by FA Esteves; available at AntWeb.org. Scale bars: 0.1 mm .
from the basolateral area of the ventral face to the apex of the lateral face; with setae increasing in length towards mandibular apex; apicalmost seta helicoid. Torular lobes closely approximated (Fig. 38A, F); torular lateral arches visible in dorsal view (Fig. 38 F ). In profile, torular lobes located at the dorsalmost part of a prominence formed by the clypeal median area and the frontal carinae (Fig. 38B). Lateral arch of the torulus projected dorsad, with a somewhat constricted rim (Fig. 38D, F). Twelve antennomeres (Fig. 54A); bulbus hemispherical, notched on the lateral margin (Fig. 54B); bulbus neck tubular (Fig. 54C, D). Antennal scape (antennomere I) surpassing the posterior margin of the head (SI: 119-120); antennomeres III and IV longer than all other preapical antennomeres save the scape; antennomere III almost as long as the apicalmost antennomere (Fig. 54A). Compound eyes present, located just anterior to the midlength of the head, surrounded by a sulcus; widest diameter of compound eyes: six or seven ommatidia (Fig. 38A, B, D). Ocelli absent. Labrum apically bilobed, with lobes broadly rounded apicolaterally (Fig. 39E); long, acute cleft at midpoint of apical margin. Middle of outer face of the labrum bulging transversely, with a short, median carina extending basad from the median cleft; the carina may be a weak, blunt pro-


Figure 57. Leg of Corrieopone nouragues; paratypes, worker caste $\mathbf{A}$ colliculate cuticular patch on the posterior face of the metatibial apex (CASENT0872031) B close-up of the colliculate cuticular patch on the metatibia (CASENT0872031) C posterior metatibial spur in posterior view (CASENT0872031) D metatibial spurs and metabasitarsus in anterior view (CASENT0923158) E basal portion of the metabasitarsus in anterior view (CASENT0923158); note the short, median carina (arrowhead) armed with a row of short, spine-like setae (colored in orange) $\mathbf{F}$ portion of the metabasitarsus anterior to its midlength, in anterior view (CASENT0923158). Images by FA Esteves; available at AntWeb.org. Scale bars: 0.05 $\mathrm{mm}(\mathbf{A}, \mathbf{B}) ; 0.2 \mathrm{~mm}(\mathbf{C}, \mathbf{D}) ; 0.1 \mathrm{~mm}(\mathbf{E}, \mathbf{F})$.
trusion or a somewhat developed ridge (Fig. 55A, B). Palpal formula 4,4 (Fig. 55C); basalmost maxillary palpomere short, bulbous. Outer face of maxillary stipes without a transverse sulcus (Fig. 55C). Galeal comb present (Fig. 55D). Galeal crown armed
with two or three elliptic setae (Fig. 55D); inner face without a translucid comb of setae (Fig. 55C, D). Premental shield without a transverse sulcus (Fig. 55C).

Mesosoma: In dorsal view, mesonotum round, wider than the dorsal face of the propodeum but narrower than the pronotum (Fig. 37A); metanotal sulcus mostly smooth, conspicuous. In profile, mesonotum dome-shaped; notopleural suture separating mesonotum from mesopleuron; promesonotum higher than propodeum; metanotal sulcus deeply impressed (Figs 37C, 40A). In profile, mesopleuron divided into anepisternum and katepisternum (Fig. 40A, B). Mesometapleural suture present (Fig. 40A-C). Metathoracic spiracle concealed by a spiracular lobe (Fig. 40A, B). Posterolateral surface of the metapleuron devoid of a longitudinal flange (Fig. 40A, C). Orifice of the metapleural gland round, opening posterolaterad, with ventral margin atop a carina located on the posterolateral margin of the metapleuron (= metapleural carina; Fig. 40C, D). Propodeal dorsum devoid of a median longitudinal impression, propodeal spines absent (Fig. 40A), propodeal posterolateral margin not carinate (Fig. 40A, C). In profile, propodeal lobe rounded, not surpassing posteriorly the dorsoposterior-most point of the rim of the propodeal foramen (Fig. 40C); propodeal spiracle slit-shaped (Fig. 40A, C). Mesosternal process bidentate; metasternal process bilobate, long (Fig. 40F). Metacoxal cavities tightly encircled by unfused annuli (Fig. 40E).

Legs: Apico-anterior portion of protibia with a brush of spatulate-costate setae, next to calcar of strigil (Fig. 41A); apicoposterior portion of protibia devoid of stout, spine-like setae (Fig. 41B). Basal third of the ventral margin of the calcar lamellate; lamella glabrous, devoid of any notch (Fig. 41B). From base to apex, microtrichia on anterior face of calcar grades from tubiform to spatulate (Fig. 56A); posterior face of calcar with lanceolate microtrichia (Fig. 41B). Anterior and ventral faces of probasitarsus densely vested with spatulate-costate setae (Fig. 41A), except for the area immediately anterior to the comb of strigil, which bears shorter, spatulate-bicuspid setae (Fig. 56B). Comb of strigil skirted posteriorly by a sulcus (Fig. 56C). Row of stout, spine-like setae on posterior face of probasitarsal notch, parallel to comb of strigil (Fig. 56C). Counting from base to apex: anterior face of the second tarsomere of foreleg with spatulatecostate setae; ventral faces of second, third, and fourth tarsomeres with stout, spine-like setae (Fig. 42C); fourth tarsomere cylindrical. Mesotibia with two spurs (Fig. 41C); anterior spur simple (Fig. 56D); posterior spur serrate, with lanceolate microtrichia on dorsal face (Fig. 41D). Mesobasitarsus devoid of any longitudinal sulcus (Fig. 56D); two rows of spine-like setae present along apical half of ventral face of mesobasitarsus and along entire ventral face of second, third, and fourth tarsomeres (Fig. 42D). Metacoxal dorsum devoid of any spine-like projection. Posterior portion of metatibial apex with a glabrous, yellowish, oblong, weakly colliculate cuticular patch (Figs 41E, 57A, B); cuticular patch with densely arranged, minute pores (visible under magnifications above 4700×; Fig. 57B); pores round to slit-shaped ( $\sim 0.29$ um length; visible at $8500 \times$ magnification); cuticle surrounding oblong cuticular patch with sparser, longer slit-shaped pores ( $\sim 0.45 \mathrm{um}$ length). Ventral face of metatibial apex with two spurs (Fig. 57D); anterior spur simple and glabrous. Metatibial posterior spur pectinate;
with lanceolate microtrichia on anterior face; with mostly glabrous posterior face, except for lanceolate microtrichia on dorsal surface (Fig. 57C, D). Metabasitarsus devoid of any longitudinal sulcus (Fig. 57D). Midline of the ventrobasal portion of metabasitarsus with a short, longitudinal carina armed with a row of short, spine-like setae (Fig. 57E); the carina is followed apically by a longitudinal row of spatulate setae that extends to the midlength of the segment (Fig. 57F); ventral face of metabasitarsus with the apical two-thirds bearing two rows of spine-like setae that skirt the midline row of spatulate setae. Dorsal faces of the mid- and hindlegs devoid of stout, spine-like setae (Fig. 42A, B). Ventral faces of the second, third, and fourth tarsomeres with two rows of spine-like setae that skirt the midline of each segment (Fig. 42E). Simple pretarsal claws on fore-, mid-, and hindlegs (Fig. 42C-F). Arolium indistinct on pro-, meso-, and metapretarsi (Fig. 42C-F).

Petiole: Sessile; surmounted by a high, unarmed, conic scale-like node that is narrow in profile and in dorsal view (Figs 37A, C, 43A). Anteroventral portion of the tergite with a lateral carina (Fig. 43A); spiracle orifice directed dorsolaterally; posterior face of the tergite with the ventral portion strigate; laterotergite delineated by a longitudinal suture (Fig. 43B). Subpetiolar process keel-like, with a round anteroventral projection (Fig. 43A), fenestra absent. Anterior disc of sternite with a round proprioceptor zone (Fig. 43B). Sternite without a posterior spatulate projection (Fig. 43B), articulation with the helcium visible in ventral view.

Gaster: Helcium infra-axial: positioned ventrad to the midheight of the anterior face of abdominal segment III (Fig. 43C). Prora well-developed on the anterior face of the sternite of abdominal segment III, lip-shaped (Fig. 43C, D). Abdominal segment IV tubular: tergite and sternite with similar lengths, tergite not arched (Fig. 43C). Presclerites of the abdominal segment IV forming an even surface with postsclerites (Fig. 43C), no girdling constriction. Small stridulitrum present on the pretergite of the abdominal segment IV (Fig. 43E, F). Pygidium not armed with spine-like setae, no acute cuticular projections, dorsum convex. Hypopygium as described for the genus (Fig. 44).

Queen description. Measurements $(N=1)$ : HL: 1.65; HW: 1.54; SL: 1.74; WL: 2.51; TL: 8.48; CI: 93; SI: 113.

Dealate. Color dark brown; clypeus and appendages lighter, reddish; apex of gaster yellowish (Fig. 53A-D). Similar to the worker caste but for the darker body color and slightly larger body length (TL 8.48), larger compound eyes, presence of ocelli, and differences in the mesosoma due to the presence of wings (Fig. 53). As described for the genus.

Etymology. The specific epithet nouragues is a non-Latin noun in apposition. It honors the Nouragues (or Norak) people, a Tupi Amerindian population who once inhabited the area (Grenand 1982), and the Nouragues Natural Reserve and Research Station.

## Acknowledgments

We thank Melanie Fichaux for finding the first specimen and calling BLF to help collect more specimens. We thank Jerôme Orivel and Frederic Petitclerc for their assis-
tance in organizing the course in French Guiana. Financial support for this study was provided by an Investissement d'Avenir grant of the Agence Nationale de la Recherche (CEBA, ANR-10-LABX-25-01). In addition, the research was supported in part by NSF DEB-1932467. We are grateful to the Nouragues Natural Reserve for allowing sampling in the protected area and the Nouragues Research Station and CNRS Guyane for logistical support. A special thanks to the Station team who made the course a success: Bran Leplat, Philippe Gaucher. In addition, we thank the Ant Course 2018 participants: Adam Khalife, Alexandre Ferreira, Cédric Chény, Charlotte Francoeur, Christian Peeters, Cody Cardenas, Frederic Petitclerc, Iryna Ivasyk, Jack Longino, Jason Williams, Jill Oberski, Julie Miller, Lily Leahy, Maria Fernanda Brito de Almeida, Mark Wong, Mélanie Fichaux, Naoto Idogawa, Pamela Eugenia Pairo, Peter Flynn, Phil Ward, Philipp Hoenle, Rachel Wells, and Philippe Gaucher.

We express our gratitude to Bernard Landry (MHNG, Genève, Switzerland) for examining one of the syntypes of Leptogenys butteli deposited in the MHNG. Many, many thanks to Roberto A Keller (Museu Nacional de Hist. Natural e da Ciência, Lisboa, Portugal) for making freely available his innumerous SEM images on AntWeb.org; this study greatly benefited from the exceptional information those images contain. We extend our gratitude to our two reviewers and the editorial team of ZooKeys, whose graceful comments, constructive criticism, and suggestions greatly improved this manuscript.

Finally, we cannot thank enough the extraordinary Michele Esposito (CASC, San Francisco, USA), whose generosity, expertise in photography, design, databasing, and overall support in the lab have contributed to this study in immeasurable ways - thank you!

## References

AntCat (2021) An online catalog of the ants of the world. http://antcat.org. [Accessed: March 01, 2021]
Arimoto K, Yamane S (2018) Taxonomy of the Leptogenys chalybaea species group (Hymenoptera, Formicidae, Ponerinae) from Southeast Asia. Asian Myrmecology 10: e010008. https://doi.org/10.20362/am. 010008
Baroni Urbani C (1975b) Contributo alla conoscenza dei generi Belonopelta Mayr e Leiopelta gen. n. (Hymenoptera: Formicidae). Mitteilungen der Schweizerischen Entomologischen Gesellschaft 48: 295-310. https://doi.org/10.5281/zenodo. 26787
Bernard F (1967) Faune de l'Europe et du Bassin Méditerranéen 3. Les fourmis (Hymenoptera Formicidae) d'Europe occidentale et septentrionale. Masson, Paris, 411 pp .
Billen J (2009) Occurrence and structural organization of the exocrine glands in the legs of ants. Arthropod Structure \& Development 38: 2-15. https://doi.org/10.1016/j.asd.2008.08.002
Bolton B (1975) A revision of the ant genus Leptogenys Roger (Hymenoptera: Formicidae) in the Ethiopian region with a review of the Malagasy species. Bulletin of the British Museum (Natural History). Entomology 31: 235-305. https://doi.org/10.5962/bhl.part. 29487

Bolton B, Brown Jr WL (2002) Loboponera gen. n. and a review of the Afrotropical Plectroctena genus group (Hymenoptera: Formicidae). Bulletin of the Natural History Museum. Entomology Series 71: 1-18. https://doi.org/10.1017/S0968045402000019
Bolton B (2003) Synopsis and classification of Formicidae. Memoirs of the American Entomological Institute 71. American Entomological Institute, Gainesville, 370pp.
Bolton B, Fisher BL (2008a) The Afrotropical ponerine ant genus Phrynoponera Wheeler (Hymenoptera: Formicidae). Zootaxa 1892: 35-52. https://doi.org/10.11646/zootaxa.1892.1.3
Bolton B, Fisher BL (2008b) Afrotropical ants of the ponerine genera Centromyrmex Mayr, Promyopias Santschi gen. rev. and Feroponera gen. n., with a revised key to genera of African Ponerinae (Hymenoptera: Formicidae). Zootaxa 1929: 1-37. https://doi.org/10.11646/zootaxa.1929.1.1
Boudinot BE, Perrichot V, Chaul JCM (2020) Camelosphecia gen. nov., lost ant-wasp intermediates from the mid-Cretaceous (Hymenoptera, Formicoidea). ZooKeys 1005: 21-55. https://doi.org/10.3897/zookeys.1005.57629
Boudinot BE, Moosdorf OTD, Beutel RG, Richter A (2021) Anatomy and evolution of the head of Dorylus helvolus (Formicidae: Dorylinae): Patterns of sex- and caste-limited traits in the sausagefly and the driver ant. Journal of Morphology 282: 1616-1658. https://doi. org/10.1002/jmor. 21410
Brown Jr WL (1963) Characters and synonymies among the genera of ants. Part III. Some members of the tribe Ponerini (Ponerinae, Formicidae). Breviora 190: 1-10. https://doi. org/10.5281/zenodo. 26976
Charles-Dominique P (2001) The Field Station. In: Bongers F, Charles-Dominique P, Forget PM, Théry M (Eds) Nouragues: Dynamics and plant-animal interactions in a Neotropical rainforest. Monographiae Biologicae, vol. 80. Springer, Dordrecht, 1-8. https://doi. org/10.1007/978-94-015-9821-7
Collingwood CA, Agosti D (1996) Formicidae (Insecta: Hymenoptera) of Saudi Arabia (part 2). Fauna of Saudi Arabia 15: 300-385. https://ia802304.us.archive.org/23/items/ ants_08420/8420_text.pdf
D’Esquivel MS, Jahyny BJB, Oliveira ML, Lacau LSR, Delebie JHC, Lacau S (2017) Thaumatomyrmex fraxini sp. nov. (Hymenoptera: Formicidae), a new ant species from the Brazilian Atlantic Forest. Sociobiology 64(2): 159-165. https://doi.org/10.13102/sociobiology.v64i2.1615
Dinerstein E, Olson DP, Joshi A, Vynne C, Burgess N, Wikramanayake E, Hahn N, Palminteri S, Hedao P, Noss R, Hansen M, Locke H, Ellis EC, Jones BS, Barber CV, Hayes R, Kormos C, Martin V, Crist E, Sechrest W, Price L, Baillie J, Weeden D, Suckling KF, Davis C, Sizer N, Moore R, Thau D, Birch T, Potapov P, Turubanova S, Tyukavina A, Souza ND, Pintea L, Brito JC, Llewellyn O, Miller AG, Patzelt A, Ghazanfar S, Timberlake J, Klöser H, Shennan-Farpón Y, Kindt R, Lillesø JP, Breugel PV, Graudal L, Voge M, Al-Shammari KF, Saleem M (2017) An ecoregion-based approach to protecting half the terrestrial realm. Bioscience 67: 534-545. https://doi.org/10.1093/biosci/bix014
Fernandes IO, Oliveira ML de, Delabie JHC (2014) Description of two new species in the Neotropical Pachycondyla foetida complex (Hymenoptera: Formicidae: Ponerinae) and taxonomic notes on the genus. Myrmecological News 19: 133-163. https://www.myrmecologicalnews.org/cms/index.php?option=com_content\&view=category\&id=589\&Itemid=363

Fernandes IO, Delabie JHC (2019) A new species of Cryptopone Emery (Hymenoptera: Formicidae: Ponerinae) from Brazil with observations on the genus and a key for new word species. Sociobiology 66: 408-441. https://doi.org/10.13102/sociobiology.v66i3.4354
Fisher BL (2006) Boloponera vicans gen.n. and sp.n. and two new species of the Plectroctena genus group (Hymenoptera: Formicidae). Myrmecologische Nachrichten 8: 111-118. http://antbase.org/ants/publications/21106/21106.pdf
Fisher BL, Guenard BS, Robson S (2015) Borneo, fANTastique! Asian Myrmecology 7: 171174. https://doi.org/10.20362/am. 007018

Fisher BL, Bolton B (2016) Ants of Africa and Madagascar: A guide to the genera. University of California Press, Berkeley, 514 pp. https://doi.org/10.1525/9780520962996
Forel A (1910) Ameisen aus der Kolonie Erythräa. Gesammelt von Prof. Dr. K. Escherich (nebst einigen in West-Abessinien von Herrn A. Ilg gesammelten Ameisen). Zoologische Jahrbücher. Abteilung für Systematik, Geographie und Biologie der Tiere 29: 243-274. https://www.zobodat.at/pdf/Zoologische-Jahrbuecher-Syst_29_0243-0274.pdf
Forel A (1913) Wissenschaftliche Ergebnisse einer Forschungsreise nach Ostindien ausgeführt im Auftrage der Kgl. Preuss. Akademie der Wissenschaften zu Berlin von H. v. Buttel-Reepen. II. Ameisen aus Sumatra, Java, Malacca und Ceylon. Gesammelt von Herrn Prof. Dr. v. Buttel-Reepen in den Jahren 1911-1912. Zoologische Jahrbücher. Abteilung für Systematik, Geographie und Biologie der Tiere 36: 1-148. https://biostor.org/reference/181439
Gibson GAP (1985) Some pro- and mesothoracic structures important for phylogenetic analysis of Hymenoptera, with a review of the terms used for the structures. Canadian Entomologist 117: 1395-1443. https://doi.org/10.4039/Ent1171395-11
Gibson GAP, Read JD, Fairchild R (1998) Chalcid wasps (Chalcidoidea): Illustrated glossary of positional and morphological terms. http://www.canacoll.org/Hym/Staff/Gibson/apss/ chglintr.htm
Gibson JC, Larabee FJ, Touchard A, Orivel J, Suarez AV (2018) Mandible strike kinematics of the trap-jaw ant genus Anochetus Mayr (Hymenoptera: Formicidae). Journal of Zoology 306: 119-128. https://doi.org/10.1111/jzo. 12580
Greenberg JA, Mattiuzzi M (2020) gdalUtils: Wrappers for the Geospatial Data Abstraction Library (GDAL) Utilities. R package version 2.0.3.2. https://CRAN.R-project.org/package=gdalUtils
Grenand P (1982) Ainsi parlaient nos ancêtres. Essai d'ethnohistoire Wayapi. ORSTOM, Paris, 408 pp .
Grimaldi M, Riéra B (2001) Geography and Climate. In: Bongers F, Charles-Dominique P, Forget PM, Théry M (Eds) Nouragues: Dynamics and plant-animal interactions in a Neotropical rainforest. Monographiae Biologicae, vol. 80. Springer, Dordrecht, 9-18. https:// doi.org/10.1007/978-94-015-9821-7
Hansen MC, Potapov PV, Moore R, Hancher M, Turubanova SA, Tyukavina A, Thau D, Stehman SV, Goetz SJ, Loveland TR, Kommareddy A, Egorov A, Chini L, Justice CO, Townshend JRG (2013) High-Resolution Global Maps of $21^{\text {st }}$-Century Forest Cover Change. Science 342 (6160): 850-853. https://doi.org/10.1126/science. 1244693
Harris RA (1979) A glossary of surface sculpturing. California Department of Food and Agriculture, Bureau of Entomology 28: 1-31. https://doi.org/10.5281/zenodo. 26215
Hawkes PG (2010) A new species of Asphinctopone (Hymenoptera: Formicidae: Ponerinae) from Tanzania. Zootaxa 2480: 27-36. https://doi.org/10.11646/zootaxa.2480.1.2

Hijmans R (2021) raster: Geographic Data Analysis and Modeling. R package version 3.4-13. https://CRAN.R-project.org/package=raster
Hölldobler B, Obermayer M, Peeters C (1996) Comparative study of the metatibial gland in ants (Hymenoptera, Formicidae). Zoomorphology 116: 157-167. https://doi. org/10.1007/BF02527156
Keller RA (2011) A phylogenetic analysis of ant morphology (Hymenoptera: Formicidae) with special reference to the poneromorph subfamilies. Bulletin of the American Museum of Natural History 355: 1-90. https://doi.org/10.1206/355.1
Kempf WW (1960) Miscellaneous studies on Neotropical ants (Hymenoptera, Formicidae). Studia Entomologica 3: 417-466. https://doi.org/10.5281/zenodo. 26016
Kempf WW (1961a) As formigas do gênero Pachycondyla Fr. Smith no Brasil (Hymenoptera: Formicidae). Revista Brasileira de Entomologia 10: 189-204. https://doi.org/10.5281/zenodo. 26019
Kempf WW (1961b) A survey of the ants of the soil fauna in Surinam (Hymenoptera: Formicidae). Studia Entomologica 4: 481-524. https://doi.org/10.5281/zenodo. 26018
Kempf WW (1964) Miscellaneous studies on Neotropical ants III (Hymenoptera: Formicidae). Studia Entomologica 7: 45-71. https://doi.org/10.5281/zenodo. 26030
Kempf WW (1975) A revision of the Neotropical ponerine ant genus Thaumatomyrmex Mayr (Hymenoptera: Formicidae). Studia Entomologica 18: 95-126. https://doi.org/10.5281/ zenodo. 26052
Kronauer DJC (2004) Trophic parasitism of a wasp (Hymenoptera: Ampulicidae: Ampulex sp.) on the ant Ectatomma ruidum (Roger, 1860) (Hymenoptera: Formicidae). Myrmecologische Nachrichten 6: 77-78. https://myrmecologicalnews.org/cms/index.php?option=com_ content\&view=category\&id=153\&Itemid=350
LaPolla J, Cover S, Mueller U (2002) Natural history of the mealybug-tending ant, Acropyga epedana, with descriptions of the male and queen castes. Transactions of the American Entomological Society 128: 367-376. https://antwiki.org/wiki/images/d/d2/Cover_2002.pdf
Larabee FJ, Gronenberg W, Suarez AV (2017) Performance, morphology and control of poweramplified mandibles in the trap-jaw ant Myrmoteras (Hymenoptera: Formicidae). Journal of Experimental Biology 220: 3062-3071. https://doi.org/10.1242/jeb. 156513
Larabee FJ, Smith AA, Suarez AV (2018) Snap-jaw morphology is specialized for high-speed power amplification in the Dracula ant, Mystrium camillae. Royal Society Open Science 5: e181447. http://dx.doi.org/10.1098/rsos. 181447
Lattke JE, Delsinne T (2016) Revisionary and natural history notes on some species of the genus Gnamptogenys Roger, 1863 (Hymenoptera: Formicidae). Myrmecological News 22: 141-147. https://myrmecologicalnews.org/cms/index.php?option=com_content\&view=c ategory\&id=635\&Itemid=366
Lehner B, Döll P (2004) Development and validation of a global database of lakes, reservoirs and wetlands. Journal of Hydrology 296: 1-22. https://doi.org/10.1016/j.jhydrol.2004.03.028
Longino JT, Branstetter MG (2020) Phylogenomic species delimitation, taxonomy, and 'bird guide' identification of the Neotropical ant genus Rasopone (Hymenoptera: Formicidae). Insect Systematics and Diversity 4: 1-33. https://doi.org/10.1093/isd/ixaa004

Mackay WP, Mackay E (2010) The systematics and biology of the New World ants of the genus Pachycondyla (Hymenoptera: Formicidae). Edwin Mellen Press, Lewiston (New York): 642 pp. http://dx.doi.org/10.13140/2.1.4271.8726
MarklH(1973)Theevolutionofstridulatorycommunicationinants.In:ButlerCG,HowsePE(Eds)Proceedings oftheInternational Union for the Study ofSocial Insects. VIIInternational Congress, London, September 1973. University of Southampton, Southampton, 258-265. https://cataglyphis. fr/Actes-SF-UIEIS/IUSSI-Londres-1973/VII-Congres-IUSSI-Londres-Septembre-1973.htm
Mayr G (1901) Südafrikanische Formiciden, gesammelt von Dr. Hans Brauns. Annalen des Kaiserlich-Königlichen Naturhistorischen Museums in Wien 16: 1-30. https://doi. org/10.5281/zenodo. 25873
NASA JPL (2013) NASA Shuttle Radar Topography Mission Water Body Data Shapefiles \& Raster Files. Distributed by NASA EOSDIS Land Processes DAAC. https://doi. org/10.5067/MEaSUREs/SRTM/SRTMSWBD. 003
NASA JPL (2020) NASADEM Merged DEM Global 1 arc second V001, distributed by NASA EOSDIS Land Processes DAAC. https://doi.org/10.5067/MEaSUREs/NASADEM/NASADEM_HGT. 001
Ouellet Dallaire C, Lehner B, Sayre R, Thieme M (2019) A multidisciplinary framework to derive global river reach classifications at high spatial resolution. Environmental Research Letters 14(2): e024003. https://doi.org/10.1088/1748-9326/aad8e9
Padgham M, Rudis B, Lovelace R, Salmon M (2017) osmdata. Journal of Open Source Software 2(14): e305. https://doi.org/10.21105/joss. 00305
Pebesma E (2018) Simple features for R: Standardized support for spatial vector data. The R Journal 10: 439-446. https://doi.org/10.32614/RJ-2018-009
Peeters C (2017) Independent colony foundation in Paraponera clavata (Hymenoptera: Formicidae): First workers lay trophic eggs to feed queen's larvae. Sociobiology 64: 417-422. https://doi.org/10.13102/sociobiology.v64i4.2092
Peeters C, Fuminori I, Wiwatwitaya D, Keller RA, Hashim R, Molet M (2017) Striking polymorphism among infertile helpers in the arboreal ant Gesomyrmex. Asian Myrmecology 9: e009015. http://dx.doi.org/10.20362/am. 009015
Poncy O, Sabatier D, Prevost MF, Hardy I (2001) The lowland high forest: structure and tree species diversity. In: Bongers F, Charles-Dominique P, Forget PM, Théry M (Eds) Nouragues: Dynamics and plant-animal interactions in a Neotropical rainforest. Monographiae Biologicae, vol. 80. Springer, Dordrecht, 31-46. https://doi.org/10.1007/978-94-015-9821-7
R Core Team (2021) R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL https://www.R-project.org/
Rakotonirina JC, Fisher BL (2013) Revision of the Pachycondyla sikorae species-group (Hymenoptera: Formicidae)inMadagascar.Zootaxa3683:447-485.https://doi.org/10.11646/zootaxa.3683.4.8
Richards OW (1977) Hymenoptera. Introduction and key to families. Second edition. Handbooks for the Identification of British Insects, Vol. 6, Part 1. Royal Entomological Society of London, London, 100 pp. https://www.royensoc.co.uk/sites/default/files/Vol06_Part01_ed2.pdf
RStudio Team (2021) RStudio: Integrated development environment for R. RStudio, PBC, Boston, MA. http://www.rstudio.com
Schmidt, CA (2013) Molecular phylogenetics of ponerine ants (Hymenoptera: Formicidae: Ponerinae). Zootaxa 3647: 201-250. https://doi.org/10.11646/zootaxa.3647.2.1

Schmidt CA, Shattuck SO (2014) The higher classification of the ant subfamily Ponerinae (Hymenoptera: Formicidae), with a review of ponerine ecology and behavior. Zootaxa 3817: 1-242. https://doi.org/10.11646/zootaxa.3817.1.1
Snodgrass RE (1910) The thorax of the Hymenoptera. Proceedings of the United States National Museum 39: 37-91. https://doi.org/10.5479/si.00963801.39-1774.37
South A (2011) rworldmap: A new R package for mapping global data. The R Journal 3(1): 35-43. https://doi.org/10.32614/RJ-2011-006
South A (2012) rworldxtra: Country boundaries at high resolution. R package version 1.01. https://CRAN.R-project.org/package=rworldxtra
Tennekes M (2018). tmap: Thematic maps in R. Journal of Statistical Software 84(6): 1-39. https://doi.org/10.18637/jss.v084.i06
Ward PS (1990) The ant subfamily Pseudomyrmecinae (Hymenoptera: Formicidae): generic revision and relationship to other formicids. Systematic Entomology 15: 449-489. https:// doi.org/10.1111/j.1365-3113.1990.tb00077.x
Wickham H, François R, Henry L, Müller K (2021). dplyr: A grammar of data manipulation. R package version 1.0.7. https://doi.org/10.1007/978-1-4842-6876-6_1
Willey RB, Brown Jr WL (1983) New species of the ant genus Myopias (Hymenoptera: Formicidae: Ponerinae). Psyche 90: 249-285. https://doi.org/10.5281/zenodo. 25285
Wilson EO (2013) Letters to a young scientist. Liveright, New York, 244 pp.
Yamane S (2007) Pachycondyla nigrita and related species in Southeast Asia. In: Snelling RR, Fisher BL, Ward PS (Eds) Advances in ant systematics (Hymenoptera: Formicidae): homage to E. O. Wilson - 50 years of contributions. Memoirs of the American Entomological Institute, 80. American Entomological Institute, Gainesville, 650-663.
Yoder MJ, Mikó I, Seltmann KC, Bertone MA, Deans AR (2010) A Gross Anatomy Ontology for Hymenoptera. PLoS ONE 5: e15991. https://doi.org/10.1371/journal.pone. 0015991

## Supplementary material I

## Table S1

Authors: Flavia A. Esteves, Brian L. Fisher
Data type: Specimen data.
Explanation note: Here, we provide specimen data of material examined (unique identifiers, collection and specimen information, depositories, and links to each specimen webpage on AntWeb.org). This material was used to contrast the morphology of Corrieopone nouragues gen. nov. with 129 species or subspecies representing all 47 currently valid extant Ponerinae genera (see also Tables S2, S3).
Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.
Link: https://doi.org/10.3897/zookeys.1074.75551.suppl1

## Supplementary material 2

## Table S2

Authors: Flavia A. Esteves, Brian L. Fisher
Data type: Morphological.
Explanation note: Here, we list and define morphological characters used to compare the morphology of Corrieopone nouragues gen. nov., sp. nov. with other Ponerinae genera. When producing this list and the associated character matrix (Table S3), our goals were to document and organize our observations. It only contains characters which we could unambiguously discretize. We share it to disclose our methodology and foster validation, replication, and reinterpretation of our results.
Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.
Link: https://doi.org/10.3897/zookeys.1074.75551.suppl2

## Supplementary material 3

## Table S3

Authors: Flavia A. Esteves, Brian L. Fisher
Data type: Morphological.
Explanation note: Table S3 documents states of characters we evaluated across ponerine genera (see character statements in Table S2). When producing this matrix, our goals were to document and organize our observations. It only contains characters which we could unambiguously discretize. We share it to disclose our methodology and foster validation, replication, and reinterpretation of our results. Character numbers correspond to those in Corrieopone genus diagnosis.
Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.
Link: https://doi.org/10.3897/zookeys.1074.75551.suppl3

## Supplementary material 4

Table S4
Authors: Flavia A. Esteves, Brian L. Fisher
Data type: Morphometric.
Explanation note: All measurements were taken at $80 \times$ power with a Leica MZ125 microscope using an orthogonal pair of micrometers, recorded to the nearest 0.001 mm , and rounded to two decimal places, as following. Head length (HL): maximum longitudinal length of the head, measured from the anteriormost portion of the projecting clypeus to the midpoint of an imaginary line traced across the posterior margin of the head. Head width (HW): maximum width of head, excluding eyes. Scape length (SL): maximum chord length of the main shaft of the antennal scape, excluding basal bulbus and bulbus neck. Weber's length (WL): with the mesosoma in profile, the diagonal length from the posteroventral corner of the propodeum to the farthest point on the anterior face of the pronotum, excluding the neck. Total length (TL): sum of HL + WL + length of segments A2 to A7. A2 to A 7 is measured as follows: maximum length of the petiole in profile (PL) +A 3 to A7, or gaster length (GL). Cephalic index (CI): HW/HL $\times 100$. Scape index (SI): SL/HW $\times 100$.
Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.
Link: https://doi.org/10.3897/zookeys.1074.75551.suppl4

